



University
of Glasgow

<https://theses.gla.ac.uk/>

Theses Digitisation:

<https://www.gla.ac.uk/myglasgow/research/enlighten/theses/digitisation/>

This is a digitised version of the original print thesis.

Copyright and moral rights for this work are retained by the author

A copy can be downloaded for personal non-commercial research or study,
without prior permission or charge

This work cannot be reproduced or quoted extensively from without first
obtaining permission in writing from the author

The content must not be changed in any way or sold commercially in any
format or medium without the formal permission of the author

When referring to this work, full bibliographic details including the author,
title, awarding institution and date of the thesis must be given

Enlighten: Theses

<https://theses.gla.ac.uk/>
research-enlighten@glasgow.ac.uk

The expression of prostanoid receptor genes in uterine and fetal tissues.

Studies in the maternal and fetal baboon and the fetal and neonatal lamb

Gordon C S Smith, B.Sc., M.B. Ch.B., M.D., M.R.C.O.G.

Submitted for the degree of PhD to the University of Glasgow.

Research carried out in:

**The Laboratory for Pregnancy and Newborn Research,
Department of Physiology,
College of Veterinary Medicine,
Cornell University,
Ithaca,
NY 14853,
United States of America.**

Supervisors:

Prof P W Nathanielsz, M.D., Ph.D., D.Sc., F.R.C.O.G.

(James Law Professor of Reproductive Physiology, Cornell University)

Prof Iain T Cameron, M.D., F.R.C.O.G.

(Regius Professor of Midwifery, University of Glasgow)

September 2000.

ProQuest Number: 10644214

All rights reserved

INFORMATION TO ALL USERS

The quality of this reproduction is dependent upon the quality of the copy submitted.

In the unlikely event that the author did not send a complete manuscript and there are missing pages, these will be noted. Also, if material had to be removed, a note will indicate the deletion.



ProQuest 10644214

Published by ProQuest LLC (2017). Copyright of the Dissertation is held by the Author.

All rights reserved.

This work is protected against unauthorized copying under Title 17, United States Code
Microform Edition © ProQuest LLC.

ProQuest LLC.
789 East Eisenhower Parkway
P.O. Box 1346
Ann Arbor, MI 48106 – 1346



Thesis 12144
copy 2

Abstract.

1. The aim of this project was to determine whether advancing gestational age and parturition were associated with alteration in the relative level of expression of genes encoding prostanoid receptors in key uterine and fetal tissues. I also sought to determine whether advancing gestational age and parturition were associated with alteration in the expression of genes encoding lipoxygenase (LOX) enzymes in key intra-uterine tissues.
2. Caesarean hysterectomy was performed on 15 pregnant baboons in the last third of pregnancy. Samples of myometrium (from multiple uterine sites), cervix, decidua and chorion were obtained. In addition, the ductus arteriosus was obtained from nine fetal baboons, 28 fetal lambs and 4 neonatal lambs. Expression of genes was studied using Northern blot analysis and in situ hybridization. Expression of genes was quantified by Northern analysis as a ratio of the signal for the gene of interest to each of three housekeeping genes (glyceraldehyde-3-phosphate dehydrogenase [GAPDH], beta-actin and cyclophilin). Statistical comparison of the effects of gestational age and labour was performed using linear regression, Student's t-test, repeated measures analysis of variance and analysis of covariance, as appropriate.
- 3 Initial studies of animals not in labour using cDNA probes demonstrated transcripts of similar size to the human genes for prostanoid EP₂, EP₃, EP₄, and FP receptor mRNA using Northern blot in myometrium. Myometrium from the lower uterine segment (LUS) had greater expression of EP₂ receptor mRNA and less expression of EP₃ mRNA compared with the fundus and corpus. However, similar levels of EP₄ and FP receptor

mRNA were observed comparing the fundus and LUS. Expression of EP₂, EP₃ and EP₄ receptor mRNA were also detected in cervix, decidua and chorion. EP₂ mRNA was most abundant in cervix, EP₃ was most abundant in myometrium and EP₄ mRNA was most abundant in decidua. The variation in myometrial expression of genes encoding EP receptor sub-types paralleled the contractile responses of paired samples (reported elsewhere).

4 When expression of prostanoid receptor genes was studied in myometrium obtained from animals both in labour and not in labour and the techniques employed were optimized (principally the use of riboprobes), transcripts of similar size to the human genes were detected for prostanoid EP₁, EP₂, EP₃, EP₄, IP, FP and TP receptor mRNA using Northern blot. There were no gestational age related changes in expression of these genes. Expression of EP₁, EP₃ and IP receptor mRNA was significantly higher in myometrium from the fundus (compared with lower segment) whereas EP₂ gene expression was significantly lower in the fundus. Labor was associated with a reduction in the regional variation of both EP₂ and IP receptor gene expression, but not EP₁ and EP₃ expression. Labor was also associated with an overall lower level of expression of EP₂ receptor mRNA.

5 When expression of prostanoid receptor genes was studied in cervix obtained from animals both in labour and not in labour, clear signals which were similar in estimated size to the human genes were detected by Northern analysis for EP₁, EP₂, EP₃, EP₄, FP, IP and TP receptors. Expression of the gene encoding the prostanoid EP₁ receptor increased with advancing gestational age prior to labor. Expression of the EP₂, FP and TP

receptor genes was much lower in animals that were delivered during spontaneous labor than in animals which were not in labor.

6 When expression of prostanoid receptor genes was studied in decidua and chorion obtained from animals both in labour and not in labour, expression of the genes encoding the EP₁ and FP receptor in decidua and the EP₄ receptor in chorion was lower with advancing gestational age. Expression of the EP₂ receptor gene was lower in labour in decidua, whereas expression of the IP receptor gene was higher in labour in both decidua (2-fold) and chorion (4-fold).

7 Expression of both the EP₃ and EP₄ receptor genes was detected in lamb ductus arteriosus and the level of expression of both genes in the fetal ductus was unaffected by maternal administration of corticosteroids. Expression of the EP₄ receptor gene was lower in the ductus obtained from term lambs compared with preterm lambs, and was lower still in neonatal lambs, whereas no variation was observed in EP₃ receptor gene expression with respect to gestational age and birth. Expression of the EP₄ receptor gene was also confirmed in fetal baboon ductus arteriosus and maternal administration of corticosteroid did not reduce EP₄ receptor gene expression in the baboon.

8 Signals of similar size to human 5-LOX and platelet 12-LOX genes were detected in myometrium, cervix, decidua and chorion. Expression of 5-LOX mRNA decreased with advancing gestational age in the cervix. In decidua, expression of 5-LOX mRNA was higher in tissues from animals in labor, whereas in chorion, 5-LOX expression was lower

in tissues from animals in labor. Expression of the platelet 12-LOX gene decreased with advancing gestational age in chorion and was lower in labor in cervix.

9 Cellular localization of the EP₃ receptor was possible in myometrium and of the EP₄ receptor gene in ductus arteriosus. In the latter case, expression of the EP₄ receptor gene was strikingly specific for the ductus arteriosus compared with neighbour vessels. Expression of the EP₃ receptor gene was evident in myometrial smooth muscles cells but not in the smooth muscle cells of muscular arteries supplying the uterus.

10 The following general conclusions are drawn from these studies:

- (a) The myometrial expression of prostanoid receptor genes varies within the uterus and the reduced contractile response of LUS to PGE₂ is paralleled by greater inhibitory EP₂ receptor expression and less contractile EP₁ and EP₃ receptor expression, a similar pattern to the cervix.
- (b) Regional variation in the expression of prostanoid receptor genes persists in labour and suggests a role for regional variation in myometrial sensitivity to prostaglandins in the polarity of uterine contraction
- (c) The pattern of expression of prostanoid receptor genes varies with advancing gestational age in cervix, chorion and decidua. Variation in the sensitivity of these tissues to the effects of PGs may have a role in the preparation for parturition in the baboon.
- (d) The pattern of expression of prostanoid receptor genes varies in association with labour in myometrium, cervix, chorion and decidua. In particular, decreased EP₂ receptor gene expression was noted in myometrium, cervix and decidua. This implies that

variation in the sensitivity of these tissues to PGs in general, and to PGE₂ in particular, might promote parturition in the baboon.

(e) The preparation of the fetus for birth also involved alterations in the relative expression of genes encoding contractile and inhibitory EP receptor sub-types. Specifically, contraction of the ductus arteriosus at birth may be stimulated by loss of inhibitory effects of PGE₂ through the EP₄ receptor in the presence of maintained contractile effects of PGE₂ through the EP₃ receptor.

(f) Variation in expression of LOX genes may play a role in the onset of parturition in the baboon. However, it is not apparent whether the role of LOX is principally to generate biologically active products, or to divert free cellular arachidonic acid to relatively inert products rather than PGs.

Table of Contents	Page
Title page	(i)
Abstract	(ii)
Table of contents	(vii)
List of figures	(x)
List of tables	(xv)
Abbreviations	(xvi)
Presentations arising from work	(xviii)
Full papers, published or in press from this work	(xx)
Acknowledgements	(xxi)
Declaration	(xxii)
Chapter 1. Prostaglandins and parturition: a review	1
1.1 General aspects of mammalian parturition	1
1.2. Uterine activity during pregnancy	2
1.2.1 Ante-partum uterine activity	2
1.2.2 Intra-partum uterine activity	2
1.2.3 Post-partum uterine activity	3
1.3. Prostaglandin pharmacology	4
1.3.1 Biosynthesis of prostaglandins	4
1.3.1.1 Phospholipase enzymes	5
1.3.1.2 Cyclo-oxygenase enzymes	5
1.3.1.3 Metabolism of PGH ₂	8
1.3.2 Prostaglandin signal transduction	8
1.3.2.1 Prostanoid receptor pharmacology	8
1.3.2.2 Prostanoid receptor genetics and second messenger coupling	10
1.3.3 Metabolism of prostaglandins	12
1.4 Prostaglandins and parturition	13
1.4.1 Synthesis of prostaglandins in labour	13
1.4.2 Effects of prostaglandins on key intra-uterine tissues	15
1.4.3 Metabolism of prostaglandins in labour	16
1.4.4 Prostaglandins and fetal cardiovascular adaptation at birth	17
1.5 Non-prostanoid control of labour	18
1.5.1 Oxytocin	19
1.5.2 Steroid hormones	20
1.5.3 Corticotrophin releasing hormone	25
1.5.4 Non-prostanoid eicosanoids	25
1.5.5 Other stimulatory factors	26
1.5.6 Other inhibitory factors	26
1.5.7 Changes in cellular function	28
1.5.7.1 Control of intracellular calcium concentration	28
1.5.7.2 Gap junctions	29
1.5.7.3 G-proteins	29
1.5.7.4 Contractile proteins	30
1.6. Clinical aspects of uterine contraction in pregnancy	30
1.6.1 Preterm labour	30

1.6.2 Induction of labour.....	31
1.6.3 Post-partum haemorrhage	32
1.7 Aims of this work.....	32
1.7.1 Variation in myometrial pharmacology according to uterine site.....	32
1.7.2 Identification of tissue-specific expression of prostanoid receptors	33
1.7.3 Identification of cell-specific expression of prostanoid receptors	34
1.7.4 The effect of labour on prostanoid receptor expression.....	34
1.7.5 Prostanoid receptor expression in the fetus.....	35
1.7.6 Comparison between prostanoid and related systems.....	35
Chapter 2. Methods.....	36
2.1 In vivo methods	36
2.1.1 Animal housing conditions	36
2.1.2 Surgery	36
2.1.2.1 Anaesthesia and pain relief.....	37
2.1.2.2 Maternal femoral vein catheterisation	37
2.1.2.3 Laparotomy.....	38
2.1.2.4 Placement of uterine electromyogram (EMG) electrodes	38
2.1.2.5 Chronic fetal preparations.....	38
2.1.2.6 Caesarean section.....	39
2.1.2.7 Caesarean hysterectomy.	39
2.1.3 Acquisition of data from chronically instrumented animals	40
2.2 In vitro methods.....	41
2.2.1 Northern blot.....	41
2.2.1.1 Extraction of RNA.....	41
2.2.1.2 Gel electrophoresis of RNA.....	43
2.2.1.3 Transfer onto membrane.....	45
2.2.1.4 cDNA probe synthesis	45
2.2.1.5 cRNA probe synthesis	46
2.2.1.6 Source of cDNAs.....	47
2.2.1.7 Hybridisation and washing	47
2.2.2 In situ hybridisation	48
2.2.3 Cell culture	49
2.3 Statistics.....	50
Chapter 3. Regional and tissue variation in prostanoid receptor gene expression.....	52
3.1 Methods	53
3.2 Results	55
3.3 Discussion.....	61
Chapter 4. The effect of labour and gestational age on expression of prostanoid receptor genes in myometrium.....	67
4.1 Methods	68
4.2 Results	70
4.3 Discussion.....	79
Chapter 5. The effect of labour and gestational age on expression of prostanoid receptor genes in cervix	83
5.1 Methods	85
5.2 Results	86
5.3 Discussion.....	93

Chapter 6. The effect of labour and gestational age on expression of prostanoid receptor genes in decidua and chorion.....	97
6.1 Methods	99
6.2 Results	100
6.3 Discussion.....	109
Chapter 7. The effect of corticosteroids, gestational age and birth on expression of EP receptor genes in ductus arteriosus	114
7.1 Methods	116
7.1.1 Obtaining fetal and neonatal lamb ductus arteriosus	116
7.1.2 Instrumented fetal lamb preparations.....	116
7.1.3 Instrumented fetal baboons	117
7.2 Results	118
7.3 Discussion.....	126
Chapter 8. The effect of labour and gestational age on expression of lipoxxygenase genes in intra-uterine tissues.....	131
8.1 Methods	132
8.2 Results	133
8.3 Discussion.....	143
Chapter 9. Overall conclusions.....	147
9.1 Variation in prostanoid receptor expression with advancing gestational age and labour	147
9.2 Regional variation in myometrial function.....	148
9.3 Tissue specific expression of prostanoid receptors	148
9.4 Cell specific expression of prostanoid receptors.....	149
9.5 The concept of stimulatory and inhibitory systems.....	149
9.6 Regulation of prostanoid receptor gene transcription	151
9.7 Technical considerations relating to prostanoid gene expression	151
9.8 Summary.....	153
References	154-
.....	188

List of Figures.....	Page
Figure 2.1	44
Photograph of RNA gel, stained with ethidium bromide, following electrophoresis and prior to Northern blot.	
Figure 3.1	54
Schematic outline of site of sampling from myometrium.	
Figure 3.2A-B.	56
Northern blot of EP ₂ and EP ₃ expression in lower uterine segment (LUS) and fundus from 8 pregnant baboons between 121-180dGA.	
Figure 3.3.	57
Densitometric quantitation of EP ₂ , EP ₃ , EP ₄ and FP receptor mRNA relative to GAPDH comparing lower uterine segment (LUS) and fundus.	
Figure 3.4.	58
Densitometric quantitation of EP ₂ and EP ₃ mRNA in different regions of baboon uterus using lower uterine segment (LUS) as a reference.	
Figure 3.5A-D.	59
Representative Northern blots of EP ₂ , EP ₃ , EP ₄ and FP receptor mRNA in myometrium from anterior corpus (MYO), cervix (CX), decidua and chorion.	
Figure 3.6A-D.	60
Quantitation by densitometry of EP ₂ , EP ₃ , EP ₄ and FP receptor mRNA in myometrium from anterior corpus (MYO), cervix (CX), decidua (DEC) and chorion (CHOR) as a ratio to GAPDH.	

Figure 4.1.	72
Northern blots of EP receptor genes in baboon myometrial mRNA from lower uterine segment (LUS) and fundus (FUN), both not in labour (NIL) and in labour (LAB)	
Figure 4.2.	73
Northern blots of non-EP prostanoid receptor genes in baboon myometrial mRNA from lower uterine segment (LUS) and fundus (FUN), both not in labour (NIL) and in labour (LAB).	
Figure 4.3.	74
Northern blots of housekeeping genes in baboon myometrial mRNA from lower uterine segment (LUS) and fundus (FUN), both not in labour (NIL) and in labour (LAB).	
Figure 4.4.	75
Densitometric quantitation of prostanoid receptor signals (as a ratio to beta-actin signal) for prostanoid receptor types in lower uterine segment (LUS) and fundus (FUN) from animals both not in labour (white bars) and in labour (shaded bars).	
Figure 4.5.	76
In situ hybridisation of EP ₃ receptor riboprobe of section from uterine fundus, obtained from animal not in labour at 177 days gestation.	
Figure 4.6	77
Expression of EP ₂ and EP ₃ receptor genes in fresh and cultured myometrium from uterine fundus of animal in term labour.	
Figure 5.1.	87
Northern blot of prostanoid EP receptor gene sub-types in baboon cervix.	
Figure 5.2.	88
Northern blot of non-EP prostanoid receptor gene sub-types in baboon cervix.	

Figure 5.3.	89
Expression of EP ₁ receptor mRNA in cervix (n=7) with advancing days of gestational age (dGA).	
Figure 5.4.	90
Expression of prostanoid receptor genes relative to beta-actin in cervix of animals in labour (LAB) and not in labour (NIL).	
Figure 6.1.	102
Northern blot of EP receptor mRNA in decidua.	
Figure 6.2.	103
Northern blot of non-EP prostanoid receptor mRNA in decidua.	
Figure 6.3.	104
Northern blot of EP receptor mRNA in chorion.	
Figure 6.4.	105
Northern blot of non-EP prostanoid receptor mRNA in chorion.	
Figure 6.5.	106
Prostanoid receptor gene expression related to gestational age among animals not in labour in decidua and chorion.	
Figure 6.6.	107
Variation in expression of prostanoid receptor genes (expressed as ratio to beta-actin) in relation to labour.	
Figure 7.1.	119
Northern blot of fetal and neonatal lamb mRNA, probed for EP ₄ , EP ₃ , beta-actin and GAPDH genes.	

Figure 7.2.	120
Densitometric quantitation of EP ₄ signal in each of the seven groups relative to beta-actin.	
Figure 7.3.	121
Densitometric quantitation of 4.2kB EP ₃ transcript to A. ratio of EP ₃ to beta-actin B. ratio of EP ₄ to EP ₃ in each of the seven groups.	
Figure 7.4.	122
Northern blot of fetal baboon ductus arteriosus probed for EP ₄ gene and beta-actin.	
Figure 7.5.	123
Densitometric quantitation of ratio of EP ₄ signal to beta-actin in baboon ductus arteriosus.	
Figure 7.6.	124
In situ hybridisation of EP ₄ receptor gene expression in baboon ductus arteriosus and aorta.	
Figure 8.1.	135
Northern blot of LOX expression in myometrium from lower uterine segment (LUS) and fundus (FUN), from animals not in labour (NIL) and in labour (LAB).	
Figure 8.2.	136
Northern blot of LOX expression in cervix from animals not in labour (NIL) and in labour (LAB).	
Figure 8.3.	137
Northern blot of LOX expression in decidua from animals not in labour (NIL) and in labour (LAB).	

Figure 8.4.	138
Northern blot of LOX expression in chorion from animals not in labour (NIL) and in labour (LAB).	
Figure 8.5.	139
Northern blot of 15-LOX expression in A. Myometrium, B. Cervix.	
Figure 8.6.	140
Densitometric analysis of 5-LOX expression (relative to beta-actin) decidua and chorion.	
Figure 8.7.	141
Densitometric analysis of 12-LOX expression in the cervix.	

List of Tables.	Page
Table 1.1 Characteristics of murine prostanoid receptor genes.	11
Table 4.1. Repeated measures analysis of variance of effect of anatomical site and labour on myometrial expression of prostanoid receptor genes, quantified densitometrically as a ratio to beta-actin.	78
Table 5.1. Relationship between gestational age and cervical prostanoid receptor gene expression as a ratio of three housekeeping genes.	91
Table 5.2. Cervical prostanoid receptor gene expression as a ratio of three housekeeping genes comparing animals not in labour and in labour.	92
Table 6.1. Variation in expression of prostanoid receptor genes (relative to beta-actin) related to advancing gestational age and labour.	108
Table 7.1. Three way analysis of variance: effect of fetal surgery, dexamethasone and gestational age and birth on expression of prostanoid receptor genes.	125
Table 8.1. Expression of 5-LOX and platelet 12-LOX in relation to gestational age and labour in baboon intra-uterine tissues.	142

Abbreviations

AA	arachidonic acid
ACTH	adrenocorticotropic hormone
ATP	adenosine triphosphate
BSA	bovine serum albumin
c	complementary
COX	cyclo-oxygenase
CRH	corticotrophin releasing hormone
CTP	cytosine triphosphate
d	deoxy
DHEAS	dihydroepiandrosterone
DNA	deoxyribonucleic acid
EDTA	ethylenediaminetetraacetic acid
EMG	electromyogram
GTP	guanosine triphosphate
HPAA	hypothalamo-pituitary adrenal axis
HPETE	hydroperoxyeicosatetraenoic acid
LOX	lipxygenase
LT	leukotriene
LUS	lower uterine segment
MLC	myosin light chain
MLCK	myosin light chain kinase
MOPS	3-(<i>N</i> -morpholino)propanesulfonic acid

NSAID	non-steroidal anti-inflammatory drug
PG	prostaglandin
PGDH	prostaglandin dehydrogenase
PL	phospholipase
PPH	post partum haemorrhage
PPROM	preterm, premature rupture of the membranes
PTHrP	parathyroid hormone related polypeptide
RNA	ribonucleic acid
TEA	triethanolamine
SDS	sodium dodecyl sulphate
SSC (1x)	0.15M NaCl and 0.015M sodium citrate, pH 7.0
TTP	thymidine triphosphate
Tx	thromboxane
UTP	uracil triphosphate
UV	ultraviolet

Presentations arising from this work.

Smith GCS, Baguma-Nibasheka M, Wu WX, Nathanielsz PW. The reduced contraction of baboon lower uterine segment to prostaglandin E₂ is paralleled by increased EP₂ and decreased EP₃ receptor mRNA compared with fundus. Society for Gynecologic Investigation, 45th Annual Meeting, Atlanta, GA, USA, March 1998, abstract published J. Soc Gyn Invest 1998;5:64A.

Smith GCS, Wu Wx, Nathanielsz PW. Prostaglandin EP receptor sub-types have characteristic patterns of expression in baboon myometrium, cervix, decidua and chorion. Society for Gynecologic Investigation, 45th Annual Meeting, Atlanta, GA, USA, March 1998, abstract published J. Soc Gyn Invest 1998;5:190A.

Smith GCS, Wu Wx, Nathanielsz PW. Baboon labor is associated with decreased uterine prostanoid EP₂ receptor expression. Society for Gynecologic Investigation, 46th Annual Meeting, Atlanta, GA, USA, March 1999, abstract published J. Soc Gyn Invest 1999;6 (supplement 1):128A.

Smith GCS, Wu WX, Nathanielsz PW. Regional, gestational and labor-related variation in expression of prostanoid receptor mRNA in baboon myometrium. Society for Gynecologic Investigation, 47th Annual Meeting, Chicago IL, USA, March 2000, abstract published J. Soc Gyn Invest 2000;7 (supplement 1): 234A.

Smith GCS, Wu WX, Nathanielsz PW. Expression of lipoxygenase genes in baboon

myometrium, cervix, decidua and chorion: effects of gestational age and labor. Society for Gynecologic Investigation, 47th Annual Meeting, Chicago IL, USA, March 2000, abstract published J. Soc Gyn Invest 2000;7 (supplement 1): 239A.

Smith GCS, Wu WX, Nathanielsz PW. Effect of gestational age and labor on the expression of prostanoid receptor genes in baboon decidua and chorion. Society for Gynecologic Investigation, 47th Annual Meeting, Chicago IL, USA, March 2000, abstract published J. Soc Gyn Invest 2000;7 (supplement 1): 128A.

Smith GCS, Wu WX, Nathanielsz PW. Decreased expression of genes encoding excitatory prostanoid receptors in baboon cervix in labor. Society for Gynecologic Investigation, 47th Annual Meeting, Chicago IL, USA, March 2000, abstract published J. Soc Gyn Invest 2000;7 (supplement 1): 250A.

Smith GCS, Wu WX, Nijland, M, Koenen, SV, Nathanielsz PW. Effect of gestational age, corticosteroids and birth on expression of prostanoid receptor genes in lamb and baboon ductus arteriosus. Society for Gynecologic Investigation, 47th Annual Meeting, Chicago IL, USA, March 2000, abstract published J. Soc Gyn Invest 2000;7 (supplement 1): 114A

Full papers published or in press from this work.

Smith, GCS, Baguma-Nibasheka, M, Wu, WX, Nathanielsz, PW. Regional variations in contractile responses to prostaglandins and prostanoid receptor mRNA in pregnant baboon uterus. *American Journal of Obstetrics and Gynecology* 1998;**179**:1545-1552.

Smith, GCS, Wu, WX, Nathanielsz, PW. Effects of gestational age and labor on expression of prostanoid receptor genes in baboon uterus. *Biology of Reproduction* (accepted pending revision).

Smith, GCS, Wu, WX, Nathanielsz, PW. Expression of prostanoid receptor genes in baboon chorion and decidua during pregnancy and parturition. *Journal of Endocrinology* (accepted pending revision).

There are three other first author papers submitted for publication on the work described in this thesis.

Acknowledgements.

I am grateful to many people who were involved in many different ways in this project. I would like to thank Peter Nathanielsz for hosting my three years at Cornell, for his helpful supervision, scientific guidance and his comments on this thesis and the papers arising from it. I am also grateful to Prof Iain Cameron for acting as my Glasgow supervisor and for his comments on the thesis. I am grateful to the Wellcome Trust for their generous funding of my fellowship to work at Cornell. I am grateful to Drs Wen Wu and Dr Ma for their assistance and instruction in the laboratory work and to Dr Sui Xing Ding, Charles Mecnas, Jim Winter and Jennifer Tame for providing the huge level of infra-structural support that is required for the *in vivo* studies. I am grateful to all the other members of the Laboratory for Pregnancy and Newborn Research between 1996-1999 for their contributions to the work described in this thesis. I am grateful to my wife, Nicola and my children Jessica and Alice for being cheerful sources of encouragement over the project and for their willingness to adventure to upstate New York.

Declaration.

This thesis is entirely my own work. The research it describes necessarily involved the contributions of a large group of people and the planning of experiments was outlined in association with Prof Nathanielsz. However, all the laboratory work described in this thesis was performed by myself. The analysis of data and writing up of these experiments was done by myself. Both Prof Nathanielsz and Prof Cameron made constructive comments on the final draft of this thesis and both Prof Nathanielsz and Dr Wen Wu made comments on the individual papers which make up this body of work.

Chapter 1. Prostaglandins and parturition: a review.

One of the key biological features of mammalian species is viviparity. By necessity, the successful completion of the process is essential for propagation of the species. Although parturition is common to all mammals, the physiological properties of the process are as diverse as mammalian species, which is to say different in many respects but with certain common features. Mammalian reproduction is characterized by (1) in vivo fertilization, (2) implantation of the conceptus and formation of the placenta, (3) development of the embryo, (4) growth of the fetus in an enlarging, essentially quiescent uterus, (5) expulsion of live born young, and (6) post-partum contraction of the uterus followed by uterine involution.

A detailed review of any aspect of mammalian parturition would require a volume many times larger than the whole of this thesis and the aims of this chapter are, first, to give an overview of the process of mammalian parturition accompanied with reference to more detailed discussion and, secondly, to discuss in greater detail the role of PGs in the control of parturition.

1.1 General aspects of mammalian parturition

One of the key differences in the control of parturition comparing mammalian species is the role and source of steroid hormones in the control of the process, namely, oestrogen, progesterone and corticosteroids¹. These hormones have key roles in all species, but the source of these hormones and their effects on the process of labour significantly differ. Much of the research on the control of parturition has been in littering animals, specifically the rat. The reason for its widespread use is largely one of easy maintenance

in a laboratory setting. More recently the mouse has been a focus for study as it is readily utilized for studies of null mutations of its genome. A number of these so-called “knock out” experiments have demonstrated that mice which are null mutant for certain genes have delayed parturition, including the FP receptor gene ², cyclo-oxygenase-1 (COX-1) ³ and steroid 5 α reductase type I ⁴.

While studies in these animals can yield important information about the physiological control of parturition in these species, some caution has to be applied when extrapolating these results to the human and the attendant clinical problems which are the stimulus to much of this research. There are important species differences in the role and source of key steroid hormones in the process of labour (see section 1.5.2 for detailed discussion). The key factor regulating parturition in rodents is involution of the corpus luteum (luteolysis). The corpus luteum is the source of progesterone in these animals and high concentrations of progesterone maintain uterine quiescence. Falling concentrations of progesterone in these animals, following luteolysis, promote labour. In sheep, progesterone has the same role but the source is the placenta. In humans and non-human primates, progesterone is also formed in the placenta after the first trimester, but there is no change in circulating concentrations of progesterone preceding labour. Many of the cellular, autocrine and paracrine mediators are common to rodents and higher species. The specific differences are discussed below and have been reviewed elsewhere ¹. Prostaglandins, which are the focus of the present study, are thought to have a key role in the process of parturition in all mammalian species studied so far ⁵.

1.2. Uterine activity during pregnancy

The activity of the myometrium in pregnancy is frequently misunderstood. The state is caricatured as the uterus being quiescent during the antepartum period and then active throughout labour. The reality is that the uterine smooth muscle is active throughout pregnancy but that the patterns of activity are complex and change with its differing physiological roles.

1.2.1 Ante-partum uterine activity

For obvious reasons, most invasive studies of ante-partum uterine activity have been performed in animal species. These studies have revealed that in the ante-partum period the uterus exhibits low amplitude, low frequency activity which has been characterized as “contractures”⁶. This activity is associated with smooth muscle depolarisation, but only induces very small increases in intra-uterine pressure and does not result in dilatation or effacement of the cervix. This activity does, however, alter the environment of the fetus, such as fetal arterial oxygen tension, and the frequency of contractures can influence maturation of fetal cardiovascular control systems⁷.

1.2.2 Intra-partum uterine activity

Uterine activity during labour is characterized by high frequency, high amplitude activity which are termed “contractions”. In the human a typical pattern of intra-partum activity in the second stage of labour would be four contractions every 10 minutes, with a peak pressure of up to 60-70mmHg, and a total duration of 60 seconds with the pressure curve being bell-shaped⁸. From this it is evident that the uterus is only actively generating tension for 2 minutes out of every 10. For another 2 minutes it is actively relaxing and for

the remaining 6 minutes out of every 10, the intra-uterine pressure is at basal levels. Therefore, considering the uterus as merely moving from a purely passive state before labour into a purely active state during labour is not merely an unfortunate simplification, but is quite wrong.

Another physiological feature of intra-partum uterine activity is that it is associated with polarity, i.e. it is directional, forcing the presenting part of the fetus towards the cervix. This suggests that the contractile role of the myometrium during labour is likely to vary according to the site of that myometrium within the uterus. The fundus and corpus of the uterus would be expected to exert more force than the lower (cervical) pole of the uterus. This is another area that has been relatively neglected and is one of the major themes of this thesis.

1.2.3 Post-partum uterine activity.

In many considerations of the physiological roles of uterine contraction, post-partum uterine activity is neglected or even completely ignored. Yet in the absence of modern healthcare (i.e. the evolutionary environment which shaped human physiology), primary post-partum haemorrhage (primary PPH, defined as blood loss of greater than 500ml within 24 hours of delivery) is one of the major causes of maternal mortality⁹. Given that failure of post-partum contraction of the uterus is the major cause of primary PPH, it is clear that development of systems to allow effective post-partum contraction of the uterus were central to the evolution of placental reproduction.

1.3. Prostaglandin pharmacology

Prostaglandins are oxygenated derivatives of arachidonic acid (AA), a fatty acid formed from membrane phospholipids by phospholipases. Prostaglandins are generally synthesized de novo in the tissue which releases them in contrast to other bioactive endogenous agents (such as the monoamines) which are commonly pre-formed and stored in intra-cellular vesicles ¹⁰. Prostaglandins exert biological effects by acting through a family of seven transmembrane domain, G-protein coupled receptors ¹¹. These receptors are coupled to a variety of second messenger systems and exert diverse biological effects ¹². Furthermore, different receptor types and/or sub-types commonly exert opposing effects on the same tissue ¹³. Broadly speaking, PGs act at sites local to their site of synthesis. Some PGs, such as TxA_2 and PGI_2 have very short half lives and degrade before they exert systemic effects ¹⁴. PGE_2 and $\text{PGF}_{2\alpha}$ are more stable, but are rapidly broken down in the lungs by 15 (OH) PG dehydrogenase (PGDH) ¹⁰. An important exception is the fetus, where circulating PGE_2 is in the nanomolar range due to high levels of PGE_2 synthesis from the placenta and low levels of catabolism due to the relatively restricted pulmonary blood flow of fetal life ¹⁵. All aspects of PG pharmacology have been previously reviewed in detail ^{12-14;16}. But an overview of the important aspects is presented below as the pharmacology of PGs is one of the central themes of this thesis.

1.3.1 Biosynthesis of prostaglandins

The main steps in the biosynthesis of prostaglandins (PGs) are: (1) formation of AA from membrane phospholipids, either by phospholipase A₂ (PLA₂) or phospholipase C (PLC), (2) the formation of the cyclic endoperoxides, PGG_2 and PGH_2 from AA by the enzyme

COX, and (3) the modification of PGH₂ to the main native PGs by modifying enzymes, such as PGE isomerase, thromboxane (Tx) synthase and PGI₂-synthase¹⁴. The PG pathway is only one of several pathways of AA metabolism which results in biologically active products¹⁰. Others include the epoxide pathway (cytochrome P450)¹⁷, the isoprostane pathway (non-enzymatic generation of isoprostanes such as 8-epi- PGF_{2α})¹⁸ and the lipoxygenase (LOX) pathway¹⁹.

1.3.1.1 Phospholipase enzymes

Arachidonic acid is formed enzymatically from membrane phospholipids, principally phosphatidyl inositol and phosphatidyl choline. The enzyme PLC forms inositol triphosphate and diacylglycerol from phosphatidyl inositol in response to activation of a range of G-protein coupled receptors and inositol triphosphate acts on heparin sensitive sites to promote intracellular calcium release²⁰. Diacylglycerol is metabolised by lipases to form AA and glycerol. Therefore, AA may be available within the cell following stimulation with a range of agonists. It is postulated that AA may itself act as a second messenger²¹. The other main pathway involved in AA release is PLA₂ and this is thought generally to be the more important pathway in PG biosynthesis, and it forms AA from phosphatidyl choline. A larger molecular weight cytosolic form (cPLA₂) and a number of smaller secretory forms (sPLA₂) have been identified, and these forms differ in their calcium concentration dependence²².

1.3.1.2 Cyclo-oxygenase enzymes

The COX enzyme forms PGH₂ from one molecule of AA and two molecules of oxygen. The same enzyme catalyses two haeme requiring steps, first, an oxygenase activity

forming PGG₂ and, secondly, peroxidase activity forming PGH₂ from PGG₂. Two isoforms of the enzyme have been identified, COX-1 and COX-2 (see ²³). The expression of COX-1 tends to be fairly constant between cells and in a given tissue in different physiological states, and it is frequently referred to as the constitutive isoform. However, dynamic up regulation of COX-1 has been described in the uterine artery in association with the menstrual cycle and pregnancy ²⁴. Expression of COX-2 varies markedly between different tissues and expression in a given tissue may be very significantly increased under physiological and pathological situations, such as labour ²⁵ and infection ²³. COX-2 is frequently referred to as the inducible form of the enzyme, although it does play a constitutive role in the physiology of the kidney, brain and endothelium ²⁶.

The COX enzymes are inhibited by the non-steroidal anti-inflammatory drugs (NSAIDs). Aspirin was the first NSAID discovered, but this category of drugs includes an extensive range of widely prescribed medicines, including indomethacin, ibuprofen, and diclofenac. These drugs are very widely employed as analgesics, anti-pyretics, anti-inflammatories, and anti-spasmodics ²⁷. An increasingly large number of drugs are available which exhibit selective activity at the two COX isoforms ²⁷.

Null mutant mice ('knock-outs') have been described for both isoforms. Mice lacking COX-1 exhibit reduced indomethacin-induced gastric ulceration and platelet aggregation ²⁸. Furthermore, the majority of pups from matings of homozygous males and females die and this appears to be due to delayed parturition³. Mice lacking the COX-2 gene exhibit a severe nephropathy and are susceptible to peritonitis ²⁹. These animals are also infertile, probably due to failure of ovulation and implantation ³⁰.

1.3.1.3 Metabolism of PGH₂

The main naturally occurring PGs which are thought to be of importance physiologically are PGI₂, PGE₂, PGD₂, PGF_{2α} and TxA₂. Each of these is formed from PGH₂ by one (or more) enzymes. Since different PGs often have different effects on a given tissue, the relative activity of these enzymes may have an important role in determining the functional response of a tissue to an increase in free AA. However, a detailed review of these pathways is outwith the scope of this thesis and is available elsewhere ¹⁴. Key enzymes include PGE synthase, PGI synthase and Tx synthase. Some PGs, for instance PGE₂, can be formed by degradation of PGH₂. Furthermore, both PGG₂ and PGH₂ can themselves stimulate the receptor for TxA₂ ¹¹.

1.3.2 Prostaglandin signal transduction

In the 1970s, there was general confusion about the possible existence of receptors for PGs. Such classifications as existed were very complex, ³¹ and some authors postulated that PGs did not act through conventional receptors but rather exerted direct effects on the cell membrane. The work of Dr Robert Coleman at Glaxo Group Research was pivotal in determining the existence of specific receptors for PGs and for understanding their physiological roles.

1.3.2.1 Prostanoid receptor pharmacology

Coleman characterised the relative potency of the main naturally occurring PGs in a wide range of tissues. These studies suggested that there were receptors for each of the main naturally occurring PGs which were at least 10-fold selective for the given PG, and he named them after the native PG: EP receptors for PGE₂, IP receptors for PGI₂, DP

receptors for PGD₂, FP receptors for PGF_{2α} and TP receptors for TxA₂¹³. In addition, using a range of synthetic agonists (and a small number of receptor antagonists), it became apparent that there were sub-types of EP receptor. To date, four distinct sub-types have been identified and these are numbered 1-4³². Broadly speaking, the IP, DP, EP₂ and EP₄ receptors inhibit smooth muscle contractility and the FP, TP, EP₁ and EP₃ receptors stimulate smooth muscle contractility¹². In addition, activation of these receptors influences many physiological processes, including platelet aggregation, renal function, thermoregulation, and immune function¹².

Highly selective agonists exist for some of these receptors such as the IP receptor (e.g. cicaprost³³), the DP receptor (e.g. BW245C³⁴), the TP receptor (U46619³⁵) and the FP receptor (fluprostenol³⁶). Selective agonists have been developed with activity at the EP₂ receptor (butaprost³¹ and AH13205³⁷) and the EP₃ receptor (GR63799X³⁸), but not for the EP₁ or EP₄ receptors. However, even among these “highly selective” agonists, BW245C and GR63799X are agonists of the EP₄ receptor in the nanomolar range³⁹. Selective antagonists exist for some of the receptors. Many antagonists have been described for the TP receptor (e.g. GR32191B⁴⁰). BW868C is a selective DP receptor antagonist⁴¹. AH6809 was originally described as an EP₁ receptor antagonist⁴² but was also demonstrated to exhibit DP blocking activity.⁴³ Both AH23848B and AH13205 antagonize the EP₄ receptor³⁹.

The “classical” prostanoid receptors described above are of the G-protein coupled class with seven transmembrane domains (see below). More recently it has been demonstrated that PGs can activate intra-nuclear receptors which mediate effects by altering gene transcription^{44;45}. The relative contribution of trans-membrane and intra-nuclear receptors in mediating the biological effects of PGs remains to be determined.

1.3.2.2 Prostanoid receptor genetics and second messenger coupling

The functional classification of Coleman has been validated, almost perfectly, by molecular biology ^{11;46}. Genes encoding all eight of the receptor types and sub-types identified pharmacologically have been cloned and sequenced in both mice and humans ^{11;12}. When recombinant receptors are expressed in cell systems, they exhibit very similar binding profiles to the native receptors ⁴⁷ and these studies have also served to underline the lack of truly specific agonists and antagonists. The characteristics of these receptors, the findings in available knock outs, and the chromosomal location of murine prostanoid receptor genes is summarized in Table 1.1.

Table 1.1 Characteristics of murine prostanoid receptor genes.

Gene	Phenotype of knock out	Second messenger	Chromosomal Location
DP	Failure of mast cell derived asthma ⁴⁸	AC (positive) ⁴⁹	14
EP ₁	Not described	IP3 ⁵⁰	8
EP ₂	Impaired ovulation, hypertension ^{51;52}	AC (positive) ⁵³	3
EP ₃	Impaired febrile response, duodenal mucosal dysfunction ⁵⁴	AC (negative) ⁵⁵	
EP ₄	Neonatal death due to patent ductus arteriosus ^{56;57}	AC (positive) ⁵⁸	
FP	Failure of luteolysis/parturition ⁵⁹	IP3 ⁶⁰	3
IP	Thrombophilia, impaired inflammatory responses, hypoalgesia ⁶¹	AC (positive) ⁶²	7
TP	Bleeding diatheses ⁶³	IP3 positive ⁶⁴	10

AC denotes adenylate cyclase IP3 denotes inositol triphosphate.

All the receptor types and sub-types which typically inhibit smooth muscle contractility have been found to be positively coupled to adenylate cyclase, i.e. the IP, DP, EP₂ and EP₄ receptors (Table 1.1). Homology studies have suggested that these receptors form a family, probably formed from mutation of a single ancestral gene ¹². Of the receptors which typically cause smooth muscle contraction, the EP₁, FP and TP receptors are usually positively coupled to the phosphatidyl inositol cascade, whereas the EP₃ receptor is usually negatively coupled to adenylate cyclase (Table 1.1).

A number of the currently identified receptor genes form multiple receptor isoforms due to alternative messenger ribonucleic acid (mRNA) splicing. The best characterized of these is the EP₃ receptor and there are at least eight alternatively spliced isoforms of the human EP₃ receptor ¹². The isoforms differ in the COOH terminal of the protein, which is the intracellular component of the receptor involved in G-protein coupling. The ligand binding site of the isoforms is the same. As stated above, the EP₃ receptor is, characteristically, negatively coupled to adenylate cyclase. However, transfection of different alternatively spliced receptor isoforms into cells demonstrates that different isoforms may be positively coupled to adenylate cyclase, or to the phosphatidyl inositol cascade ^{12;65}. It has also been demonstrated that agonist-induced desensitisation appears to vary directly with the length of the intracellular component ⁶⁶. Multiple bands have been demonstrated on Northern blot for the TP ⁶⁷ and IP ⁶⁸ receptors, and pharmacological studies have suggested isoforms of these receptors ^{69;70}.

1.3.3 Metabolism of prostaglandins

Circulating PGs generally undergo rapidly pulmonary degradation: 95% of infused PGE₂ is inactivated in a single passage through the lungs ¹⁰. In the fetus, pulmonary blood flow

is less than 10% of the combined ventricular output and this is one factor which results in elevated circulating concentrations of PGE₂ in fetal life ¹⁵. The initial step of inactivation of some prostanoids (e.g. PGI₂ and TxA₂) is spontaneous hydrolysis, as they are very unstable. For PGE₂ and PGF_{2α}, the initial step of inactivation is oxidation of the 15-OH to ketone by the enzyme 15-OH-PGDH and further oxidative steps of E series PGs yield metabolites which are chiefly excreted in the urine ¹⁰. PGDH activity is thought to be a major local determinant of local concentrations of PGE₂ and PGF_{2α} in intra-uterine tissues ⁷¹.

1.4 Prostaglandins and parturition

Prostaglandins are thought to have a key role in the initiation of parturition in many species, including the human and non-human primate ⁵. Exogenous PGs can induce delivery of the conceptus at any gestational age in human clinical situations although much lower doses of PGs are required if the administration of PG is preceded by a progesterone receptor antagonist such as mifepristone ⁷². COX inhibitors prolong pregnancy ^{73;74} and can reverse the altered levels of expression of key genes in myometrium in established labour ⁷⁵. PGs are frequently referred to as a final common pathway in the process of parturition and have been implied in both term labour and preterm labour arising from diverse aetiologies ⁵.

1.4.1 Synthesis of prostaglandins in labour

Increased PG biosynthesis has been described in a number of tissues in association with both term and preterm labour. Labour at term is associated with increased circulating fetal ⁷⁶ and maternal ⁷⁷ concentrations of PGs as well as increased concentrations of PGs

in amniotic fluid ^{78;79}. The causal role of these findings in the initiation of labour are supported by the fact that these rises appear to precede the onset of labour. Analysis of key intra-uterine tissues also demonstrates increased expression of both PLA₂ and COX-2, but not COX-1 ⁸⁰. Increased expression of these enzymes in the baboon is observed in the amnion, myometrium, decidua and cervix and in some of these tissues the increase precedes the onset of labour ⁸¹. Studies of amniotic fluid PGs also suggest variation within the cavity of the uterus, with higher concentrations of PGs in the forewaters (i.e. the liquor in the lower pole of the uterus) ⁸². Consistent with this, myometrial expression of COX-2 appears to be greater in samples obtained from the lower pole of the uterus than the upper pole in the baboon, and myometrial COX-2 increases in the lower pole in late gestation, whereas it does not increase in samples from the fundus ⁸¹.

It is postulated that infection may account for a significant proportion of preterm births ⁸³. Clear links between infection, inflammation, and PG synthesis have been demonstrated in humans. Amniotic fluid concentrations of PGs appear to be elevated in preterm labour associated with infection compared with both idiopathic preterm labour and with preterm women not in labour ⁷¹. There is an extensive literature on the effects of cytokines on expression of key enzymes determining the concentrations of PGs, such as PLA₂, COX-2 and PGDH in cells from key intra-uterine tissues, in particular the amnion ⁸⁴. The close link between inflammation and parturition is underlined by the observation that physiological term labour is associated with an infiltrate of inflammatory cells, particularly neutrophils and macrophages, and the process of parturition has much in common with an inflammatory process ⁸⁵.

1.4.2 Effects of prostaglandins on key intra-uterine tissues

Prostaglandins have important effects on key uterine tissues involved in parturition. A number of naturally occurring PGs and synthetic derivatives have effects on myometrial contractility. Using a range of these agents, the prostanoid receptors present in myometrial strips from the lower uterine segment (LUS) obtained from pregnant women at the time of Caesarean section were characterized. These studies demonstrated that activation of DP, IP and EP₂ receptors inhibited myometrial contractility, whereas FP, TP and EP₃ receptor activation stimulated myometrial contractility⁸⁶. The study predated the identification of the EP₄ receptor, and characterization of this receptor is limited by the lack of any selective agonist¹¹. These studies also suggested that the bulk of the contractile effect of PGE₂ on myometrium was mediated by the EP₃ receptor, but there was also possibly an effect mediated through the EP₁ receptor. Studies on cultured human myometrial cells indicate that EP₁, EP₃ and FP receptor activation all appear to stimulate protein kinase C and calcium influx^{87,88}. Interestingly, the intrinsic sensitivity of the EP₃ receptor for PGE₂ is generally 5 to 10-fold greater than the EP₂ receptor⁴⁷. Consistent with this, low concentrations of PGE₂ have been shown to stimulate myometrium, whereas higher concentrations relax myometrium⁸⁶.

An important species difference exists between primates and non-primates: both the human and baboon exhibit inhibitory responses to selective EP₂ agonists^{86;89}, whereas ovine myometrium does not⁹⁰. Expression of the gene encoding the EP₂ receptor decreases in association with advancing gestational age in the human⁹¹ and in association with labour in the rat^{92;93}. Conversely, expression of the gene encoding the excitatory FP receptor is increased in labour in both the rat and human⁹¹⁻⁹⁴.

The factors which regulate prostanoid receptor expression in the uterus in humans and non-human primates have not been studied. Studies in the pregnant rat have demonstrated that myometrial expression of the EP₂ receptor gene is unaffected by either exogenous oestradiol or by blocking the effects of endogenous oestradiol using the oestrogen receptor antagonist ICI 164384 ⁹³. Expression of the FP receptor gene in myometrium was increased by exogenous oestrogen or a progesterone receptor antagonist and was reduced by exogenous progesterone or an oestrogen receptor antagonist ⁹³. Unlike the oxytocin receptor gene, expression of the FP receptor gene was unaffected by stretch ⁹⁴.

PGs also have important effects on the cervix and these have been exploited clinically, where administration of exogenous synthetic PGE₂ is widespread for the pre-treatment of the cervix prior to induction of labour ⁹⁵. The effects of PGs on the cervix are thought to be due both to associated uterine contraction and to direct effects of PGs on the cervix such as decreasing cervical collagen content ⁹⁶. The receptors mediating the direct effect of PGE₂ on the cervix have not yet been identified.

Both the decidua and fetal membranes are sources of PGs ⁵. It is assumed that the effects of PGs released from these tissues are paracrine, specifically, acting on myometrium ⁹⁷. However, there are minimal data on possible effects of PGs on the fetal membranes or decidua. This is partly due to the difficulty in characterizing the effects of hormones on these tissues.

1.4.3 Metabolism of prostaglandins in labour

It is clear that enzymes which are important in the biosynthesis of PGs will have an important role in determining local concentration of PGs in key intra-uterine tissues in association with labour. In the case of PGI₂ and TxA₂ the compounds have very short half

lives and biosynthesis is likely to be the major determinant of local concentrations. However, in the case of PGE₂ and PGF_{2α}, which are thought to have key roles in parturition⁵, metabolism of PGs is also postulated to be important⁷¹. As discussed above, the enzyme PGDH catalyses the first step in inactivation of these compounds. Prior to the onset of labour, this enzyme is expressed in high levels in the chorion⁷¹. Since the chorion lies between the amnion and myometrium and the amnion is seen as the major source of PGs for stimulating myometrial contractility, it has been postulated that chorionic PGDH may have a key role in regulating parturition. Consistent with this it has been shown that both spontaneous term labour and preterm labour in the presence of genital tract infection are associated with decreased expression of PGDH in chorion (see ref⁷¹ for review).

1.4.4 Prostaglandins and fetal cardiovascular adaptation at birth

Preparation for parturition clearly involves preparation of the fetus for life ex utero as well as preparation of maternal tissues to expel the fetus. One of the most acute needs of the neonate is to establish gaseous exchange through the lungs following loss of the umbilico-placental circulation. There are clearly many aspects to this adaptation. One of the best understood roles for PGs in this context is in the control of the ductus arteriosus, which is a shunt blood vessel of fetal life, extending between the main pulmonary artery and the descending aorta. In fetal life, the ductus diverts approximately 90% of the right ventricular output away from the lungs to the aorta and, ultimately, to the placenta⁹⁸. Following birth, the pressure gradient across the ductus reverses and flow is from the aorta to the pulmonary artery. Closure of the ductus occurs in the first 24-48 hours of fetal life⁹⁹.

Like the uterus, PGs have a key role in mediating contraction of the smooth muscle at the time of birth. In fetal life, high local and circulating concentrations of PGE₂ inhibit ductal smooth muscle contractility⁹⁹ and following birth, circulating concentrations of PGE₂ fall promoting contraction of the vessel¹⁵. The ductus has high intrinsic contractile tone: administration of a PG synthesis inhibitor in pregnancy results in contraction of the ductus¹⁰⁰. Ductal tone may be related to an intrinsically high sensitivity of the contractile proteins to calcium whereas PGE₂, acting through adenylate cyclase, inhibits intracellular calcium sensitivity in the vessel¹⁰¹. PGE₂ relaxes the ductus through the EP₄ receptor³⁹ but can also contract the vessel through the EP₃ receptor¹⁰². Mice lacking the EP₄ receptor die at birth due to patent ductus arteriosus^{57;103}. It is as yet unclear why absence of a gene encoding an inhibitory receptor should result in failure of closure of the vessel in the neonatal mouse.

Although the pattern of control of the ductus arteriosus by PGE₂ (PGE₂ inhibits its contractility) is the opposite of myometrium (PGE₂ stimulates contractility), these two smooth muscles exhibit parallels. First, they both express mixed populations of stimulatory and inhibitory prostanoid receptor types and EP receptor sub-types. Secondly, current models for the control of both smooth muscles tend to focus on factors determining the concentration of PGs to which these vessels are exposed. In neither case is it yet clear whether variation in the relative expression of prostanoid receptor types and sub-types might act have a role in the physiological regulation of contractility.

1.5 Non-prostanoid control of labour

The PG system is only one of many that have been demonstrated to have a role in parturition. The process of preterm labour, in particular, may be the result of diverse

pathophysiological processes, and the relative importance of PGs may differ according to the aetiology. Furthermore, many physiological processes have levels of redundancy such that elimination of a single system may have relatively little effect on a process to which it is known to contribute. Thus the phenotypes of many 'knock out' mice are surprisingly normal. A number of other systems involved in the control of the uterus during parturition are discussed below.

1.5.1 Oxytocin

Oxytocin is a nine amino acid peptide which contracts myometrium through the phosphatidyl inositol pathway ¹⁰⁴ and may also stimulate uterine activity by increasing COX-2 expression ¹⁰⁵. It was first localized in the posterior pituitary, although has been more recently identified as being produced in the decidua suggesting autocrine and/or paracrine roles in addition to its well described endocrine effects ¹⁰⁶. In fact, the possible role of oxytocin has been the subject of some debate and some authors maintain its effects on the uterus were entirely confined to the stimulation of postpartum contraction of the uterus and involution since oxytocin is released from the posterior pituitary in association with breast feeding, but circulating levels of oxytocin are not detectably elevated prior to the onset of labour ¹⁰⁷. Consistent with this view, mice which are null mutant for the oxytocin gene have normal labour and delivery, although the neonates die due to failure of lactation in the mother ¹⁰⁸. However, as well as the normal caveats which apply when interpreting these data, species differences in the control of parturition undermine extrapolation of this observation to the human. In mice which are null mutant for COX-1 and oxytocin, parturition occurs normally whereas mice null mutant for COX-1 alone have delayed parturition: this is explained by the opposing luteotrophic effect of

oxytocin and the luteolytic effect of COX-1 mediated increases in $\text{PGF}_{2\alpha}$ ³. The luteotrophic effect of oxytocin complicates interpretation of the loss of myometrial effects of oxytocin in the knock out model.

Synthetic oxytocin has been widely used for many years in the induction of labour and management of postpartum haemorrhage. More recently, selective blockers of the oxytocin receptor have been developed as uterine tocolytics ¹⁰⁹. These agents have resolved some of the debate about the role of oxytocin in late gestation and in labour. Studies on non-human primates have shown that the oxytocin receptor antagonist atosiban inhibits the switch from low amplitude, low frequency contractures, to the high amplitude, high frequency contractions of labour at term ⁶ and a randomised controlled clinical trial demonstrated that the antagonist atosiban had similar tocolytic efficacy to ritodrine ¹¹⁰. The discrepancy between the effectiveness of atosiban in inhibiting labour and the lack of any rise in circulating concentrations of oxytocin during labour may be explained by the drug blocking the autocrine and paracrine effects of decidual oxytocin ^{106;111;112} or by an increase in the number or affinity of oxytocin receptors in key tissues during labour ¹¹³. In the rat, increased expression of oxytocin receptor gene in labour is mediated both by progesterone withdrawal ^{114;115} and mechanical stretch of the uterus ¹¹⁵.

1.5.2 Steroid hormones

The role of steroid hormones differs markedly between species. The key event triggering the onset of parturition in rodents is involution of the corpus luteum (luteolysis) which is mediated by $\text{PGF}_{2\alpha}$ acting through the prostanoid FP receptor ². Loss of luteal progesterone results in parturition. Extrapolation of findings in rodent species to the human and non-human primate is limited by the fact that luteolysis has no role in

parturition in these higher species ¹. In some ruminants, such as the cow, labour is also due to PGF_{2α} mediated luteolysis resulting in progesterone withdrawal ¹¹⁶. However, in sheep, parturition is not related to luteolysis. It was observed that administration of corticosteroids to pregnant ewes induced labour and delivery and catheterisation of fetal lambs demonstrated that there was a rise in cortisol in fetal blood preceding labour ¹¹⁷. The hypothesis that activation of the fetal hypothalamo-pituitary-adrenal axis (HPAA) initiates parturition was supported by the observation that labour can be delayed by interruption of the HPAA at the level of the adrenal, hypothalamus or para-ventricular nucleus ¹¹⁸. Corticosteroids appear to act by inducing placental expression of 17α-hydroxylase which converts progesterone to oestradiol ¹¹⁹. The role of this process in inducing labour is demonstrated by the observations that labour in the sheep is preceded by a sharp rise in maternal circulating concentrations of oestrogen ¹²⁰ and exogenous oestradiol induces labour in the sheep ¹²¹, whereas administration of exogenous progesterone delays labour ¹²². Oestradiol stimulates expression of a cassette of genes in the uterus which promote contraction ¹²³.

Although sheep and primates share the properties that luteolysis occurs early in gestation and progesterone is produced by the placenta ¹²⁴, there are important differences. For instance, there is no fall in progesterone preceding the onset of labour in primates ^{125;126}. Further, administration of exogenous progesterone does not prevent or delay labour ¹²⁷. There is evidence that supports a role for progesterone maintenance of uterine quiescence, however, as administration of a progesterone receptor antagonist induces uterine contraction, although cervical dilatation is apparently less efficient than in normal labour ¹²⁸. This has given rise to speculation that there may be a progesterone withdrawal at the level of its signal transduction system (see below). Furthermore, there is no abrupt

rise in oestradiol in primates as observed in sheep prior to labour, although there is a steady rise in maternal circulating concentrations of oestrogens with advancing gestational age in primates ¹²⁵. Administration of exogenous oestradiol does not induce labour in rhesus monkeys ¹²⁹. However, as in the sheep, a key role has been postulated for the fetal HPA.

The primate (human and non-human) adrenal has a unique fetal zone which produces very large quantities of androgenic precursors of oestradiol ¹³⁰. It is hypothesized that stimulation of the fetal adrenal by adrenocorticotrophic hormone (ACTH) may stimulate the formation of precursors which are metabolised to oestrogens which in turn promote parturition. The hypothesis that androgens from the fetal adrenal might stimulate labour was tested by administering exogenous androstenedione to pregnant rhesus monkeys and this was associated with preterm labour ¹³¹. This would appear to act through the formation of oestradiol as co-administration of an aromatase inhibitor (which blocks the conversion of androstenedione to oestradiol) blocked the effect of androstenedione ¹³². This seems inconsistent with the observation that exogenous oestradiol on its own did not cause labour ¹²⁹. The difference may be due to a necessity for the formation of oestradiol at the site of action. Alternatively, it may be related to effects of androstenedione on androgen receptors which are expressed in key intra-uterine tissues ¹³³.

The control of parturition by steroids in the human appears, as one would expect, to be most similar to non-human primates compared with other species. Progesterone has an important maintenance function for pregnancy. In the first trimester, the corpus luteum is the major source of progesterone but luteolysis occurs early in pregnancy and the placenta is the major source of progesterone in the second and third trimesters ¹³⁴. Removal of the corpus luteum after the first trimester does not terminate human

pregnancy. There is no fall in progesterone preceding labour in women ¹³⁵, but administration of progesterone receptor antagonists can induce delivery in all three trimesters, although the successful termination of pregnancy is more commonly successful when a PG agonist is administered after the progesterone antagonist ⁷². However, exogenous synthetic progesterone does not inhibit labour in humans ¹³⁶.

The effect of progesterone in other species, the effect of progesterone blockade in both the human and non-human primate and the lack of effect of exogenous progesterone have led to theories that allow a role for progesterone withdrawal in the absence of changes in serum concentrations. Possible explanations include altered levels of 17- β ,20 α -hydroxysteroid dehydrogenase resulting in local increases in the ratio of oestrogen: progesterone ¹³⁷, functional antagonism of the progesterone receptor by transforming growth factor- β ¹³⁸, increased expression of heat shock proteins ¹³⁹, and altered expression of progesterone receptor sub-types ¹⁴⁰. The relative contribution of these multiple pathways affecting the response of the uterus to progesterone in the physiological control of parturition remains to be determined.

There is a steady rise in circulating oestrogens with advancing gestation in women ¹⁴¹. Oestradiol and oestrone are formed in approximately similar quantities from maternal and fetal precursor dihydroepiandrosterone sulphate (DHEAS) whereas oestriol is formed principally from fetal DHEAS from the adrenal ¹⁴¹. Release of fetal ACTH may be stimulated by the fetal response to pathological stimuli, such as hypoxia ¹⁴². Alternatively, conversion of maternal cortisol to cortisone by the enzyme 11 β steroid dehydrogenase may reduce suppression of the fetal HPAA by maternally-derived corticosteroid ¹⁴³. Furthermore, corticotrophin releasing hormone (CRH) from the placenta may stimulate ACTH and thus the endocrine changes leading to parturition ¹⁴⁴.

As in the sheep, corticosteroids have important effects on the developing fetus to promote lung maturation and other developmental changes to promote adaptation at birth ¹⁴⁵. These effects are very widely exploited clinically and the use of corticosteroids to accelerate fetal lung maturity at the time of threatened preterm delivery is one of a relatively small numbers of pharmacological interventions in perinatology which reduced the number of perinatal deaths ¹⁴⁵. However, unlike the sheep, betamethasone and dexamethasone do not induce labour: the human placenta lacks 17- α hydroxylase ¹. There are mechanisms, however, by which corticosteroids may influence the process of parturition. First, transplacental passage of corticosteroid suppresses the fetal HPAA and reduces circulating levels of oestrogens and this prolongs gestation ¹⁴⁶. However, corticosteroids stimulate release of CRH from the placenta ¹⁴⁷, which would tend to promote parturition (see below). Secondly, corticosteroids inhibit expression of PGDH in placental and chorion cells which will tend to promote parturition ¹⁴⁸. These data suggest that the cortisol released from the fetal adrenal, although lacking the pivotal role of this hormone in the sheep, may influence the process of labour in the human and non-human primate.

It is currently assumed that, like the primate, the onset of human parturition is related to activation of the fetal HPAA and the combined effects of fetal cortisol and a rise in oestrogen mediated by increased precursors from the fetal adrenal. If this is the case, it suggests that, as in the rhesus monkey, local production of oestrogen is critical, since administration of exogenous synthetic oestrogens to pregnant women does not induce labour ¹⁴⁹. However, the adverse effects of diethylstilboestrol on the genital tract of the developing female fetus ¹⁵⁰ has led to considerable caution in evaluating the effects of

oestrogen receptor agonists to pregnant women and, interestingly, topical oestradiol appears to be an effective agent in priming the cervix ^{151;152}.

1.5.3 Corticotrophin releasing hormone

Corticotrophin releasing hormone is a 41 amino acid peptide hormone, first identified in the hypothalamus, where it is released into the portal circulation to stimulate release of ACTH. Shortly after its identification in the hypothalamus, CRH was demonstrated in the human placenta ¹⁵³. In humans and some – but not all – non-human primates, CRH rises in the third trimester ¹⁵³. Release of CRH is stimulated by oestrogen, corticosteroids and PGE₂ and inhibited by progesterone ^{154;155}. Although CRH does not appear to exert a direct effect of myometrial contractility ¹⁵⁶ expression of the CRH-R1 receptor is increased in association with labour ¹⁵⁷. The precise role of CRH in the direct control of myometrial contractility remains to be determined. However, placentally-derived CRH may stimulate myometrial contractility indirectly by activating the fetal HPAA ¹⁵⁵. Serum concentrations of CRH between 16-20 weeks gestation appear to predict the onset of labour at term ¹⁵⁵. Furthermore, the relationship between placental CRH and inflammation and also with fetal stress suggest that the hormone may have a role in the aetiology of preterm labour as well ¹⁴⁴.

1.5.4 Non-prostanoid eicosanoids

PGs are only one of a number of pathways of locally active agents formed from AA. Other enzymatic pathways are the LOX pathway and the epoxide pathway (cytochrome P450). I could find no data on the possible role of the epoxide pathway in the control of labour. There are a limited number of studies on the LOX pathway. LOX enzymes

generate hydroperoxyeicosatetraenoic acid (HPETE) compounds from AA. There are three main types of enzyme, 5-LOX, 12-LOX and 15-LOX which generate 5HPETE, 12HPETE and 15HPETE compounds, respectively. Of these enzymes, it has been shown that labour in the human is associated with increased expression of 5-LOX gene and protein ¹⁵⁸. Consistent with this, 5-HETE has been shown to increase the contractility of isolated strips of human myometrium whereas 12-HETE was without effect ¹⁵⁹.

There is another pathway for AA, namely, the non-enzymatic formation of isoprostanes, such as 8-epi- PGF_{2α}. These agents act both on the TP receptor ¹⁶⁰ and on a novel receptor ¹⁶¹ and have been shown to have a number of physiological roles in other organs. 8-epi- PGF_{2α} has been shown to have a moderate stimulatory effect on isolated strips of human myometrium ¹⁶² but the role of this agent, if any, in the control of parturition is unknown.

1.5.5 Other stimulatory factors

A range of other hormones have been shown to mediate contraction of the uterus. Specifically, monoamines, such as 5-hydroxytryptamine, noradrenaline and histamine contract isolated strips of myometrium ¹⁶³⁻¹⁶⁵. The role of these agents in the control of labour remains obscure, but it is postulated that release of some of these agents by mast cells may promote uterine contraction during labour and may act synergistically with other systems, such as PGs ¹⁶⁵.

1.5.6 Other inhibitory factors

A range of other inhibitory factors have been identified which may be involved in maintaining uterine quiescence in the antepartum period, for instance, relaxin,

parathyroid hormone related polypeptide (PTHrP) and nitric oxide. In these cases plausible explanations of how these agents may be involved in parturition can be elaborated. For instance, in the case of PTHrP, this agent profoundly inhibits the contractility of myometrial strips from the uterine corpus, lower segment and fundus of the baboon ¹⁶⁶ and release of PTHrP from the amnion may be reduced in association with labour ¹⁶⁷. Despite such associations, information on the causative role of these agents in the regulation of parturition remains to be determined.

It cannot be assumed that because an agent has an inhibitory effect on myometrial contractility, that its effect is purely to inhibit the process of parturition. In the case of both relaxin and nitric oxide (NO), both agents may have effects which promote parturition in some key tissues and inhibit parturition in others. Relaxin inhibits myometrial contractility but has effects on the cervix which would tend to increase cervical compliance ¹⁶⁸. Although, synthetic human relaxin is not effective in priming the cervix prior to induction of labour ¹⁶⁹, increased expression of the gene is implicated in preterm, premature rupture of the fetal membranes (PPROM) ¹⁷⁰. Similarly, NO donors inhibit the contractility of isolated strips of human myometrium ¹⁷¹ and were investigated as potential uterine tocolytics in the management of preterm labour ¹⁷². However, increased activity of endogenous NO is implicated in cervical changes at term and in labour ¹⁷³ and NO donors are effective agents for priming the human cervix prior to termination of pregnancy ¹⁷⁴.

1.5.7 Changes in cellular function

The above principally deals with specific hormone signalling systems. However, general changes also take place in myometrial cellular function which would be expected to influence the response of the uterus to these agents.

1.5.7.1 Control of intracellular calcium concentration

Smooth muscle contraction is mediated by the interaction between the 20kDa myosin light chain (MLC₂₀) and actin which results in activation of myosin adenosine triphosphate (ATP)ase and contraction. The interaction between myosin and actin is regulated by phosphorylation of the serine 19 residue on MLC₂₀¹⁷⁵. Phosphorylation and dephosphorylation of this residue is mediated by key regulatory enzymes, myosin light chain kinase (MLCK) and a smooth muscle myosin phosphatase (SMPP-1M), respectively¹⁷⁶. Raised intracellular calcium concentration ($[Ca^{2+}]_i$) has a key role in initiating smooth muscle contraction, forming a complex with the regulatory protein calmodulin, and the calcium calmodulin complex stimulates contraction by activating MLCK²⁰.

Current models of the control of myometrial contractility invoke a key role for the control of intracellular calcium in the regulation of myometrial contractility¹⁷⁵. $[Ca^{2+}]_i$ may be elevated either by influx of calcium from the extracellular space or by release of calcium from intracellular stores. In human myometrium, agonists induce an increase in $[Ca^{2+}]_i$ due both to influx of calcium from the extracellular space and release from intracellular stores¹⁷⁷. Influx of calcium from the extracellular space is mediated by both voltage sensitive calcium channels and receptor operated channels. The mechanisms which

regulate $[Ca^{2+}]_i$ and the relationship between a given $[Ca^{2+}]_i$ and contraction have been reviewed at length ¹⁷⁸.

1.5.7.2 Gap junctions

One of the features of intrapartum uterine contraction is that contraction is global, i.e. coordinated across the uterus ⁸. A prerequisite for this is the ability of contraction to be propagated through the uterus. In the heart, communication of cells through gap junctions results in a functional syncytium. Unlike the heart, the normal state of the uterus is not that of a syncytium. However, there is a dramatic increase in the expression of gap junctions in myometrium in association with labour which allows excitation of uterine cells to be propagated and facilitates global, coordinated uterine contraction ¹⁷⁹.

1.5.7.3 G-proteins

Many of the signal transduction systems which are thought to have key roles in stimulating contraction during parturition are of the super-family of G-protein coupled receptors ¹⁸⁰. G-proteins are membrane associated proteins which bind guanosine triphosphate (GTP) and couple occupancy of the receptor by an agonist with activation or inhibition of a second messenger ¹⁷⁶. In the case of an inhibitory second messenger, such as adenylyl cyclase, a stimulatory G-protein would mediate relaxation. In the case of a stimulatory second messenger, however (such as PLC), the same G-protein would promote contraction. Conversely, an inhibitory effect of a G-protein on an inhibitory second messenger would initiate contraction. The prostanoid EP₃ receptor typically mediates contraction of smooth muscle in this manner, namely, by a G-protein mediated decrease in adenylyl cyclase activity ¹⁸¹. Labour is associated with decreased myometrial

expression of $G \alpha s$ ¹⁸² which stimulates the inhibitory second messenger adenylate cyclase. The possible role of variation in G-protein expression in controlling human labour is outwith the scope of this thesis and has been reviewed in detail elsewhere ¹⁸⁰.

1.5.7.4 Contractile proteins

There is much less information on the possible role of changes in calcium sensitivity (i.e. the relationship between $[Ca^{2+}]_i$ and contractility) in the physiological and pharmacological control of myometrial contractility. Studies of human myometrium have inferred possible effects of stimulatory agonists on calcium sensitivity from apparent dissociations of calcium concentration and force ¹⁸³. A single study of β -escin permeabilised smooth muscle demonstrated a greater intrinsic calcium sensitivity of human pregnant myometrium compared with non-pregnant ¹⁸⁴ which may be related to a greater degree of contraction for a given level of activation of MLCK ¹⁸⁵. There are, however, multiple mechanisms by which the sensitivity of the contractile proteins to a given $[Ca^{2+}]_i$ can be altered ¹⁷⁸ and these have not been the subject of intense study in relation to myometrial activation in labour.

1.6. Clinical aspects of uterine contraction in pregnancy.

1.6.1 Preterm labour

Preterm labour is frequently referred to as if it is a distinct disease. However, it is a purely descriptive term, being labour and delivery before the start of the 37th completed week following the first day of the last menstrual period. Further, by implication, it is arbitrary as well as descriptive. Labour on the 258th day since the LMP is preterm, whereas labour on the 259th day is not. Preterm delivery may be the result of a diverse

range of clinical precipitating factors including, but not limited to, PPRM, fetal abnormality, polyhydramnios, multiple pregnancy, infection, intra-uterine growth restriction and uterine anomaly ¹⁴⁵. Many cases have no obvious precipitating cause and are referred to as idiopathic. Infection is one key antecedent of many cases of preterm labour, both with and without PPRM.

The ultimate goal of treating preterm labour is the stated aim of many basic studies of myometrial physiology and pharmacology. However, of the three principle clinical goals discussed, management of preterm labour is perhaps the one with the most ambiguous indication. Preterm labour is undoubtedly a major source of morbidity and mortality. Administration of steroids to pregnant women in preterm labour between 1-7 days prior to preterm delivery accelerates fetal lung maturation and reduces perinatal morbidity and mortality ¹⁴⁵. Tocolytic therapy concurrent with steroid administration is justified on the grounds that it might delay delivery to allow steroids to have their maximum effect. However, although tocolytic therapy does delay delivery, it has no effect on perinatal morbidity or mortality ¹⁸⁶. The lack of a clear beneficial effect of tocolysis may be related to the pathophysiology of preterm labour. For example a significant proportion of cases of preterm labour is thought to be secondary to intra-uterine infection and delaying delivery is potentially harmful in this context ^{83;145}. However, preterm labour is the end-point of diverse pathological processes. It is likely that inhibition of preterm uterine contraction confers a differing degree of benefit (or harm) depending on the aetiology.

1.6.2 Induction of labour

In contrast to the treatment of preterm delivery, induction of labour has been shown, in the context of post-dates pregnancy, to reduce both maternal morbidity (Caesarean

section)¹⁸⁷ and may also reduce perinatal mortality¹⁸⁸. The apparent protective effect of induction of labour post-term on risk of Caesarean section is likely to be due to the increased risk of Caesarean section associated with advancing gestational age post-term¹⁸⁹ and the apparent reduction in perinatal mortality may be due to the increased risk of unexplained stillbirth post term¹⁹⁰. However, at a given gestational age, a woman whose labour has been induced is more likely to require a Caesarean section than a woman in spontaneous labour¹⁸⁹. The challenge is how to induce labour in a manner than does not increase the risk of operative delivery.

1.6.3 Post-partum haemorrhage

Failure of contraction of the uterus postpartum with consequent primary postpartum haemorrhage kills approximately 125,000 women in the world each year⁹. In this case there is no ambiguity in the purpose of treatment: it is to contract the uterus and maintain it in a contracted state. An effective, tonic constrictor of the pregnant human uterus that has long-term stability at tropical room temperatures would have considerable potential to reduce global maternal mortality. However, this aspect of the control of the uterus is relatively ignored in the context of myometrial pharmacology.

1.7 Aims of this work.

1.7.1 Variation in myometrial pharmacology according to uterine site

The vast majority of studies of myometrium from pregnant women use biopsies obtained from the lower segment of the uterus, obtained at the time of LUS Caesarean section. It is possible to obtain biopsies from the fundus and corpus of the uterus, but only a small number of studies use this technique. The alternative to this is to use samples obtained at

the time of Caesarean hysterectomy. However, this procedure is performed very infrequently in modern obstetric practice and, commonly, the indication for this radical procedure, e.g. severe atonic PPH, would mean that the results obtained from such specimens could not be extrapolated to the physiological situation. One aim of this project was, therefore, to address regional variation in gene expression in the primate uterus. To this end, the principle technique employed for obtaining tissues was total Caesarean hysterectomy.

1.7.2 Identification of tissue-specific expression of prostanoid receptors

Prostaglandins are known to be important in the control of myometrium and cervix in labour. Although the decidua and fetal membranes are also sites of PG synthesis, it is currently assumed that PGs from these sites have principally a paracrine role by influencing the myometrium or cervix. However, it is much harder to characterize decidual or chorionic function than myometrium function. The possibility that PGs produced in these tissues may have an autocrine effect has not been fully addressed. An aim of this project was to determine whether prostanoid receptor expression could be demonstrated in the fetal membranes and decidua.

Another aspect of this question related to the complex family of prostanoid receptor types and sub-types of EP receptor. Currently, native PGs are typically employed to induce labour. The use of PGE₂ to prime the cervix is dose limited by the effect of PGE₂ to stimulate myometrial contractility. I sought to determine whether there was evidence that prostanoid receptor expression differed in different intra-uterine tissues.

1.7.3 Identification of cell-specific expression of prostanoid receptors

Although analysis of total prostanoid receptor gene expression in a given tissue allows inferences to be drawn about the role of a given gene in the control of parturition, this approach ignores the cellular complexity of intra-uterine tissues. Even taking the simple example of smooth muscle in the uterine wall, there are two types of smooth muscle, namely, myometrial and vascular. There are also many other non-muscle cells types, such as fibroblasts and leukocytes (the latter infiltrate the uterine wall in association with labour). One aim of the project was to determine the cellular localization of prostanoid receptor genes.

1.7.4 The effect of labour on prostanoid receptor expression

A number of groups have made systematic study of expression of families of genes in labour in order to determine systems which are likely to have key roles in the control of parturition. These studies have led to the concept of a cassette of genes encoding myometrial contraction associated proteins which have a key role in labour. Despite the key role postulated for PGs in the control of labour, there has been no systematic study, to my knowledge, of the expression of prostanoid receptors in myometrium in any species. One aim of this work was, therefore, to describe the effect of labour on expression of all eight currently recognized G-protein coupled prostanoid receptors.

As discussed above, I also postulated that prostanoid receptors may have a role in the control of other, non-myometrial tissues, in the process of labour. I sought to infer the possible role, stimulatory or inhibitory, of prostanoid receptors in other tissues in labour, by determining the effect of labour on the level of expression of each of the currently recognized prostanoid receptor genes.

1.7.5 Prostanoid receptor expression in the fetus

Given that successful reproduction necessitates that delivery of the fetus by the maternal uterus has coincided with sufficient fetal development to allow independent survival of the newborn, it is clear that the processes required for fetal adaptation must mature in a parallel fashion with the preparation for labour in uterine tissues. Given that the fetal ductus arteriosus, like the uterus, is critically regulated by PGE₂, I sought to determine whether there were any parallels in developmental changes in prostanoid signal transduction in the ductus. I sought, therefore, to determine the effect of advancing gestational age and birth on the expression of prostanoid receptor genes in ductus arteriosus smooth muscle.

1.7.6 Comparison between prostanoid and related systems

A key role has been postulated for prostanoids in the control of parturition. However, as discussed above there are other pathways of AA metabolism which may generate eicosanoids with a role in the control of key intra-uterine tissues. While the expression of COX enzymes has been systematically studied in relation to labour in a number of species, there has been no systematic study of the expression of LOX genes in relation to labour in the human and non-human primate. A further aim of this work was, therefore, to determine whether expression of LOX genes changed in association with labour in key intra-uterine tissues in the baboon.

Chapter 2. Methods.

2.1 In vivo methods

All animal work was overseen by the Cornell Institutional Animal Care Committee and was under the terms of the current US animal legislation. Baboons were bred in one of two facilities in the USA (San Antonio and Chicago) and transferred to Cornell at approximately 80 days gestational age (dGA). Gestational age was assessed by a known date of mating and by early ultrasonography.

2.1.1 Animal housing conditions

Animals were housed in individual cages. At least 4 weeks prior to study, all animals were given a full medical examination to determine that they were in good health. At this time they were acclimated to a controlled light: dark cycle and animals which were destined for protocols involving chronic instrumentation were fitted to a jacket and tether restraint system ⁶. A nutritionally balanced maintenance diet (Teklad 25% Monkey Diet (W) 8663, Harlan Inc., Madison, WI) and water were supplied to animals ad lib. Feed was supplemented daily with fresh fruits and/or vegetables and one chewable multivitamin tablet daily (Topco, Skokie, IL).

2.1.2 Surgery

All surgical procedures were carried out under general anaesthesia under conditions of strict asepsis.

2.1.2.1 Anaesthesia and pain relief

Anaesthesia was induced by ketamine given by intra-muscular injection. The animal was then transferred to an operating theatre and anaesthesia was maintained by inhalation using halothane, 1-2%, initially administered through a face mask and subsequently through an endotracheal tube. Post operatively, all animals received analgesia for three days (continuous intra-arterial infusion of Buprenorphine [Reckitt & Coleman, Ltd., Richmond, VA], $0.03\text{mg.kg}^{-1}.\text{day}^{-1}$) and prophylactic antibiotics for 5 days (Oxacillin, Bristol-Myers Squibb, Princeton, NJ) $100\text{mg.kg}^{-1}.\text{day}^{-1}$) via continuous intravenous infusion. All catheters were continuously infused with heparinized saline (25 units of heparin. ml^{-1} at 0.5ml.h^{-1} to the mother and at 0.125ml.h^{-1} to the fetus). Heparinized saline was also used as the vehicle for infusion of post-operative medications. When required with fetal preparations, the oxytocin antagonist Atosiban (RW Johnson Pharmaceutical Research Institute, Raritan, NJ), $0.188\text{g.kg}^{-1}.\text{min}^{-1}$ was given to alleviate uterine contraction activity.

2.1.2.2 Maternal femoral vein catheterisation

A 3cm incision was made in the long axis of the anatomical course of the femoral artery approximately 1cm below the inguinal ligament. The femoral vessels were exposed by blunt dissection and the distal end of each vessel was tied. Catheters were advanced into the vessel to 15cm, secured to surrounding connective tissue by interrupted 3/0 silk sutures, tunnelled subcutaneously to the exit point on the animal's dorsum and connected to the tether system. The skin incision was closed with 3/0 Vicryl.

2.1.2.3 Laparotomy.

The abdomen was opened through a mid-line incision. This was usually sub-umbilical, but occasionally extended above the umbilicus. The length of the incision varied with the procedure to be undertaken. Bipolar diathermy was applied to secure haemostasis. The rectus sheath was identified and cut in the midline. The linea alba was identified, divided in its vertical axis and the two halves of the rectus abdominis separated in this plane. The peritoneum was grasped with two small artery forceps, confirmed empty and opened with a blade. The peritoneal incision was extended with scissors. Repair was effected using a mass closure techniques employing 1 Vicryl in a continuous suture. The skin was closed using sub-cuticular 3/0 Vicryl.

2.1.2.4 Placement of uterine electromyogram (EMG) electrodes.

The abdomen was opened as described above, with an incision of no more than 4cm. The anterior surface of the uterus was identified. Five EMG electrodes were stitched into the anterior uterine wall. The wires entered a common sheath which was secured to the parietal peritoneum on the anterior abdominal wall by a suture. The electrodes were connected to the tether system as described for the femoral vascular catheters.

2.1.2.5 Chronic fetal preparations

A detailed description of the techniques involved here is outwith the scope of this thesis as fetal preparations are peripheral to the main focus of the work. In brief, the uterus was opened with a scalpel and the cut edge was grasped with Babcock forceps. A needle was passed through the exposed fetal membranes and amniotic fluid withdrawn and stored at 42C. The membranes were then excised and grasped with the Babcock forceps along with

the myometrium. When the fetal skin was incised, its cut edge was also incorporated with the membranes and myometrium to prevent loss of amniotic fluid. Intra-uterine instrumentation varied between preparations. As a minimum it would include an intra-amniotic catheter and fetal carotid arterial catheter and variably included jugular venous catheter, carotid arterial flow probe, EMG electrodes (biceps and/or triceps), ECG electrodes, and tracheal catheter. The catheters, with or without leads, were connected to the tether system as described for the maternal instrumentation.

2.1.2.6 Caesarean section.

The abdomen was opened as described above. The uterine incision was transverse through the lower part of the uterine corpus. The full thickness of myometrium was opened with a blade to expose the fetal membranes and the cut edges of myometrium grasped with Babcock's forceps for haemostasis. At this stage, specimens of amniotic fluid were frequently removed. The membranes were then incised and the uterine incision extended with scissors. The fetus was delivered, and the cord clamped and divided. The fetal membranes were carefully stripped off and the placenta removed intact with gentle traction on the umbilical cord. The uterine cavity was explored manually and confirmed empty. The uterine incision was closed with a single layer of continuous locked 2/0 Dexon. Closure was otherwise as described above.

2.1.2.7 Caesarean hysterectomy.

The abdomen was opened as described above and the uterus was incised in the midline. After delivery of the fetus, placenta and membranes, the round ligaments were clamped and tied. The broad ligament was opened and the peritoneum was opened anteriorly. The

peritoneal incisions on either side were joined anteriorly and the bladder reflected inferiorly to below the level of the cervix. The peritoneum over the posterior aspect of the uterus was incised and the inferior cut edge was reflected inferiorly. In this way the rectum, if adherent to the Pouch of Douglas, was reflected inferior to the ultimate incision in the vagina. The infundibulo-pelvic ligament was then clamped, divided and tied (2/0 Dexon, lateral to the ovaries if oophorectomy was also being performed). Clamps were advanced lateral to the uterus to secure the uterine arteries. These pedicles were cut and tied (2/0 Dexon). The cardinal ligaments were then clamped, cut and tied (2/0 Dexon). The anterior vaginal wall at the level of the external cervical os was grasped and opened. The incision was extended laterally and the vascular angles were clamped. The uterus was freed by cutting the posterior wall of the vagina and the specimen was removed for immediate harvesting of tissues. The vaginal vault was closed with continuous locking 2/0 Dexon and the abdomen was closed as described above.

2.1.3 Acquisition of data from chronically instrumented animals

Maternal and fetal catheters and electrodes exited the animal's dorsum and entered the tether on its jacket. The tether exited the cage and was connected to transducers (for the catheters) and pre-amplifiers (for the electrodes). These signals were digitised, averaged over 6 second periods, and stored in an electronic form in a computer hard drive. They were also printed out on a dot matrix printer to provide a hard copy for each animal. A detailed discussion of the electronics of the system is outwith the scope of this thesis and has been described elsewhere ⁶.

2.2 In vitro methods

All reagents employed were molecular biology grade. Water was ultra filtered, deionized to 18.3 M Ω resistance and had less than 2 parts of organic impurity per billion. Glassware was carefully washed, rinsed with ultra filtered water and baked for at least 5 hours at 270C. As far as possible, disposable plastic ware was used which was sterile and individually wrapped. If it was not, it was autoclaved for 30 minutes. Items which were not suitable for either baking or autoclaving were treated with a commercial RNase inhibitor (RNase Zap, *Ambion*, Austin, TX, USA).

2.2.1 Northern blot.

Northern blot is a technique which allows the quantification of a specific ribonucleic acid (RNA). The process of Northern blot involves (1) extraction and quantitation of RNA, (2) electrophoresis of denatured samples in an agarose gel, (3) transfer of RNA from the gel to a solid medium where it is fixed (nitrocellulose or nylon membrane), (4) hybridisation of the blot with a labelled probe and (5) autoradiography of the probed blot.

2.2.1.1 Extraction of RNA.

Commercial kits were employed for extraction of RNA which were selective for polyadenylated RNA. For samples of less than 0.2g, Micro Fast track, version 2.2 was employed and for samples between 0.2 to 1.0g, Fast Track, version 2.0, was employed, both kits are made by *Invitrogen*, San Diego, CA. The overall process is similar for both kits and involves homogenisation and incubation of the sample in a buffer containing protein/RNase degrader, mixing of the sample with oligo dT cellulose to allow binding of polyadenylated RNA to the cellulose, washing of the cellulose pellet to remove protein,

deoxyribonucleic (DNA) and ribosomal RNA, and then elution of polyadenylated RNA from the pellet. The detailed protocol was as follows.

A sample of tissue stored at -80°C was placed in a sterile plastic tube containing 200mM NaCl, 200mM Tris (pH 7.5), 1.5mM MgCl_2 , 2% sodium dodecyl sulphate (SDS) and RNase and protein degrader. The sample was then homogenised for 1 minute and incubated at 45°C for 1 hour. The specimen was spun at 4000g for 5 minutes, the supernatant transferred to a clean labelled tube, 5M NaCl was added and mixed. DNA in the specimen was sheared by drawing the sample into a syringe through a 21G needle 4 times. The oligo dT cellulose pellet was added and the specimen mixed in a rocker for 1 hour. The suspension was then spun at 3000g for 5 minutes and the pellet washed three times in 500mM NaCl, 10mM Tris-Cl (pH 7.5). The pellet was then re-suspended in 250mM NaCl, 10mM Tris-Cl (pH7.5) and spun at 3000g for 5 minutes and this was also repeated 3 times. The cellulose pellet was re-suspended in the same buffer, transferred to a spin column and washed three times. The spin-column was transferred to a clean microfuge tube and the polyadenylated RNA was eluted by two washes with 200 μl of 10mM Tris-Cl pH 7.5, heated to 65°C . The RNA was precipitated with 0.15 volume of 2M sodium acetate and 2.5 volumes of 100% ethanol, placed in a -80°C freezer until solid, thawed and centrifuged at 14,000g for 30 minutes at 4°C and the ethanol removed. The pellet of RNA was suspended in 10 mM Tris (pH 7.5) and the concentration and purity of RNA determined in a spectrophotometer using the ratio of the optical density at 260nm and 280nm with a ultraviolet (UV) light source. This procedure typically yielded RNA with a 260:280 ratio of 2.1-2.2.

2.2.1.2 Gel electrophoresis of RNA

The samples of the test RNA and tube containing a standard series of RNA size markers (Gibco) were concentrated by centrifugation in a Speed Vac for 10-20 minutes. Each pellet was then dissolved in 15 μ l of 17.4% (vol/vol) formaldehyde, 50% (vol/vol) freshly de-ionised formamide, 20mM MOPS [3-(*N*-morpholino)propanesulfonic acid], 5mM sodium acetate, 0.025% bromophenol blue and 1mM ethylenediaminetetraacetic acid (EDTA), pH 7.0. Re-suspension was effected by vigorous pipetting and brief vortexing. Samples were concentrated at the bottom of the tube by a 5 second spin at 5000g. They were then heated for 5 min at 65C and chilled on ice for 3 minutes. The samples were loaded into wells of a 1.4% (wt/vol) agarose gel containing 0.66M formaldehyde, 20mM MOPS and 0.28 μ g per ml ethidium bromide in an electrophoresis tank containing 0.66M formaldehyde and 20mM MOPS. The voltage was generally set at 100-110V for large gels and 70-80 for small gels. After the electrophoresis was completed (typically 3-4 hours), the gel was photographed under UV light (see Figure 2.1).

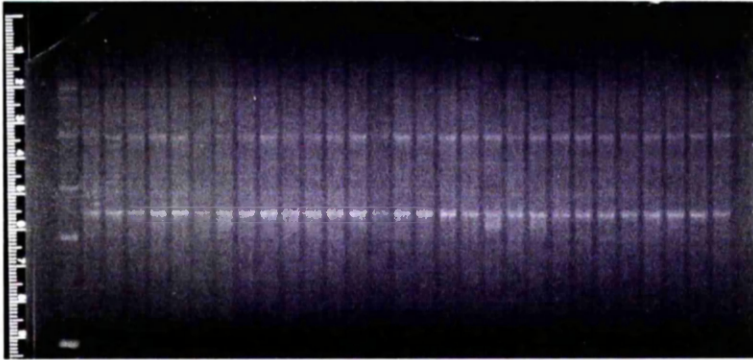


Figure 2.1 Photograph of RNA gel, stained with ethidium bromide, following electrophoresis and prior to Northern blot. 2 μ g of myometrial mRNA loaded in each lane, except first lane = 2.5 μ g of RNA ladder.

2.2.1.3 Transfer onto membrane

The blot was washed 4 times for 5 minutes in water. The nylon membrane (Gene Screen plus, NEN, Dupont, DE, USA) was cut to the size of the gel, washed in water and then in 10X SSC (1X SSC is 0.15M NaCl and 0.015M sodium citrate, pH 7.0). The RNA was transferred onto the membrane by capillary blotting for 24 hours in 10X SSC. The gel was viewed under UV light to confirm that the RNA was transferred. The membrane was then washed briefly in 2X SCC and left over night. The process of drying was sufficient to fix the RNA onto the membrane. It was stored in a sealed bag at 4C until hybridisation.

2.2.1.4 cDNA probe synthesis

Probes were made from full length human complementary DNA (cDNA). The cDNAs were removed from the plasmid by restriction enzymes and separated in a low melting point agarose gel. The DNA corresponding to the probe was cut out of the gel and stored at -20C. Prior to use, the insert (in gel) was heated at 65C for 10 minutes. cDNAs were labelled with [α -³²P] deoxy-cytosinetriphosphate (dCTP) (3000Ci/mmol) using the random priming method to specific activities of approximately 1×10^9 cpm/ μ g using a number of commercial kits, including NEG 013H (NEN, Dupont, Boston, MA, USA), Decaprime and StripeZ DNA (both Ambion) using a method described in detail elsewhere ¹⁹¹. All kits used the same principle of denatured cDNA in an appropriate buffer with random decamers, deoxynucleoside mixture (dATP, dGTP and dTTP, [α -³²P] dCTP and large fragment DNA polymerase. Labelled and unlabelled DNA were then separated from unincorporated nucleotide using Sephadex G-50 chromatography. The probe was quantified in a scintillation counter and, prior to use, was boiled for 5 minutes and chilled on ice for 3 minutes. The advantage of the Decaprime kit over the NEG kit

was that it required less template DNA resulting in less unlabelled DNA in the final probe. The advantage of the StripEZ DNA kit was that one of the nucleotides was chemically modified allowing the probe to be removed with stripping solutions at 65C with minimal loss of blotted RNA.

2.2.1.5 cRNA probe synthesis

The prostanoid receptor cDNAs had been cloned into the pcDNA-3 vector (Invitrogen), which includes promoters for phage polymerases SP-6 and T-7, allowing synthesis of sense and anti-sense riboprobes. The plasmid was linearized by an appropriate restriction enzyme for the given promoter and type of probe. Probes were synthesized using a commercial kit (StripEZ RNA [Northern] or MaxiScript [in situ], both *Ambion*, TX, USA) and labelled with [α - 32 P] uracil triphosphate (UTP) (800Ci/mmol) (NEN Life Science) for Northern analysis and with 35 S UTP (1250Ci/mmol) for in situ hybridization (see below). In both kits, the linearized plasmid is incubated in appropriate buffer (containing DTT and spermidine); unlabelled ATP, CTP and GTP; labelled UTP; the appropriate RNA polymerase and a ribonuclease inhibitor. The StripEZ RNA kit differed in that one of the unlabelled nucleotides was modified and this allowed the probe to be stripped using commercially supplied buffers at 65C with minimal loss of blotted RNA. Template DNA was removed by addition of 2U of RNase-free DNase and incubation for 15 minutes at 37C. Probes was separated from unincorporated nucleotide using a Sephadex G-50 spin column (Probe Quant G-50, Pharmacia Amersham) and quantified. Unlabelled competitor complementary RNA (cRNA) was synthesized using a commercial kit (MegaScript, Ambion) which was optimized for large scale synthesis of

RNA, allowing the unlabelled competitor to be added in greater than 100-fold excess to the labelled probe as a control when employing in situ hybridization (see below).

2.2.1.6 Source of cDNAs

The human EP₂ receptor cDNA was obtained from Dr D.F. Woodward of Allergen, Irvine, CA 92713, USA. The human prostanoid receptor EP₃, EP₄, IP and FP cDNAs were obtained from Dr Mark Abramovitz of Merck Frosst, PO Box 1005, Quebec H9R 4P8, Canada. The human TP receptor cDNA was obtained from Oxford Biomedical (MI, USA) and the mouse EP₁ receptor cDNA was obtained from Dr Y Sugimoto of Kyoto University, Japan. The plasmids (all TRIsScript, Ambion) containing the glyceraldehyde-3-phosphate dehydrogenase (GAPDH), beta-actin and cyclophilin cDNAs with RNA polymerase promoters were purchased from Ambion. The cDNAs for human 5-LOX, platelet 12-LOX, and 15-LOX and for murine leukocyte 12-LOX were all purchased from Oxford Biomedical.

2.2.1.7 Hybridisation and washing

Blots were pre-hybridised and hybridised in the same buffer except the latter contained the nucleic acid probe. In initial experiments using cDNA probes, hybridisation buffer of the following composition was employed: freshly de-ionised formamide (50% vol/vol), sodium phosphate (50mM), NaCl (0.8M), SDS, 2% wt/vol, herring sperm DNA (100µg/ml), yeast transfer RNA (20µg/ml) and 1X Denhardt's (1X = 1% solution of bovine serum albumin [BSA], Ficoll and polyvinylpyrrolidone). In later experiments, commercial hybridisation buffer was employed which was also based on 50% formamide (Northern Max or Ultrahyb, both Ambion).

When cDNA probes were employed, pre-hybridisation (>5 hours) and hybridisation (>18 hours) were carried out at 42°C in sealed bags. Blots were washed sequentially in 2X SSC at 42°C, 0.5X SSC and 0.1%SDS at 65°C (15 minutes), and 0.2X SSC and 0.1%SDS at room temperature (10 minutes). When cRNA probes were employed, pre-hybridisation (>1 hour) and hybridisation (>18 hours) were carried out at 65-68°C in bottles in a hybridisation oven (Hybaid, UK). Membranes were washed at 65°C: twice for five minutes in 2X SSC and 0.1% SDS and twice for one hour in 0.1X SSC and 0.1%SDS. In both cases probe was added to the pre-hybridisation buffer to a concentration of 1 million counts per minute per ml of buffer. Following washing, Kodak X-Omat film was exposed to the membrane with an intensifying screen at -80°C. Following autoradiography, membranes were stripped according to the manufacturer's protocol. Densitometric analysis of autoradiographic signals was performed by scanning autoradiographs into Adobe Photoshop and then using Scan Analysis (Biosoft, Cambridge, UK) to quantify signals.

2.2.2 In situ hybridisation

Frozen sections (5-8 microns thick) cut onto commercially prepared poly-L-lysine coated slides (Sigma Chemical Company) were fixed in freshly prepared 4% paraformaldehyde in 0.1M phosphate buffer (5 minutes), washed twice in 0.1M phosphate buffer, immersed in triethanolamine (TEA)-HCl (3.71g TEA, 2ml 6M NaOH and 198ml of water), pH 8.0 and then TEA and acetic anhydride (0.25%) for 10 minutes. They were then washed in 2X SSC for 5 minutes and briefly in 70% ethanol and allowed to air dry. The specimens were incubated for 2 hours in a humidified container (55°C) with 50-100µl of pre-hybridization buffer (50% freshly deionized formamide, 0.3M NaCl, 20mM Tris-HCl

(pH 8.0), 5mM EDTA, 10mM sodium phosphate buffer (pH 8.0), 1x Denhardt's solution, in RNase free water), and for least 16 hours in hybridization buffer (i.e. pre-hybridization buffer plus probe = 1×10^6 cpm per specimen, between 50-100 μ l). Control slides were hybridized with a sense riboprobe or an antisense probe in the presence of an excess of unlabelled competitor. Slides were then washed three times in 4X SSC and 4mM DTT; three times in NTE buffer (0.5M NaCl, 10mM Tris-HCl and 5mM EDTA, pH 8.0) at 37C (second NTE wash was for 30 minutes with 30 μ g/ml Ribonuclease A); 2X SSC and 1mM DTT; 0.1X SSC and 1mM DTT at 60C; and, finally 0.1SSC at room temperature. The slides were then dehydrated in a graded series of ethanol: 50%, 85% (both plus 0.3M ammonium acetate) and 100%. They were air dried overnight, dipped in emulsion (Kodak NTB2) and exposed for 1-4 weeks at 4C. Following developing and fixing they were counter stained with haematoxylin and eosin, mounted (Permount) and covered with a glass coverslip.

2.2.3 Cell culture

All procedures were carried out using strict aseptic technique. All manipulations of cells were performed in a laminar flow hood. All reagents used were tissue culture grade. If reagents were not already sterile (e.g. culture medium), they were filter sterilized. All reagents employed were obtained from the Sigma Chemical Company, unless otherwise stated. Myometrial cells were established in primary culture using the methodology described by Casey et al, ¹⁹² with certain modifications. Approximately a gram of myometrium was obtained at surgery and placed in Hank's balanced salt solution (buffered with HEPES [20mM] and sodium bicarbonate to pH 7.4) containing penicillin (1000U/ml), streptomycin (1mg/ml) and amphotericin (2.5 μ g/ml). The tissue was minced

into fragments of about 1mm^3 using two sterile scalpel blades and transferred to Hank's balanced salt solution containing collagenase (type II, Gibco BRL) 2mg/ml, DNase (0.2mg/ml = 1400U/ml), penicillin (200U/ml) streptomycin (200 $\mu\text{g}/\text{ml}$) and amphotericin 0.5 $\mu\text{g}/\text{ml}$. The tissues was incubated in this solution (with agitation) for 12-16 hours. The suspension was then drawn into a 30ml syringe 4 times (to disperse the cells further) and filtered through 4 layers of sterile gauze cloth. The filtrate was centrifuged at 600g for 10 minutes and the pellet suspended Ham's F-12 Dulbecco's minimal essential medium containing 10% fetal bovine serum (v/v), penicillin (100U/ml), streptomycin (0.1mg/ml) and amphotericin (0.25 $\mu\text{g}/\text{ml}$). The cells were then placed in a tissue culture flask and incubated at 37C in 95% air 5% CO₂. The culture medium was changed every 48-72 hours. Cell growth was assessed by visual inspection using a phase contrast microscope and confluence was reached in about 7 days. When confluent, the culture medium was removed and replaced with Hank's balanced salt solution (lacking calcium and magnesium) containing HEPES (25mM, pH7.2) and Trypsin-EDTA solution and incubated for 10 minutes at 37C until the cells detached. The cell suspension was added to two volumes of culture medium, centrifuged for 10 minutes at 600g and then re-suspended in culture medium and pelleted again. The cells were then re-suspended in culture medium and counted. The cells were then plated at a density of 100,000 per cm^2 .

2.3 Statistics

Continuous variables were summarized by the mean and standard error of the mean (SEM). Comparison of two means was made by Student's t-test. Comparison of three or more means was made using analysis of variance (ANOVA). Comparison of paired data was performed using Student's paired t-test when comparing two groups and by repeated

measures ANOVA when comparing three or more groups. Multiple comparisons following ANOVA was performed using Tukey's method. Comparison of means adjusted for a continuous covariate was performed using analysis of covariance. Correlation was determined using both the Pearson correlation coefficient and by Spearman's rho. Statistical significance was assumed at the 5% level. Statistical analysis was performed using SPSS version 9.0 (SPSS Inc, Chicago IL, USA) and Stata version 6.0 (Stata Corporation, College Station, TX, USA).

Chapter 3. Regional and tissue variation in prostanoid receptor gene expression

To deliver the fetus, the uterus must exert force with a degree of polarity to direct the baby through the cervix and birth canal, which necessitates a relative relaxation of the LUS. Following delivery, however, the whole uterus must contract to prevent hemorrhage. Effective delivery of the fetus and contraction of the empty uterus post-partum are essential for neonatal and maternal survival. But the process by which differential, regional control of the uterus is effected is as yet unknown.

I sought to test the hypothesis that differential control of the upper and lower poles of the uterus would be reflected in regional variation in the response to PGs and that any such variations would in turn be paralleled by regional differences in the expression of prostanoid receptor mRNA. Furthermore, given the large family of receptor types and sub-types, there is clear potential for the use of selective agonists and antagonists, as appropriate, in the clinical situations described section 1.6. Therefore, I also sought to describe the pattern of expression of prostanoid receptors in key intra-uterine tissues.

3.1 Methods.

Caesarean section hysterectomy was performed on 8 animals at the following days gestational age (dGA): 121, 153, 159, 162, 162, 177, 177, 180 (term=180-185 days) and samples of myometrium were obtained from different regions of the uterus. The cervix was un-effaced and closed in all animals. Uterine electromyogram leads had been sited in 3 hysterectomy animals going to term (delivered at 177, 177 and 180dGA). Analysis of EMG traces of the 48 hours preceding surgery revealed no contraction activity. No drugs of any form had been administered to any of the animals in the 2 weeks preceding surgery. Samples were dissected immediately following removal of the uterus and flash frozen. The LUS was defined as the portion of the uterus superior to the internal os of the cervix. The fundus was defined as the portion of the uterus superior to the upper limit of the uterine cavity. Samples from the corpus were obtained from both the anterior and posterior wall. Upper and lower corpus were defined as lying above and below (respectively) the midline between the upper limit of LUS (which I took as 1cm superior to the internal os of the cervix) and the lower limit of fundus. The site of sampling are outlined schematically in Figure 3.1.

Northern blot was performed as described in Chapter 2. The composition of the pre-hybridisation buffer was freshly de-ionised formamide (50% vol/vol), sodium phosphate (50mM), NaCl (0.8M), sodium dodecyl sulphate (SDS, 2% wt/vol), herring sperm DNA (100µg /ml), yeast transfer RNA (20µg /ml) and 1X Denhardt's (1X = 1% solution of BSA, Ficoll and polyvinylpyrrolidone) and hybridisation buffer was the same with the probe added. Random primed DNA probes were used in all Northern blots described in this chapter.

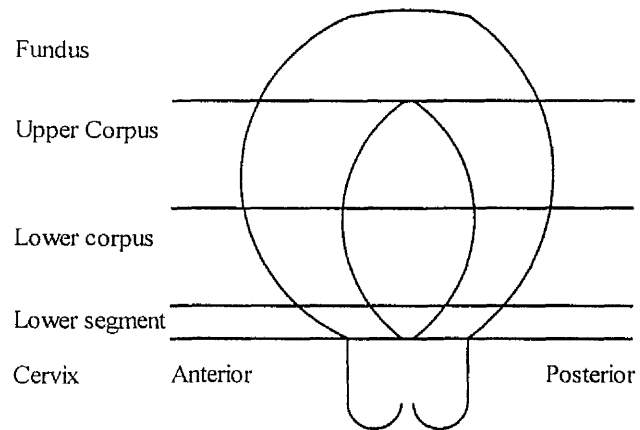


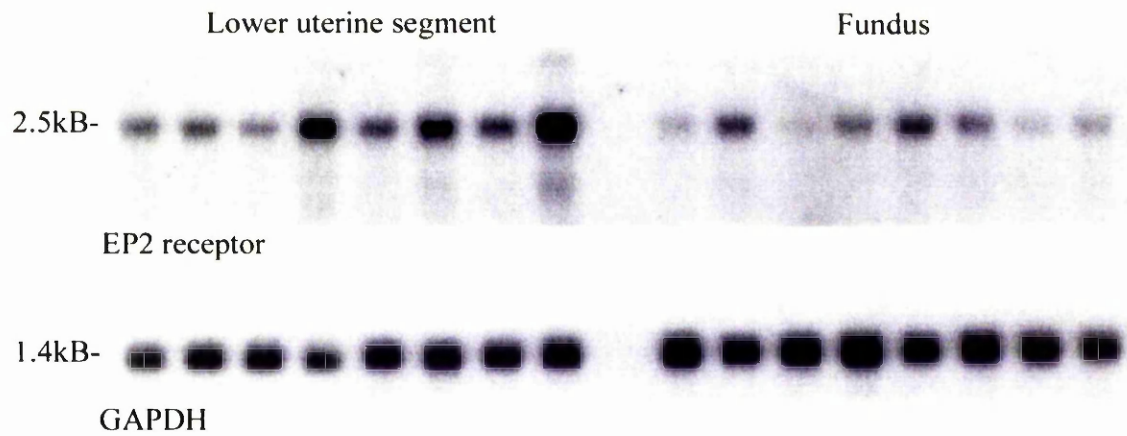
Figure 3.1 Schematic outline of site of sampling from myometrium. The figure represent the appearance of the uterus cut in a longitudinal section in the midline.

3.2 Results.

A preliminary screen for the eight currently recognized prostanoid receptor types and sub-types (EP₁₋₄, DP, IP, TP and FP) using Northern blot analysis and cDNA probes identified EP₂, EP₃, EP₄ and FP receptor mRNA in baboon myometrium. Samples of RNA from LUS and fundus from the 8 hysterectomies were then analysed for these receptors. Compared with fundus, LUS exhibited more EP₂ (Figures 3.2A and 3.3A), less EP₃ (Figures 3.2B and 3.3B) and similar levels of EP₄ and FP mRNA (Figures 3.3C and 3.3D). The corpus also had less EP₂ and more EP₃ mRNA compared with LUS (Figure 3.4). There was no correlation between the level of expression of any of these genes in either LUS or fundus with the gestational age of the animal at the time of Caesarean section.

Northern blot analyses of prostanoid receptor mRNA in myometrium (anterior corpus), cervix, decidua and chorion demonstrated EP₂, EP₃ and EP₄ receptor mRNA in all four tissues. FP receptor mRNA was only detected in myometrium and cervix (Figure 3.5). There was a characteristic distribution of EP receptor mRNA in the four tissues, with the EP₂ receptor being most abundant in cervix, EP₃ most abundant in myometrium and EP₄ most abundant in decidua (Figure 3.6). There was no difference in EP₄ or FP receptor mRNA comparing myometrium and cervix (Figure 3.6). EP₁, IP, DP and TP expression were not reliably detected by Northern blot in any of these tissues using cDNA probes, as were employed in these analyses.

A.



B.

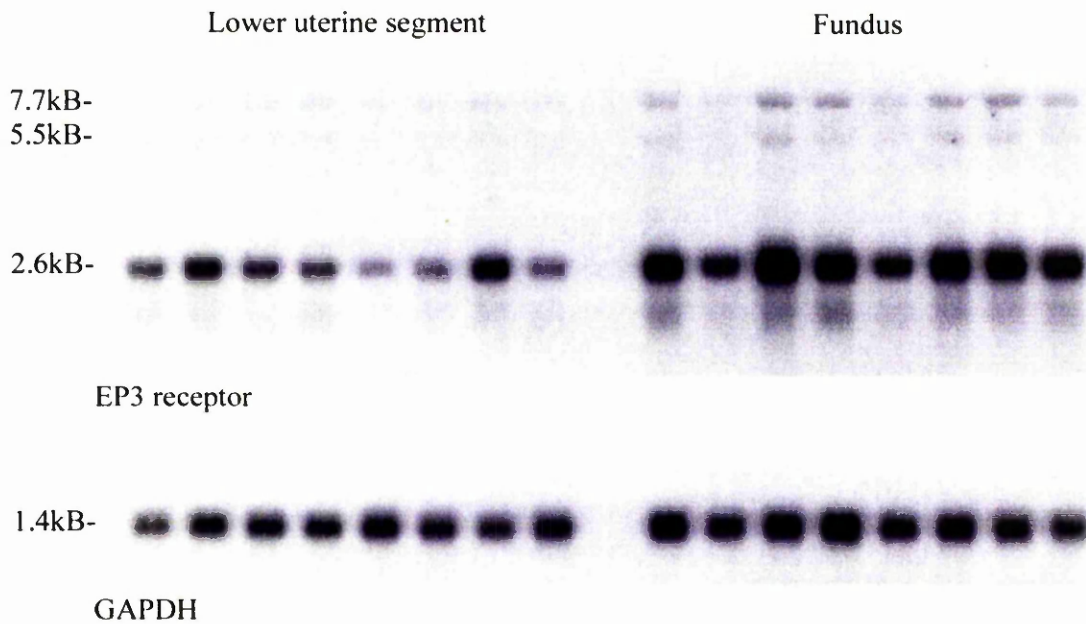


Figure 3.2A-B. Northern blot of EP₂ and EP₃ expression in lower uterine segment (LUS) and fundus from 8 pregnant baboons between 121-180dGA. Variation in loading was corrected by subsequent probing of the same blot for GAPDH. Each lane was loaded with 2μg polyadenylated RNA (EP₃) or 5μg (EP₂).

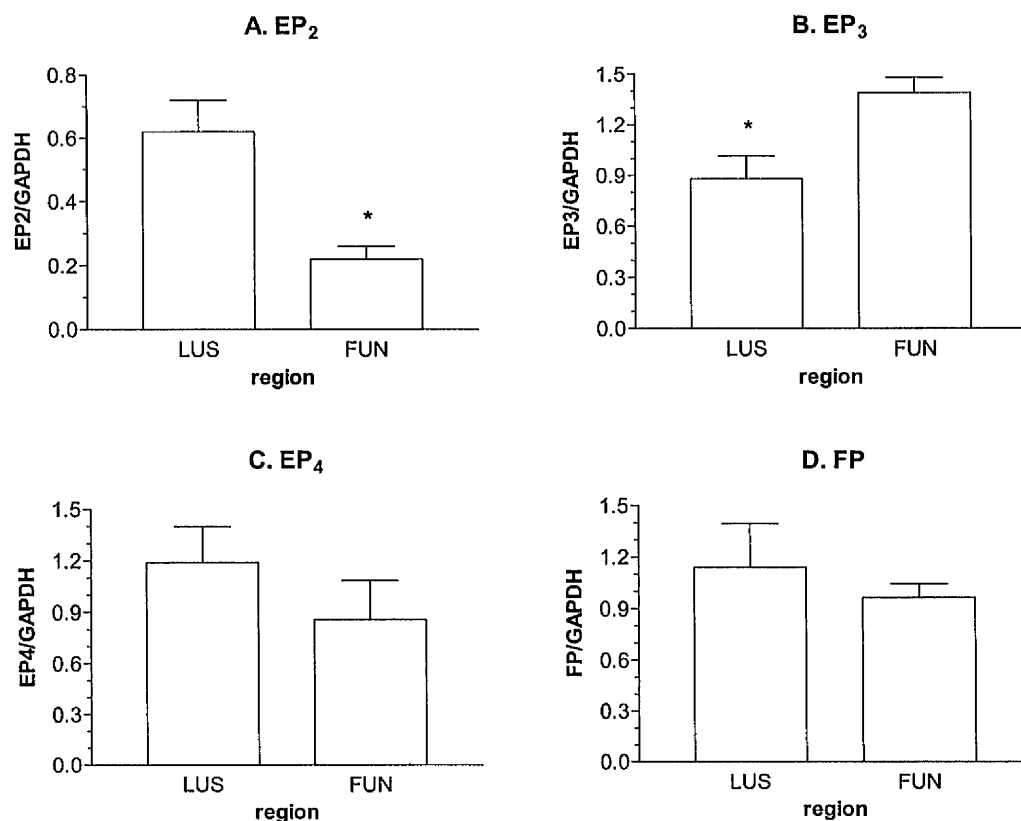


Figure 3.3. Densitometric quantitation of EP₂, EP₃, EP₄ and FP receptor mRNA relative to GAPDH comparing lower uterine segment (LUS) and fundus. Densitometry for EP₃ was performed on the signal of approximately 2.4kB (see Figure 2A). Statistical comparison of LUS vs. fundus (means \pm SEM, n=8) by Student's t-test, *p<0.01.

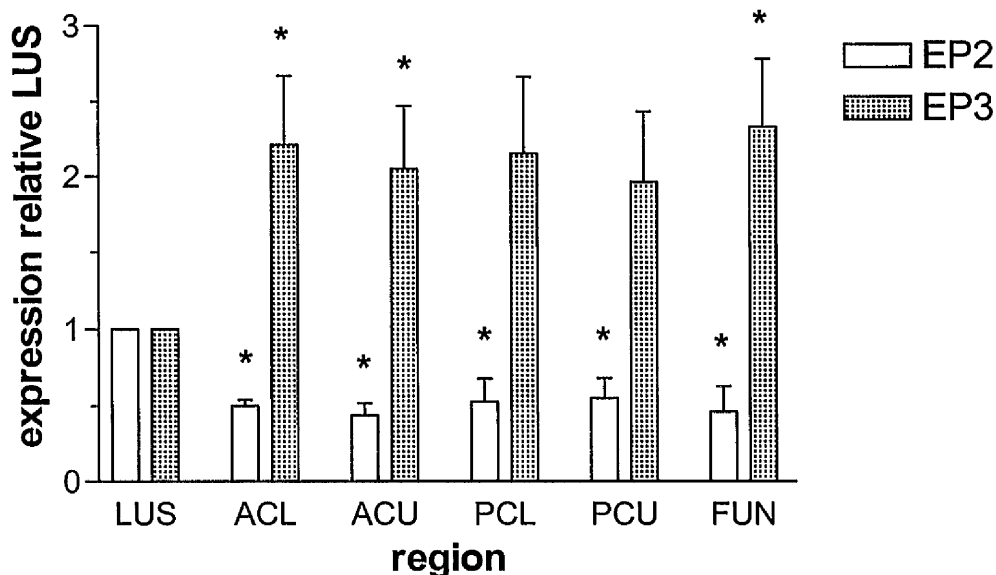


Figure 3.4. Densitometric quantitation of EP₂ and EP₃ mRNA in different regions of baboon uterus using lower uterine segment (LUS) as a reference. Each lane was loaded with 5µg of myometrial mRNA extracted from one of 6 regions of a single animal's uterus: LUS= lower uterine segment, ACL = anterior corpus lower, ACU = anterior corpus upper, PCL = posterior corpus lower, PCU = posterior corpus upper, FUN = fundus. Blots were probed for EP₂ or EP₃ and then for GAPDH. The signal for a given probe was expressed as a ratio to the signal for GAPDH. For a given blot, the signals in the other 5 regions were expressed as a ratio to the signal in LUS (means ± SEM, n=6-7). The data were then pooled and means and 95% CI were calculated for each region with the LUS as unity and a significant difference (*) with LUS was assumed where 95% CI excluded 1.0.

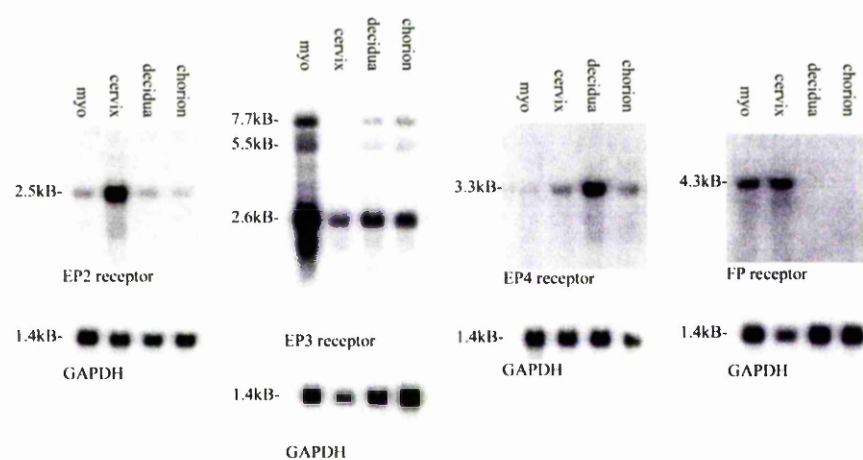


Figure 3.5A-D. Representative Northern blots of EP₂, EP₃, EP₄ and FP receptor mRNA in myometrium from anterior corpus (MYO), cervix (CX), decidua and chorion. Each lane was loaded with 10µg polyadenylated RNA.

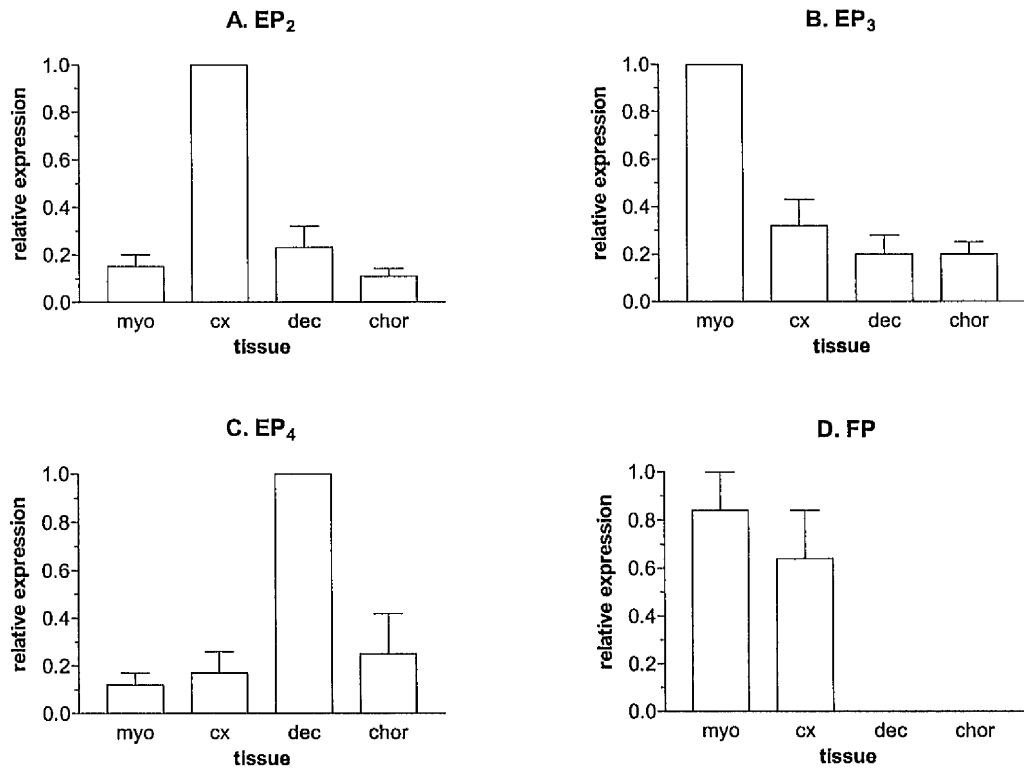


Figure 3.6A-D. Quantitation by densitometry of EP₂, EP₃, EP₄ and FP receptor mRNA in myometrium from anterior corpus (MYO), cervix (CX), decidua (DEC) and chorion (CHOR) as a ratio to GAPDH. In each blot (see Figure 3.5 for representative blots), the highest signal was expressed as unity and the others were expressed as a ratio to it. The ratios for 3-5 blots from different animals were then pooled and the 4 regions compared by ANOVA, plus Tukey's multiple comparisons. EP₂ mRNA expression was significantly greater in the cervix than in the other 3 tissues ($p < 0.05$, $n = 5$), EP₃ mRNA was significantly greater in MYO than in the other 3 tissues ($p < 0.05$, $n = 4$), EP₄ was significantly greater in decidua than in the other tissues ($p < 0.05$, $n = 3$). There was no significant difference in EP₄ or FP ($n = 3$) expression comparing MYO and CX.

3.3 Discussion.

When considering the regulation of myometrial contractility, most attention is focused on factors controlling delivery of the fetus. However, effective post-partum contraction of the uterus is probably equally important from an evolutionary perspective. The processes of delivery of the fetus and post-partum contraction are quite different. Delivery of the fetus requires phasic contraction with a relative relaxation of the lower segment. Post-partum contraction requires tonic smooth muscle activation of the whole uterus. Therefore, a possible means by which differential regulation of uterine contraction (i.e. pre- and post-delivery of the fetus) might be studied is by addressing regional variation in the control of contractility.

One reason why these issues have not been fully addressed is the difficulty in obtaining specimens of healthy pregnant human myometrium from regions other than the LUS. There are some reports of analysis of Caesarean hysterectomy specimens and one report demonstrated regional variation in oxytocin receptors ¹⁹³. However, these reports are anecdotal and are necessarily limited by the opportunities to obtain healthy tissues. There are some studies comparing lower segment strips with biopsies of fundus also obtained at the time of lower segment Caesarean section ¹⁹⁴. However, these did not utilize full thickness strips of fundus which introduces a potential for bias in comparing LUS and fundus.

At the same time as samples were obtained for Northern analysis, paired samples were obtained for contractility studies. It was found that PGE₂ contracted myometrial strips from the fundus but not the lower segment ¹⁹⁵. (These findings are reported in a paper where I was first author, but I have not included them in this thesis. Although I directed these experiments and devised the experimental protocol, the actual practical work was

carried out by a colleague and, therefore, I have excluded these data from this thesis.) The LUS is formed during pregnancy from the cervix ¹⁹⁶. Given the fibrous nature of the cervix, it might be expected that the LUS would exhibit a generalized reduced contractility compared to the fundus. However, its response to $\text{PGF}_{2\alpha}$ was virtually identical to fundus ¹⁹⁵. These observations implied that the lack effect of PGE_2 on LUS was a manifestation of differences in prostanoid signal transduction.

As discussed in Chapter 1, four receptor sub-types have been identified pharmacologically which mediate the effects of PGE_2 ¹¹. EP_1 and EP_3 receptors generally stimulate and EP_2 and EP_4 receptors generally inhibit smooth muscle¹¹. I tested the hypothesis that the reduced response of LUS to PGE_2 compared with fundus was secondary to higher expression of inhibitory EP receptors and/or lower expression of stimulatory EP receptors in LUS. I found that LUS had more inhibitory EP_2 receptor mRNA and less contractile EP_3 receptor mRNA than the fundus, but similar levels of EP_4 mRNA (Figure 3.3). This supported the hypothesis that varying receptor populations explain the different magnitude of the effects of PGE_2 on LUS and fundus. This hypothesis was lent further support as the levels of FP receptor mRNA were similar in LUS and fundus (Figure 3.3), also paralleling the response to native PGs ¹⁹⁵. Samples of myometrium from the corpus showed similar levels of EP_2 and EP_3 mRNA to the fundus (Figure 3.4).

Given that the LUS is formed from the cervix ¹⁹⁶, I hypothesized that the cervix might exhibit similar quantitative differences in prostanoid receptor mRNA when compared with the corpus. EP_2 , EP_3 and EP_4 receptor mRNA were expressed in myometrium, cervix, decidua and chorion (Figure 3.5). There was a characteristic pattern of expression of the sub-types with the EP_2 receptor mRNA most abundant in cervix, EP_3 in

myometrium and EP₄ in decidua. EP₄ and FP receptor expression were similar in myometrium and cervix (Figure 3.6). Therefore, the pattern of prostanoid receptor mRNA comparing the cervix and corpus paralleled the pattern comparing LUS and fundus. I conclude that different proportions of EP₂ and EP₃ receptors in LUS are likely to be due to its cervical origin.

The foregoing assumes that levels of mRNA parallel the amount of functional receptors in the respective tissues. This assumption is supported by the contractility studies ¹⁹⁵. It might be further supported by immunocytochemistry or Western blot to confirm translation of mRNA into protein. However, antibodies against the various EP receptor sub-types are not yet widely available. The size of transcripts observed in the present study was similar to previous reports of Northern blots from the human. As in previous studies in the human, multiple bands were present using the EP₃ probe ¹⁹⁷, whereas discrete bands were observed using the EP₂ ¹⁹⁸, EP₄ ¹⁹⁹ and FP ²⁰⁰ probes. Furthermore, the estimated size of transcripts was similar to these reports of Northern analysis of human RNA. Such variation as was observed (less than 20%) may be related to variation in un-translated regions of the gene which has been previously been inferred by the discrepancy between the size of the full length cDNA and the size of transcripts in Northern analysis ²⁰¹ as well as inevitable variation due to experimental error.

These pharmacological differences in LUS and fundus might explain the differential regional control of the uterus before and following delivery of the fetus. I postulate that high levels of PGE₂ from the fetal membranes during labour ²⁰² stimulate EP₂ receptors in LUS during labour, which would favour selective relaxation of LUS to allow passage of the fetus. Following delivery of the membranes, PGE₂ levels will fall, which would favour stimulation of the EP₃ receptor, as its affinity for PGE₂ is more than an order of

magnitude greater than EP₂ receptor ⁴⁷. Furthermore, the activity of other uterotonic agents with a purely contractile effect on the LUS, such as PGF_{2α}, would stimulate contraction without opposition by EP₂-mediated inhibition.

The complex family of prostanoid receptors in decidua and chorion were also of interest. Prostaglandins have been postulated to mediate the clinically evident relationship between infection and PPRM, but current models invoke an indirect action on the membranes, namely, by stimulating uterine activity which then promotes membrane rupture ⁹⁷. The presence of multiple EP receptor sub-types in chorion implies a direct effect of PGE₂ on the fetal membranes. The presence of EP₂, EP₃ and EP₄ receptors on decidua also suggests that PGE₂ acts directly on the decidua. It was interesting that the EP₄ receptor appeared to have much higher levels of expression in the decidua than in adjacent tissues. All these receptors are coupled to adenylate cyclase, EP₂ and EP₄ positively and EP₃ negatively ¹¹. Establishing the effects of cyclic AMP dependent protein kinases should shed light on the effects of PGE₂ on these tissues.

These data have very clear clinical significance. The high level of EP₂ expression in the cervix (Figure 3.5A) may be important for drug development to induce labour. PGE₂ is widely employed to prime the cervix ⁹⁵. The use of PGE₂ reduces the incidence of prolonged labour, Caesarean and instrumental vaginal delivery ²⁰³. However, uterine hyper-stimulation (probably EP₃ receptor mediated ⁸⁶) is the dose-limiting side effect of vaginal PGE₂ ²⁰⁴. PGE₂ stimulation of glycosaminoglycan synthesis by cervical fibroblasts is mediated by activating adenylate cyclase ²⁰⁵. Given that the EP₂ receptor is coupled positively to adenylate cyclase ¹¹ and EP₂ expression is high in the cervix (Figures 3.5 and 3.6), it is possible that the effect of PGE₂ on the cervix is mediated, at least in part, by the EP₂ receptor. Selective EP₂ agonists (which are currently available ¹³)

may, therefore, be as effective in priming the cervix as PGE₂, but would not be expected to cause uterine hyper-stimulation.

There is preliminary evidence to support the above from randomised controlled trials. There are two reports that misoprostol is more effective in priming the cervix than PGE₂ without increasing the risk of hyper-stimulation ^{206;207}. The affinity of PGE₂ for the EP₃ receptor is 14-fold greater than the EP₂ receptor ⁴⁷. However, the affinity of misoprostol for the EP₃ receptor is less than 4-fold greater than EP₂ ⁴⁷. If the EP₂ receptor mediates cervical priming, it would be predicted that the greater the relative affinity of a drug for the EP₂ receptor compared with the EP₃ receptor, the safer and more effective it will be in priming the cervix. However, this is somewhat at odds with the apparent efficacy of sulprostone, a selective EP₁ and EP₃ agonist, in priming the cervix in early pregnancy ²⁰⁸. Sulprostone is a potent stimulator of myometrial contractility ⁸⁶ and it is possible that selective agonists of different EP receptor sub-types might prime the cervix by different mechanisms of action. The possible role of EP₂ receptor activation in the control of cervical compliance during labour may be inferred by whether the gene is up-regulated or down-regulated in labour. This issue is addressed in a later chapter.

The finding that PGF_{2α} contracted both the LUS and fundus ¹⁹⁵ and that expression of the FP receptor gene was similar in the two regions (Figure 3.3) provide a rational basis for the empirical usefulness of 15-methyl-PGF_{2α} in the treatment of PPH refractory to other uterotonic agents ²⁰⁹.

PG synthesis inhibitors are currently used in the treatment of pre-term labour but contract the fetal ductus arteriosus ²¹⁰. It has been proposed that selective antagonists of prostanoid receptors may be useful in the treatment of pre-term labour and have fewer effects on the fetus than COX inhibitors ^{39;99}. However, the complex pattern of multiple

EP receptor sub-types and FP receptor expression in key intra-uterine tissues controlling parturition (Figure 3.5) may limit the usefulness of a drug acting on a single receptor type or sub-type in the treatment of pre-term labour.

Chapter 4. The effect of labour and gestational age on expression of prostanoid receptor genes in myometrium

It is evident from the human clinical use of exogenous synthetic PGs to induce labour that the response to a standard dose of PGE₂ is highly variable between women, from virtually no effect to excessive stimulation of uterine contraction necessitating immediate delivery by Caesarean section ²¹¹. This observation suggests that the sensitivity of the uterus to PGs might be a factor controlling myometrial contractility at term and in labour. In the preceding chapter it was established that variation in contractile responses to native PGs was paralleled by variation in the expression of prostanoid receptor types and subtypes. In the present chapter, I sought to test the hypothesis that variation in the expression of prostanoid receptors at term and in labour has a role stimulating myometrial contractility in primate parturition. I quantified the expression of prostanoid receptor genes in baboon myometrium obtained from 15 animals in the last third of pregnancy, five of which were in spontaneous labour. Given that I had also previously shown that expression of receptor genes varied comparing different regions of the uterus, I also determined (a) whether regional variation in prostanoid receptor expression persisted in labour and (b) the cellular localization of prostanoid receptor mRNA expression.

4.1 Methods

The general methods were as described in Chapter 2. The animals employed were as follows. Ten animals were not in labour and in the last 1/3 of pregnancy at the following days gestational age (dGA): 121, 128, 141, 153, 159, 162, 162, 177, 177, 180 (term=180-185 days). The cervix was un-effaced and closed in all of these animals. Uterine electromyogram leads had been sited in three of these animals going close to term (the animals delivered after 170 dGA, term is approximately 185dGA). Analysis of EMG traces of the 48 hours preceding surgery revealed that myometrial activity was solely in the contractures mode with no contraction activity. No drugs of any form had been administered to any of the animals in the two weeks preceding surgery. Caesarean hysterectomy was also performed on five animals in spontaneous labour. Of these, four had EMG electrodes sited. These animals had a baseline cervical examination at the time of their first EMG contraction activity and were re-examined when they had a sustained switch from contractures to contractions (>30 minutes). The gestational ages at hysterectomy were 164, 184, 191, 193. The cervical dilations for these animals were 6cm, 3cm, 3cm and 2cm, respectively (the cervix was closed in all four at the baseline examination). In a fifth animal without EMG electrodes, Caesarean section was performed at 172dGA and at the time of the procedure it was found that the cervix was 4cm dilated and fully effaced and a hysterectomy was performed. Samples of myometrium were obtained from the fundus and lower segment, flash frozen in liquid nitrogen and stored at -80C until use. The lower segment and fundus were defined described in Chapter 3.

Primary culture of myometrial cells was established from a sample of uterine fundus obtained from an animal in spontaneous labour at 193 days gestational age. In situ

hybridisation was performed on frozen sections of an animal at 177 days gestation, where hysterectomy was performed prior to the onset of labour, from the uterine fundus.

4.2 Results.

Clear signals of similar molecular weight to the cloned human prostanoid receptors genes were detected in myometrium for the EP₁, EP₂, EP₃, EP₄, FP, IP and TP receptors genes using riboprobes, both from the lower segment and uterine fundus and from animals in labour and not in labour (Figures 4.1 and 4.2). No clear signal above background could be detected using an antisense riboprobe for the human DP receptor. All blots were stripped and re-probed for each of three housekeeping genes, GAPDH, beta-actin and cyclophilin (Figure 4.3).

Densitometric analysis of these signals (expressed as a ratio to beta-actin) demonstrated that the level of expression of genes encoding the EP₄, FP and TP receptors were similar comparing samples obtained from the LUS and the fundus and comparing animals in labour and not in labour (Table 4.1).

The only signal that demonstrated a change in the overall level of expression in labour was EP₂ receptor mRNA, which was markedly decreased in myometrium obtained from animals in labour. Furthermore, expression of this gene was greater in the lower segment, but the magnitude of the difference between the lower segment and fundus diminished in association with labour (Figure 4.4 and Table 4.1). The expression of IP receptor mRNA also differed comparing lower segment and fundus and the magnitude of the difference also significantly differed comparing samples from animals in labour and not in labour (Figure 4.4 and Table 4.1). Expression of mRNA encoding the EP₁ and EP₃ receptors was also significantly different comparing samples from the lower segment and the fundus, but the magnitude of the differences comparing the two regions was not significantly affected by labour (Figure 4.4 and Table 4.1). There were no significant changes in the expression of prostanoid receptor genes related to gestational age among the animals

delivered prior to the onset of labour. All but one of the comparisons which were statistically significant with beta-actin were also significant when the signal was expressed as a ratio to GAPDH or cyclophilin (data not shown). The exception was comparison of the IP receptor signal between the lower segment and fundus. This was significant at $P=0.02$ and 0.0004 with beta-actin and cyclophilin, respectively, but the P value was 0.15 when related to GAPDH.

In situ hybridisation was attempted for the EP_2 , EP_3 , EP_4 and IP receptors. Clear signals above background were only detected for the EP_3 receptor (Figure 4.5A), which is most abundantly expressed in myometrium (Chapter 3). Hybridisation of the EP_3 riboprobe could be prevented by co-incubation with an excess of unlabelled probe (Figure 4.5B). The gene encoding the EP_3 receptor was expressed in myometrial cells (Figure 4.5C), but not in vascular smooth muscle cells of muscular arteries in the uterine wall (Figures 4.5A and 4.5D).

Expression of the genes encoding the prostanoid EP_2 and EP_3 receptors could not be detected in cultured baboon myometrial cells. Visual inspection of the cells demonstrated that they retained the characteristic spindle shape of smooth muscle cells in culture.

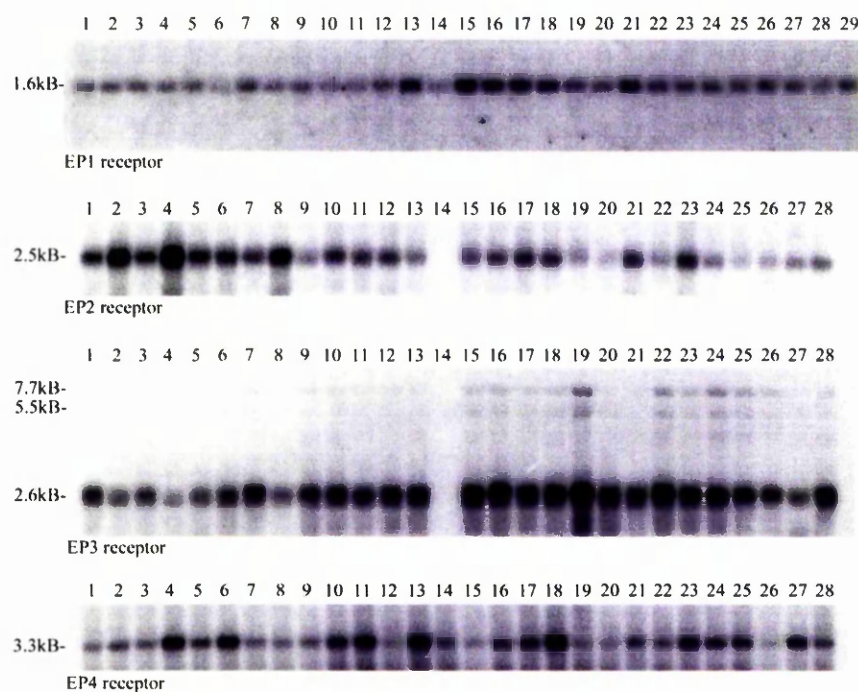


Figure 4.1. Northern blots of EP receptor genes in baboon myometrial mRNA from lower uterine segment (LUS) and fundus (FUN), both not in labour (NIL) and in labour (LAB). 2µg each lane loaded in each lane, probed for all four currently recognized EP receptor genes. Lanes as follows: **EP₁: 1-9 = LUS, NIL; 10-14 = LUS, LAB; 15-24 = FUN, NIL; 25-29 = FUN, LAB. **EP₂** and **EP₃**: 1-8 = LUS, NIL, 9-13 = LUS, LAB, 15-23 = FUN, NIL; 24-28 = FUN, LAB. **EP₄**: 1-9 = LUS, NIL; 10-14 = LUS, LAB; 15-23 = FUN, NIL; 24-28 = FUN, LAB.**

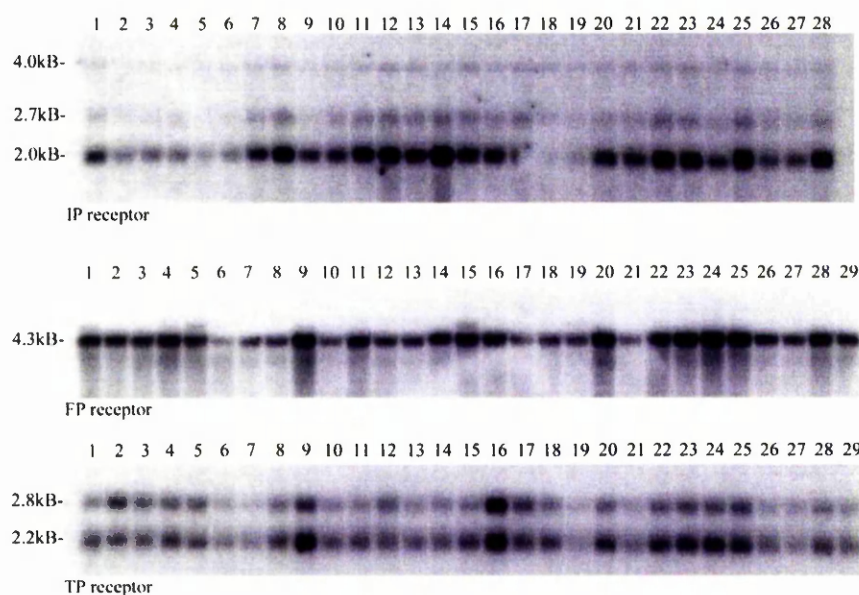


Figure 4.2. Northern blots of non-EP prostanoid receptor genes in baboon myometrial mRNA from lower uterine segment (LUS) and fundus (FUN), both not in labour (NIL) and in labour (LAB). 2µg each lane, probed for the IP, FP and TP receptor genes. Lanes as follows: IP: 1-9 = LUS, NIL; 10-14 = LUS, LAB; 15-23 = FUN, NIL; 24-28 = FUN, LAB. FP and TP: 1-9 = LUS, NIL; 10-14 = LUS, LAB; 15-24 = FUN, NIL; 25-29 = FUN, LAB.

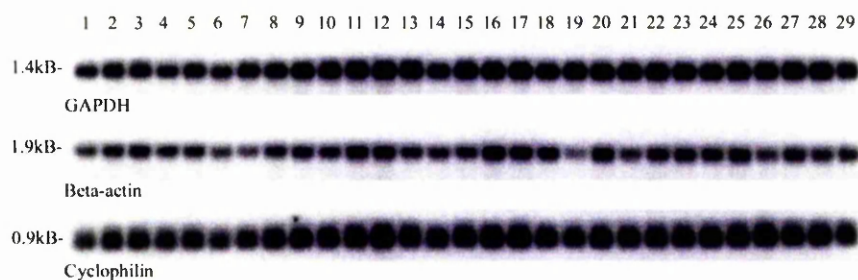


Figure 4.3. Northern blots of housekeeping genes in baboon myometrial mRNA from lower uterine segment (LUS) and fundus (FUN), both not in labour (NIL) and in labour (LAB). 2 μ g each lane consecutively probed and stripped for the GAPDH, beta-actin and cyclophilin genes. Lanes as follows: 1-9 = LUS, NIL; 10-14 = LUS, LAB; 15-24 = FUN, NIL; 25-29 = FUN, LAB.

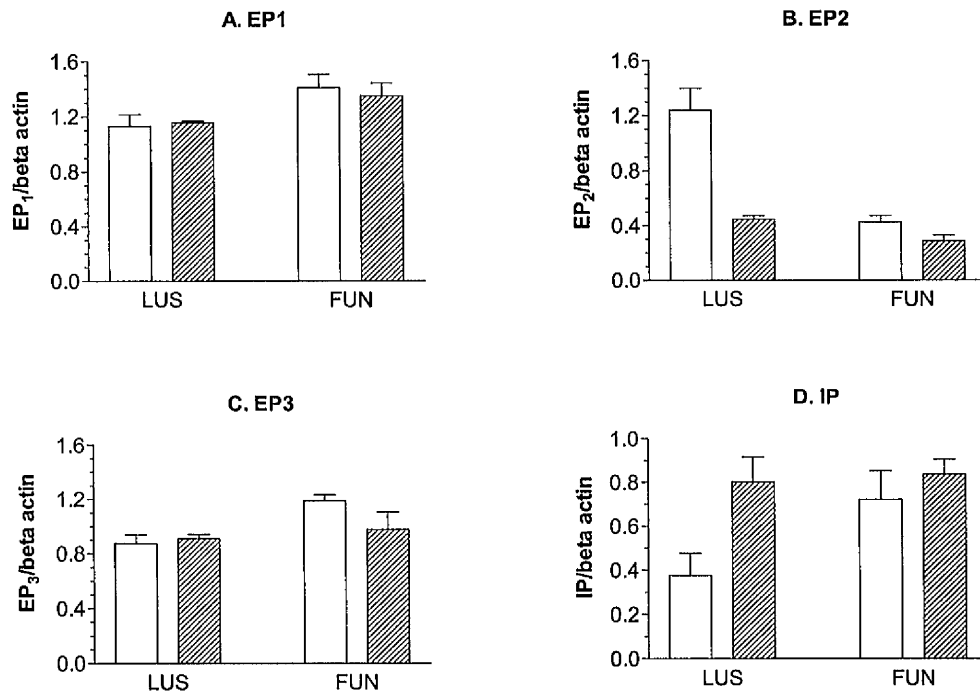


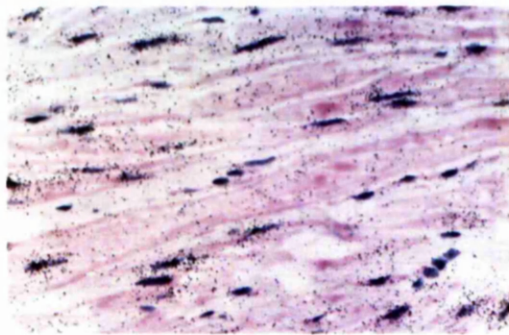
Figure 4.4. Densitometric quantitation of prostanoid receptor signals (as a ratio to beta-actin signal) for prostanoid receptor types in lower uterine segment (LUS) and fundus (FUN) from animals both not in labour (white bars) and in labour (shaded bars). A. EP₁ receptor B. EP₂ receptor C. EP₃ receptor and D. IP receptor. Significant differences between groups, compared with repeated measures ANOVA: effect of labour, EP₂ only; uterine site, EP₁, EP₂, EP₃ and IP; interaction between uterine site and labour, EP₂ and IP only. For details of statistical analysis, see table 4.1.



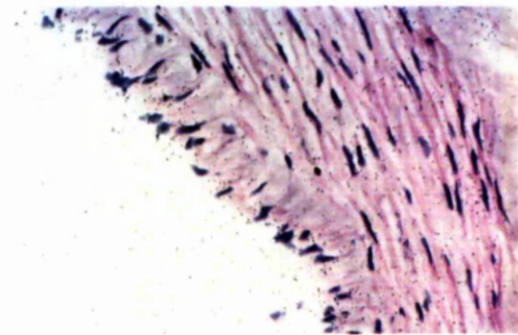
A.



B.



C.



D.

Figure 4.5. In situ hybridisation of EP₃ receptor riboprobe of section from uterine fundus, obtained from animal not in labour at 177 days gestation. A. Low power, dark field, probe without competitor. **B.** Low power, dark field, probe plus unlabelled competitor. **C.** Medium power of myometrial smooth muscle cells. **D.** Medium power vascular smooth muscle cells from muscular artery within uterine wall.

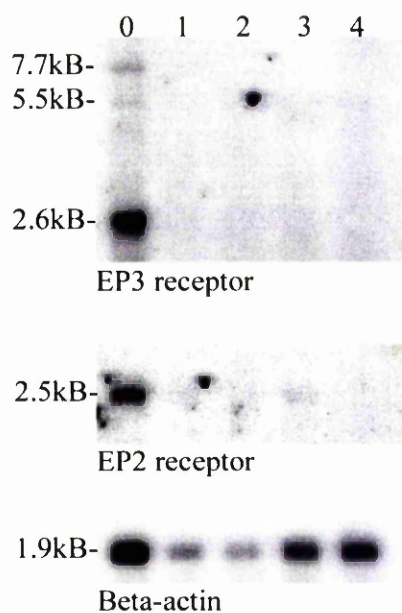


Figure 4.6 Expression of EP₂ and EP₃ receptor genes in fresh and cultured myometrium from uterine fundus of animal in term labour. Lane 0, flash frozen intact fundus, lanes 1-4 first four passages of primary culture of myometrium from same animal. 0.5 µg RNA loaded in each lane.

Table 4.1. Repeated measures analysis of variance of effect of anatomical site and labour on myometrial expression of prostanoïd receptor genes, quantified densitometrically as a ratio to beta-actin.

Gene	Lower segment		Fundus				LAB vs. NIL	LUS vs. Fundus	Interaction
	mean (SEM)		mean (SEM)		P=	P=			
	NIL	LAB	NIL	LAB					
EP ₁	1.13 (0.08)	1.16 (0.13)	1.40 (0.10)	1.35 (0.09)	0.94	0.02	0.57		
EP ₂	1.24 (0.16)	0.45 (0.03)	0.42 (0.05)	0.29 (0.04)	0.0003	0.002	0.02		
EP ₃	0.88 (0.06)	0.91 (0.03)	1.19 (0.04)	0.98 (0.13)	0.23	0.01	0.10		
EP ₄	0.82 (0.12)	0.69 (0.07)	0.63 (0.06)	0.66 (0.10)	0.67	0.17	0.28		
FP	1.88 (0.18)	1.66 (0.20)	1.70 (0.19)	1.45 (0.12)	0.36	0.24	0.89		
IP	0.38 (0.10)	0.80 (0.12)	0.72 (0.13)	0.84 (0.07)	0.12	0.02	0.05		
TP	1.26 (0.12)	1.13 (0.07)	1.23 (0.08)	0.86 (0.08)	0.07	0.13	0.16		

NIL = not in labour, LAB = labour, LUS = lower uterine segment. Not in labour n= 8-10, labour, n=5

NIL = not in labour, LAB = labour, LUS = lower uterine segment. Not in labour n= 8-10, labour, n=5

4.3 Discussion.

There are a number of lines of evidence in a range of species that have indicated a key role for PGs in the control of parturition, and these were reviewed in Chapter 1. Much of the research on the potential role of PGs in the initiation and promotion of labour has focused on factors determining the concentration of endogenous PGs in intra-uterine tissues, in particular the relative balance of the expression and activity of PG synthetic enzymes (COX-1 and COX-2)²¹² and PGDH, which converts PGs to inactive metabolites¹⁴⁸. A number of other studies have looked at signal transduction aspects of myometrial smooth muscle which may affect the response of the uterus to PGs, such as expression of G-proteins²¹³ and genes encoding gap junctions²¹⁴. However, there has been much less research on the role of variation in the expression of prostanoid receptor types and subtypes in the control of myometrial contractility in the human and non-human primate.

The most striking observation of the present study was that labour was associated with a much lower level of expression of the inhibitory prostanoid EP₂ receptor and this was observed both in the fundus and lower segment. Studies of isolated strips of human myometrium have demonstrated a profound inhibitory effect of EP₂ agonists on contractility and a profound excitatory effect of EP₃ receptor activation⁸⁶. Reduced expression of EP₂ receptors would be anticipated to increase the stimulatory effect of a given concentration of PGE₂ on the myometrium acting through EP₁ and EP₃ receptors and, therefore, promote the process of parturition. I hypothesize, therefore, that reduced expression of the inhibitory EP₂ receptor in primate myometrium may have a key role in promoting myometrial contraction during labour. If confirmed in the human, this suggests that an EP₂ receptor antagonist might have a role in the induction of labour and that an EP₂ receptor agonist may have a role in the treatment or prophylaxis of preterm

labour, although the effect of such a drug may be diminished in active labour due to loss of EP₂ receptor expression.

I confirmed the previous observation (Chapter 3) of regional differences in EP₂ and EP₃ receptor gene expression comparing lower segment and fundus and that these differences were maintained in labour. Interestingly, the magnitude of the difference in EP₂ receptor gene expression between the lower segment and fundus was significantly diminished by labour, although a difference persisted.

By optimizing techniques, principally the use of riboprobes, I could also detect EP₁, TP and IP receptor gene expression using Northern analysis, whereas these genes could not be detected using the initial methodologies outlined in Chapter 3. These studies demonstrated regional variation in both the EP₁ and IP receptor genes. In the case of the EP₁ receptor gene, expression was greater in the fundus than the lower segment, but labour had no statistically significant effect on either the absolute level of expression or the difference between the two regions. In the case of the IP receptor gene, expression was lower in the lower segment before labour, but not in tissue obtained during labour. The higher level of expression of the excitatory EP₁ is consistent with the previous observation that strips of myometrium from the fundus exhibited a greater contractile response to PGE₂ than strips from the lower segment¹⁹⁵. The greater level of expression of the inhibitory IP receptor in the lower segment during labour suggests that PGI₂ may inhibit myometrial contractility in the lower segment during labour, but this issue requires further study.

Relating the current observations to previous studies there are interesting parallels and differences. Reduced expression of EP₂ receptor mRNA in myometrium in association with labour (or advancing gestational age) has now been demonstrated in the baboon

(present study), rat ⁹² and human ⁹¹, but not in sheep ²¹⁵. This species difference in genetic expression is paralleled by functional differences in response of myometrial strips, since both human ⁸⁶ and baboon myometrium ⁸⁹ relax in response to EP₂ receptor agonists, whereas these drugs are without effect in ovine myometrium ⁹⁰.

The lack of an effect of labour on expression of the FP receptor gene differs from studies using human LUS which demonstrated increased FP receptor mRNA in biopsies taken from women in labour at the time of Caesarean section, as well as animal studies in the rat ⁹² and sheep ²¹⁵. The contractile response to PGF_{2α} and expression of FP receptor mRNA is the same in myometrial strips from the lower segment and fundus of baboons ¹⁹⁵. I interpreted this observation as possibly indicating a major role for the FP receptor in post-partum contraction of the uterus, consistent with the efficacy of FP agonists in the management of refractory atonic PPH ²⁰⁹. It is possible, therefore, that expression of FP receptor mRNA could increase in the final stages of labour, and that the time of sampling of myometrium with respect to the total duration of labour, may explain this difference.

The only receptor gene that could reliably be localized using in situ hybridisation was the EP₃ receptor, which is the prostanoid receptor most abundantly expressed in myometrium (Chapter 3). As expected from functional studies ⁸⁶, this gene was noted to be highly expressed in myometrial cells (Figures 5A and 5C). Interestingly, the gene did not appear to be expressed in the smooth muscle of small uterine arteries (Figures 5A and 5D). This observation suggests that there may be differential control of myometrial and vascular smooth muscle in the uterus. The blood vessels supplying the uterine muscle would be expected to be dilated most when the uterus was most metabolically active, i.e. during labour. Therefore, uterine vascular smooth muscle should be in a relaxed state when myometrial smooth muscle is in its most active state. On the basis of the in situ

hybridisation findings, differential regulation of myometrial and vascular smooth muscle may be effected by differential expression of prostanoid receptor genes.

Further studies should attempt to confirm that variation in levels of gene expression in the present study are reflected in variation in the number of functional receptors.

Furthermore, elucidation of the factors which control EP₂ receptor gene expression in primate myometrium may shed light on the control of primate myometrial contractility at the molecular level. The loss of expression of the prostanoid EP₂ and EP₃ receptor genes in cultured baboon myometrial cells was disappointing as it was hoped that the factors controlling expression of these genes might have been studied in this model.

Chapter 5. The effect of labour and gestational age on expression of prostanoid receptor genes in cervix

Prostaglandins (PGs) are known to have a key role in the control of cervical compliance in human pregnancy⁹⁵. Appreciation of the physiological role of PGs led to clinical trials of exogenous synthetic PGE₂ as a means of improving cervical compliance prior to induction of labour. A large number of randomized controlled trials have demonstrated that PGE₂ is an effective agent in priming the cervix and that its use prior to induction of labour reduces the risk of failed induction and operative delivery²¹¹. The principal side effect of PGE₂ in this context is uterine hyper-stimulation. Although PGE₂ reduces the overall risk of Caesarean section, the risk is increased in the interval immediately following insertion of PG due to occasional induction of hypertonic uterine activity²¹¹.

In Chapter 3, I demonstrated that the cervix exhibited much greater levels of expression of the adenylate cyclase coupled EP₂ receptor than myometrium, decidua or chorion¹⁹⁵. I hypothesized that the EP₂ receptor may mediate the clinically important effects of PGE₂ on cervical compliance. Characteristically, expression of genes encoding receptors for hormones that have a stimulatory role in parturition is increased in the given tissue in association with labour⁷⁵. This suggests that if any of the prostanoid receptors had a major role in mediating the changes in cervical compliance associated with labour, that there would be an increase in the expression of the gene encoding the receptor. Conversely, if any receptor mediated an inhibitory effect on these changes, it suggests that the expression of the gene may be decreased in labour. In this chapter I report quantitative Northern analysis of all the currently recognized G-protein coupled prostanoid receptor genes in the cervix obtained from pregnant baboons by total

Caesarean hysterectomy, comparing animals in labour and not in labour at the time of surgery.

5.1 Methods.

The same animals described in Chapter 4 were employed and the general methods were as outlined in Chapter 2.

5.2 Results.

Clear signals of similar molecular weight to the human prostanoid EP₁, EP₂, EP₃, EP₄, FP, IP and TP receptors genes were detected using Northern analysis of cervical mRNA (Figures 5.1 and 5.2). Multiple transcripts were observed for the EP₃, IP and TP receptor genes. No clear signal above background could be detected using an antisense riboprobe for the human DP receptor.

There was a significant increase in the level of expression of the EP₁ receptor with advancing gestational age (Figure 5.3 and Table 5.1). There were no gestational age-related changes in the level of expression of any of the other receptor genes (Table 5.1).

There were marked and highly significantly lower levels of expression of the EP₂, FP and TP receptor genes in samples obtained from animals in labour compared with those not in labour (Figure 5.4 and Table 5.2). There was no significant difference in the levels of expression of the genes encoding the EP₁, EP₃, EP₄ or IP receptors comparing animals in labour and not in labour (Table 5.2).

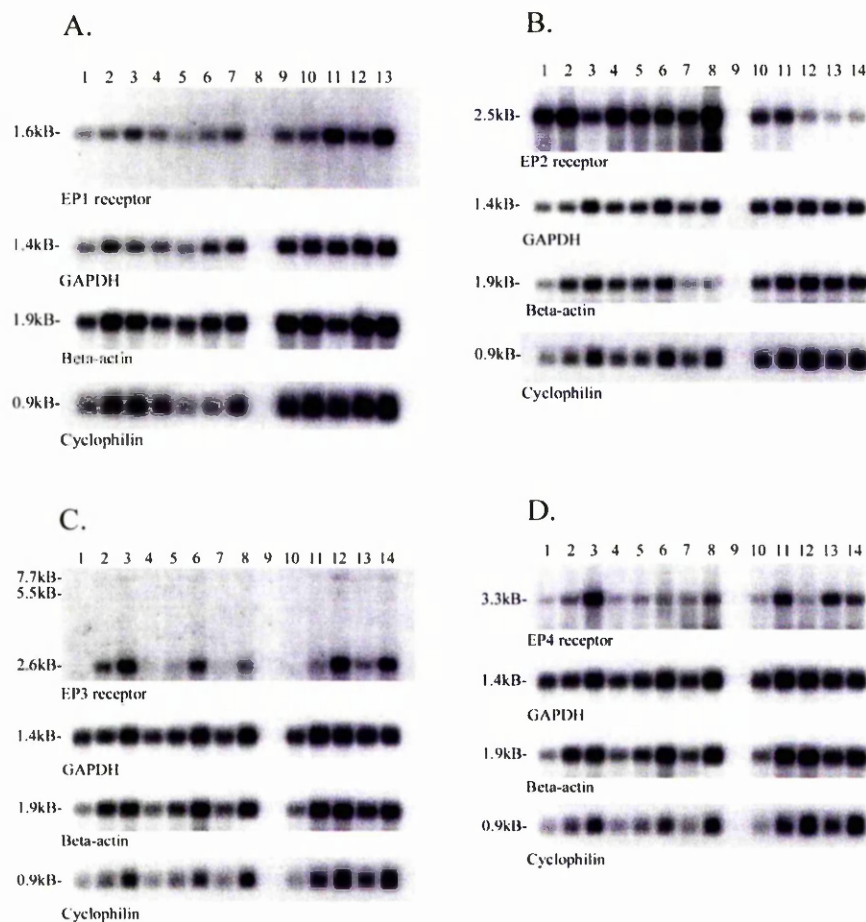


Figure 5.1. Northern blot of prostanoid EP receptor gene sub-types in baboon cervix. A. EP₁ receptor, B. EP₂ receptor, C. EP₃ receptor, D. EP₄ receptor. In each case the membrane was re-probed for each of the three housekeeping genes, GAPDH, beta actin and cyclophilin. The estimated molecular weight of transcripts is given in kilobases (kB) from interpolation of markers as described in Methods. The same quantity of mRNA (either 2 or 4 µg) was loaded in each lane in a given blot. A. Lanes 1-7 not in labour (NIL), lanes 9-13 in labour (LAB). B-D. Lanes 1-8 NIL, lanes 10-14 LAB.

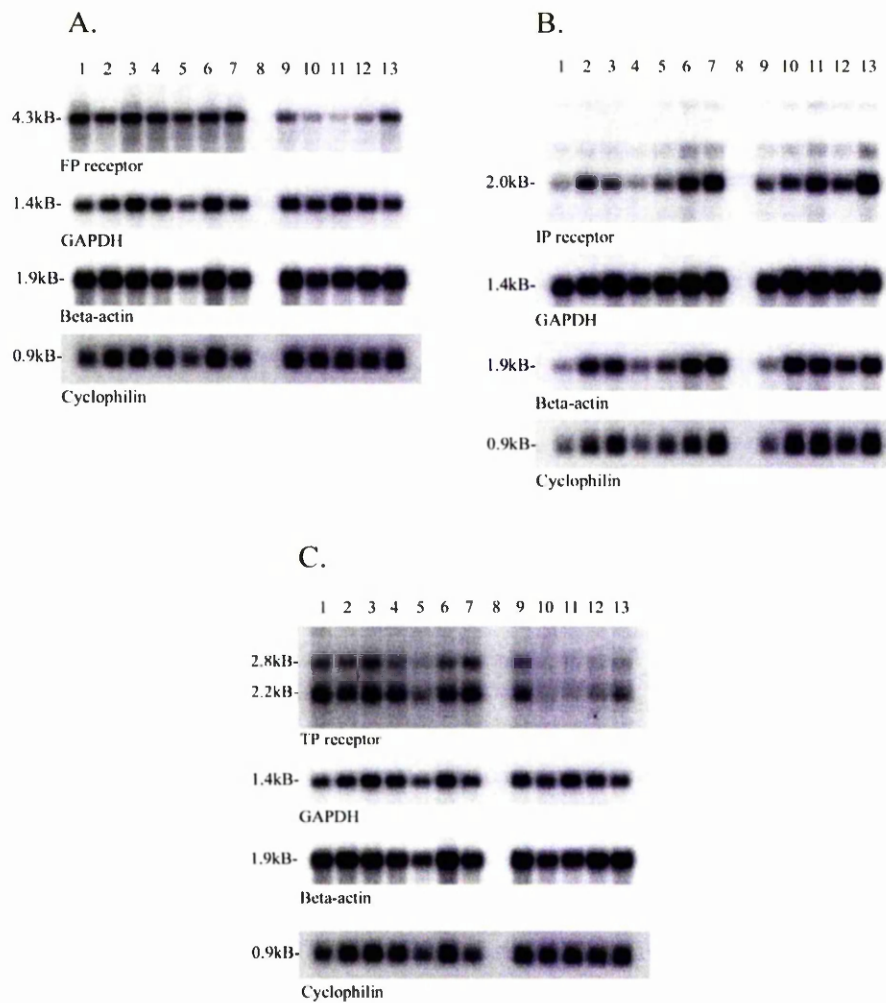


Figure 5.2. Northern blot of non-EP prostanoid receptor gene sub-types in baboon cervix. A. FP receptor, B. IP receptor, C. TP receptor. In each case the membrane was re-probed for each of the three housekeeping genes, GAPDH, beta actin and cyclophilin. The estimated molecular weight of transcripts is given in kilobases (kB) from interpolation of markers as described in Methods. The same quantity of mRNA (either 2 or 4 μ g) was loaded in each lane in a given blot. All blots lanes 1-7 not in labour, lanes 9-13 in labour.

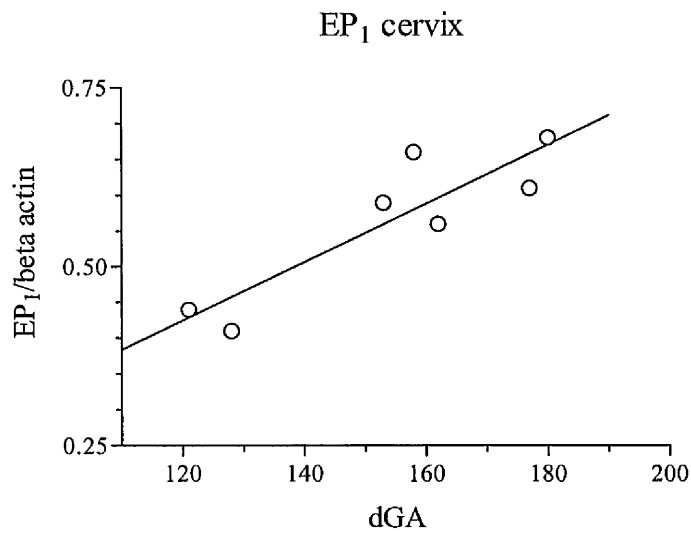


Figure 5.3. Expression of EP₁ receptor mRNA in cervix (n=7) with advancing days of gestational age (dGA). Simple linear regression of EP₁ receptor gene expression (relative to beta-actin, expressed as densitometric ratio) related to gestational age in animals not in labour (n=7). See Table 5.1 for statistical analysis.

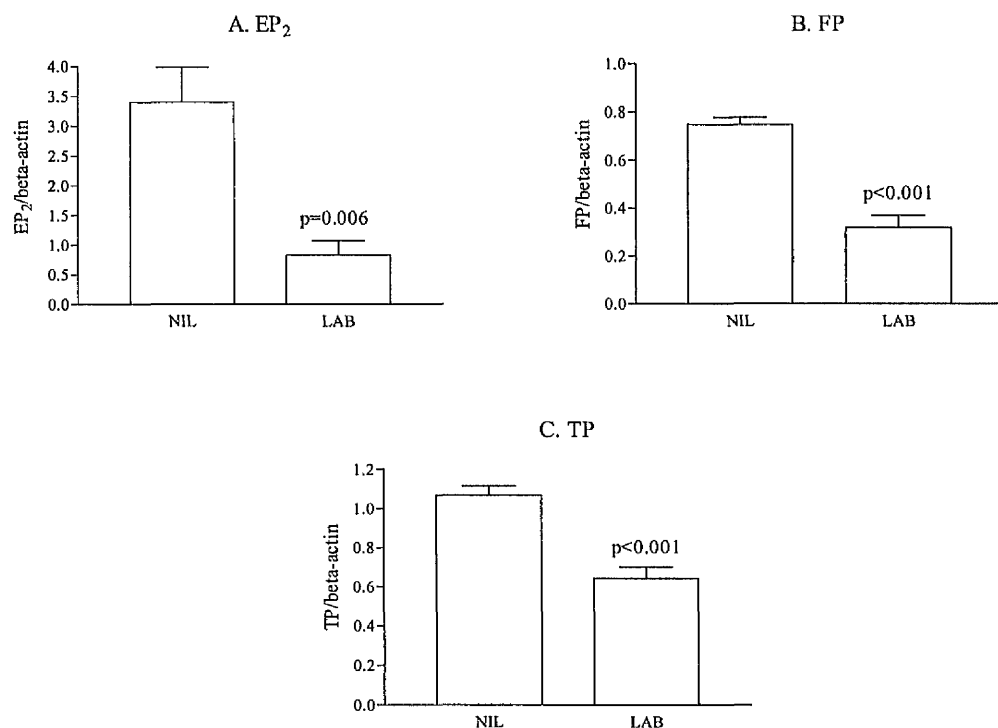


Figure 5.4. Expression of prostanoïd receptor genes relative to beta-actin in cervix of animals in labour (LAB) and not in labour (NIL). A. EP₂ receptor gene (NIL n=8, LAB n=5), B. FP receptor gene (NIL n=7, LAB n=5), C TP receptor gene (NIL n=7, LAB n=5). See Table 5.2 for statistical analysis. Columns are means and bars are SEM.

Table 5.1. Relationship between gestational age and cervical prostanoid receptor gene expression as a ratio of three housekeeping genes.

	GAPDH		Beta-actin		Cyclophilin	
	p=	r ²	p=	r ²	p=	r ²
EP ₁ (n=7)	0.04	0.61	0.007	0.79	0.01	0.76
EP ₂ (n=8)	0.08		0.91		0.03	0.59
EP ₃ (n=8)	0.69		0.94		0.80	
EP ₄ (n=8)	0.56		0.69		0.91	
IP (n=7)	0.15		0.27		0.53	
FP (n=7)	0.22		0.91		0.32	
TP (n=7)	0.18		0.85		0.28	

The p value is given for simple linear regression, where the ratio of the gene of interest to the given housekeeping gene is the dependent variable and gestational age is the independent variable. Virtually identical results were obtained using Spearman's rho. None of the animals were in labour at the time of delivery. The relationship between EP₁ gene expression and gestational age was positive (Figure 5.3).

Table 5.2. Cervical prostanoid receptor gene expression as a ratio of three housekeeping genes comparing animals not in labour and in labour.

	GAPDH				Beta-actin				Cyclophilin			
	NIL (n=7-8)	Labour (n=5)	p=	95% CI difference	NIL (n=7-8)	Labour (n=5)	p=	95% CI difference	NIL (n=7-8)	Labour (n=5)	p=	95% CI difference
	mean (SEM)				mean (SEM)				mean (SEM)			
EP ₁	1.11 (0.71)	1.20 (0.12)	0.49		0.56 (0.04)	0.74 (0.11)	0.12		0.70 (0.08)	0.69 (0.07)	0.91	
EP ₂	2.75 (0.35)	0.80 (0.18)	0.002	0.92-2.98	3.40 (0.57)	0.83 (0.25)	0.006	0.89-4.25	2.69 (0.35)	0.74 (0.20)	0.002	1.16-2.71
EP ₃	1.71 (0.37)	2.13 (0.64)	0.54		1.47 (0.26)	1.63 (0.42)	0.73		2.26 (0.40)	1.99 (0.40)	0.66	
EP ₄	0.64 (0.08)	0.68 (0.10)	0.76		0.57 (0.06)	0.57 (0.08)	0.97		0.86 (0.08)	0.75 (0.13)	0.42	
IP	0.43 (0.06)	0.54 (0.09)	0.30		0.82 (0.06)	0.97 (0.11)	0.23		1.11 (0.09)	1.13 (0.11)	0.86	
FP	1.28 (0.08)	0.56 (0.13)	0.0008	0.38-1.26	0.75 (0.03)	0.32 (0.05)	<0.0001	0.30-0.56	1.02 (0.08)	0.42 (0.08)	0.0003	0.35-0.84
TP	1.83 (0.14)	1.08 (0.14)	0.004	0.30-1.21	1.07 (0.05)	0.64 (0.06)	0.0002	0.26-0.59	1.46 (0.13)	0.83 (0.07)	0.004	0.26-0.99

Means are compared using Student's t test. CI denotes confidence interval, NIL denotes not in labour, GAPDH denotes glyceraldehyde-3-phosphate dehydrogenase, SEM denotes standard error of the mean.

5.3 Discussion.

Administration of PGE₂ is well established as a method for preparing the cervix prior to induction of labour ²¹¹. The identification of multiple sub-types of EP receptor ^{11;13} suggested that there may be a role for selective agonists of EP receptor sub-types in cervical priming. In Chapter 3, I compared the level of expression of EP and non-EP prostanoid receptors in the baboon and had found that there were high levels of expression of the adenylate cyclase-coupled EP₂ receptor in the cervix. I hypothesized that this may be the receptor which mediates the clinically important effects of PGE₂. A significant weakness in the hypothesis was the efficacy of sulprostone in priming the cervix ²⁰⁸ since the drug is virtually devoid of EP₂ receptor agonist activity ¹³.

In this chapter, I report that expression of the gene encoding the EP₂ receptor was actually lower in the cervix of animals that were in labour compared with animals not in labour (Table 5.2 and Figure 5.4A). This suggests that the EP₂ receptor does not mediate the effects of PGE₂ on the cervix which promote dilatation. Consequently, I now hypothesize that the EP₂ receptor might inhibit changes in the cervix which promote dilatation, that loss of EP₂ receptor expression may be a factor which promotes cervical dilatation, and that the effects of PGE₂ on the cervix which promote dilatation are mediated through a receptor other than the EP₂ receptor. The EP₁ receptor is a candidate since its level of expression increased with advancing gestational age (Figure 5.3). However, sulprostone, 16,16 dimethyl-PGE₁ and misoprostol are all effective agents for priming the cervix and all three drugs are agonists of the EP₃ receptor ¹³ whereas misoprostol is a weak EP₁ receptor agonist ¹¹. It seems most likely, therefore, that the EP₃ receptor mediates the effects of PGE₂ which promote dilatation of the cervix. I propose that a marked reduction in the expression of the inhibitory EP₂ receptor during labour

results in an uncovering of EP₃ receptor-mediated effects, promoting changes in cervical compliance and, consequently, cervical dilatation. This hypothesis is supported by the observation that the EP₂ and EP₃ receptors mediate opposing effects on adenylate cyclase¹¹.

A further unexpected finding in the present study was the lower level of expression of the genes encoding the FP and TP receptors in the cervix obtained from animals in labour (Figure 5.4B and C, Table 5.2). PGF_{2α} has been employed clinically as a means of inducing labour, but it requires an approximately 10-fold greater dose than PGE₂ for a given therapeutic effect and this is associated with a much greater level of systemic adverse effects²¹⁶. There are very little data on the possible role of the TP receptor in the peripartum control of the cervix. But the current study suggests that a decrease in FP and TP receptor mediated effects may stimulate changes in the cervix which promote cervical dilatation.

Activation of the prostanoid EP₂ receptor usually stimulates adenylate cyclase¹⁹⁸, whereas both FP and TP receptor activation generally stimulate calcium influx^{67;217}. In many tissues, including myometrium, activation of these receptors results in opposing effects⁸⁶. It seems paradoxical, therefore, that receptors which mediate opposing effects might exhibit parallel changes in their level of expression in association with labour. The physiological significance of an increase or decrease in the overall level of expression of a given gene in the whole tissue depends on the cell type(s) that express the gene, the second messenger that the receptor is coupled to within the given cell type, the effect of activation of the given second messenger on the activity of the cell, and the role of that cell type in promoting cervical dilatation. I hypothesize that the parallel levels of expression of the EP₂, FP and TP receptor genes in labour might reflect changes in the

level of expression of these genes in different cells types with different roles in promoting cervical dilatation. I attempted to identify the cell types expressing prostanoid receptor genes in the cervix using in situ hybridization, but could not reliably detect signals above background due to diffuse low levels of expression.

The foregoing assumes that there is a correlation between expression of the gene encoding the receptor and the amount of functional receptor present. This assumption is supported by the data presented in Chapter 3 which demonstrated parallels between regional variation in the contractile response to PGs ¹⁹⁵ and the expression of genes encoding prostanoid receptor genes in baboon myometrium. Ideally, however, the findings of our present study should be confirmed by other techniques such as radioligand binding or Western blot analysis. However, radioligand binding has limitations as a technique to characterize prostanoid receptor populations due to the lack of truly selective ligands ¹³, a problem underlined by studies using cells expressing recombinant prostanoid receptors which have demonstrated that many ligands which are supposed to be highly selective for a given receptor or receptor sub-type bind extensively to other receptors ⁴⁷. The use of Western blot analysis is limited by the fact that antibodies for all the receptor types and sub-types are not yet widely available. The size of bands observed were similar to those observed in myometrium (Chapter 4) which were themselves similar to previous studies using human tissues.

Assuming that the current findings can be extrapolated to the human, these data have implications for drug development. First, it has previously been demonstrated that the major effects of PGE₂ on human myometrium are mediated through the EP₃ receptor ⁸⁶. If, as seems likely on the basis of our studies in the baboon, the EP₃ receptor is also the major effector of PGE₂ in the cervix, it raises the possibility that an EP₃ receptor

antagonist might be an effective uterine tocolytic, which would also have an inhibitory effect on cervical compliance. Such a drug would have the advantage over COX inhibitors that it would not attenuate the inhibitory effects of EP₂ receptor activation⁸⁶. It would also not be expected to contract the fetal ductus arteriosus, the dose limiting side effect of indomethacin²¹⁰, since this is mediated by loss of PGE₂ acting through the EP₄ receptor⁹⁹. The present observations also suggest a possible role for an EP₂ receptor antagonist in the induction of labour. This is supported by the finding reported in Chapter 4 that expression of the EP₂ receptor gene is also reduced in myometrium in association with labour. Similarly, in human myometrium, decreased EP₂ receptor gene expression has been described with advancing gestational age⁹¹. Finally, the previous demonstration of a profound inhibitory effect of EP₂ receptor activation on myometrial contractility⁸⁶ and the current proposal that the same receptor might exert inhibitory effects on cervical dilatation suggest that an EP₂ agonist may have a role in the management of preterm labour.

Chapter 6. The effect of labour and gestational age on expression of prostanoid receptor genes in decidua and chorion

The fetal membranes and decidua are thought to have a key role in the control of parturition in the human and non-human primate. In analyses of cases of preterm labour, approximately 30-40% of births are associated with PPROM ¹⁴⁵. Furthermore, the membranes are thought to have an important paracrine role in the control of myometrial contractility, both in spontaneous term labour ²¹⁸ and in preterm labour secondary to intra-uterine infection ⁹⁷. The decidua is also thought to have an important paracrine role in the control of the myometrium being both a source of PGs and other important regulators, such as oxytocin ¹¹¹.

The amnion is thought to be a critical site of PG synthesis in relation to parturition in a number of species, including the human ²¹⁸. The chorion also expresses PG synthetic enzymes, although a key role has been postulated for the chorion by acting as a barrier for diffusion of PGs from the amnion to the myometrium due to high levels of expression of PG dehydrogenase (PGDH): expression of PGDH falls in association with labour ^{148;219}, implicating a role for this barrier function in maintaining uterine quiescence.

Currently, most models for the role of PGs in the fetal membranes and decidua consider the membranes as sources of either PG biosynthesis or sites of PG catabolism ⁵. Furthermore, any effects of PGs from the fetal membranes and decidua are thought to be paracrine, mediated through changes in myometrial contraction ⁹⁷. However, in Chapter 3, I demonstrated a complex pattern of expression of genes encoding prostanoid receptors in the chorion and decidua of pregnant baboons which suggests that PGs synthesized in the membranes and decidua may also have autocrine effects.

I hypothesize that PGs may have direct effects on decidua and chorion which in turn may have a role in the initiation of parturition in primates. Characteristically, genes encoding signal transduction proteins of systems which have a role in labour are expressed at higher levels in tissues (both myometrial and non-myometrial) from animals which are in labour compared with animals not in labour, and there are many examples of such phenomena^{25;113;139;214}. As an initial test of the hypothesis that direct effect of PGs on decidua and chorion may have a role in the initiation of parturition, I sought to determine whether expression of prostanoid receptor genes varied according to gestational age and whether the animal was in labour or not in labour at the time the tissue was obtained.

6.1 Methods

The same animals described in Chapter 4 were employed and the general methods were as outlined in Chapter 2.

6.2 Results.

Clear signals of similar molecular weight to the cloned human prostanoid receptors genes were detected in decidua to probes for the EP₁, EP₂, EP₃, EP₄, FP, IP receptors (Figures 6.1 and 6.2). No clear signal above background could be detected using an antisense riboprobe for the human DP receptor. Clear signals of similar molecular weight to the cloned human prostanoid receptors genes were detected in chorion to probes for the EP₁, EP₂, EP₃, EP₄, and IP receptors (Figures 6.3 and 6.4). No clear signal above background could be detected using antisense riboprobes for the human DP or FP receptor genes. Weak signals could be detected in both tissues to a TP receptor probe (Figures 6.2 and 6.4). However, the signal to noise ratio for the TP receptor gene transcript was too low in both tissues for meaningful quantitative analysis.

There was a statistically significant negative correlation between the level of expression of the EP₁ and FP receptor genes in decidua and EP₄ expression in the chorion and the gestational age at the time of hysterectomy among animals which were not in labour (Figure 6.5). There were no other significant relationships between gene expression and gestational age in either tissue (Table 6.1). Results of similar statistical significance were obtained when correlation was tested using Spearman's rho (data not shown).

There was a significantly lower level of expression of the EP₂ receptor gene in decidua from animals in labour (Figure 6.6A) but a higher level of expression of the IP receptor gene in decidua (Figure 6.6B) and chorion (Figure 6.6C) from animals in labour. There were no other significant differences in the level of expression of the other prostanoid receptor genes in either tissue comparing samples from animals in labour and not in labour (Table 6.1).

For the signals which were observed to vary with gestational age (EP₁ and FP in decidua and EP₄ in chorion) a further comparison was made of tissues obtained from animals in labour with animals not in labour, where the comparison was adjusted for gestational age using analysis of covariance. In all three cases there was still no significant change in levels of expression associated with labour after adjusting for the effect of gestational age (EP₁ decidua, $P = 0.65$; FP decidua, $P = 0.95$; EP₄ chorion, $P = 0.65$).

All the above comparisons were made expressing the level of the given gene as a ratio to beta-actin. Significant results were also observed when levels were expressed relative to one of the other housekeeping genes, with one exception. There was a negative correlation of borderline statistical significance between gestational age and EP₁ expression in chorion relative to beta-actin ($P = 0.05$). This association was not statistically significant when the EP₁ signal was related either to GAPDH ($P = 0.87$) or cyclophilin ($P = 0.11$).

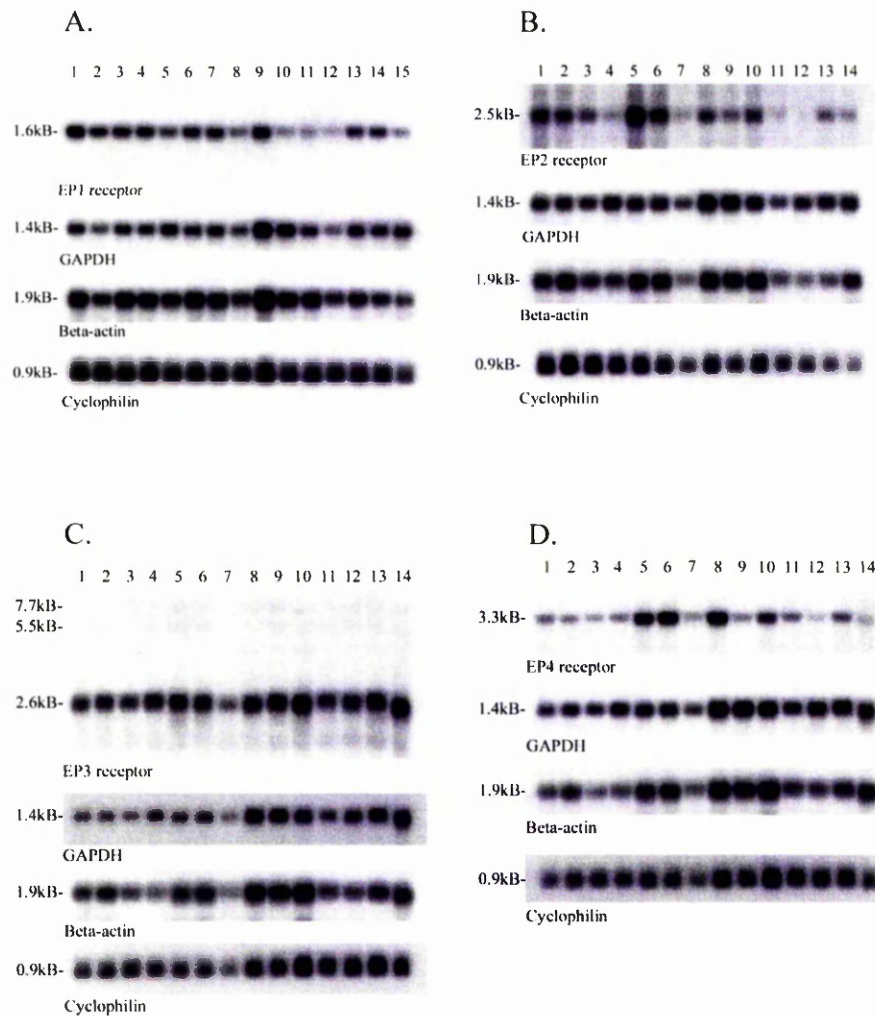


Figure 6.1. Northern blot of EP receptor mRNA in decidua. Either 2 or 4 μ g each lane, same quantity loaded in each lane in given blot) probed for A. EP₁ receptor gene, B. EP₂ receptor gene, C. EP₃ receptor gene, D. EP₄ receptor gene. All membranes consecutively stripped and re-probed for three housekeeping genes. Estimate size of transcripts estimated in kilobases (kB).

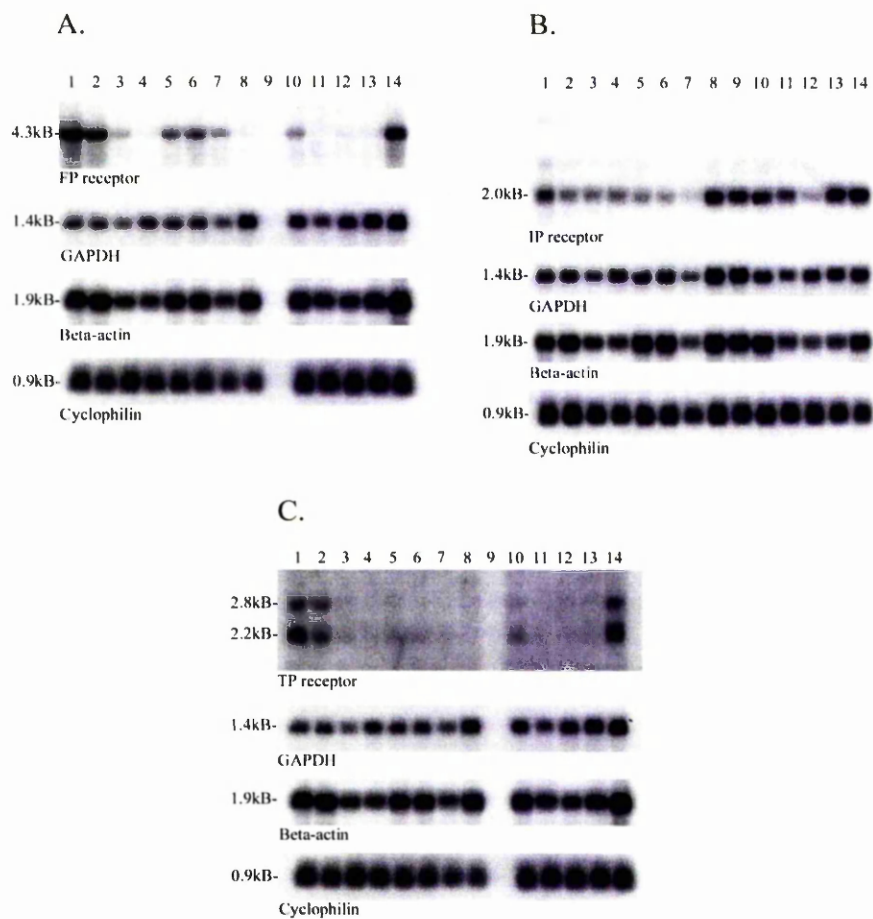


Figure 6.2. Northern blot of non-EP prostanoid receptor mRNA in decidua. Either 2 or 4 μ g each lane, same quantity loaded in each lane in given blot, probed for A. FP receptor gene, B. IP receptor gene, C. TP receptor gene. All membranes consecutively stripped and re-probed for three housekeeping genes. Estimate size of transcripts estimated in kilobases (kB).

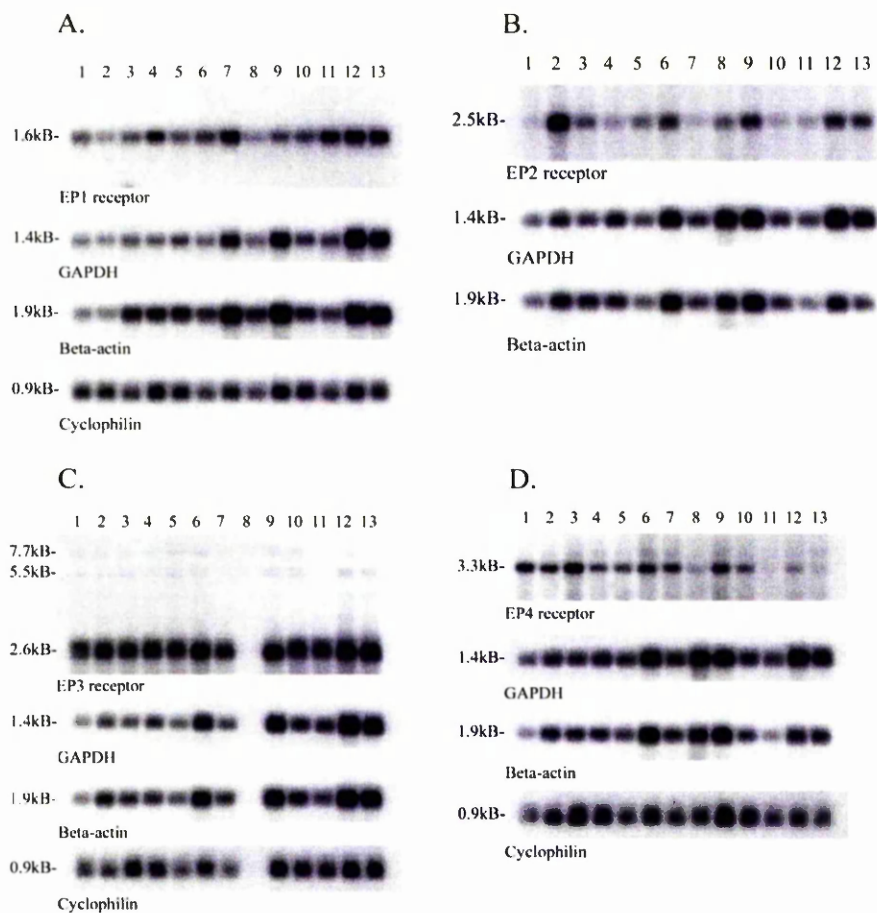


Figure 6.3. Northern blot of EP receptor mRNA in chorion. Either 2 or 4 μ g each lane, same quantity loaded in each lane in given blot, probed for A. EP₁ receptor gene, B. EP₂ receptor gene, C. EP₃ receptor gene, D. EP₄ receptor gene. All membranes consecutively stripped and re-probed for three housekeeping genes. Estimate size of transcripts estimated in kilobases (kB).

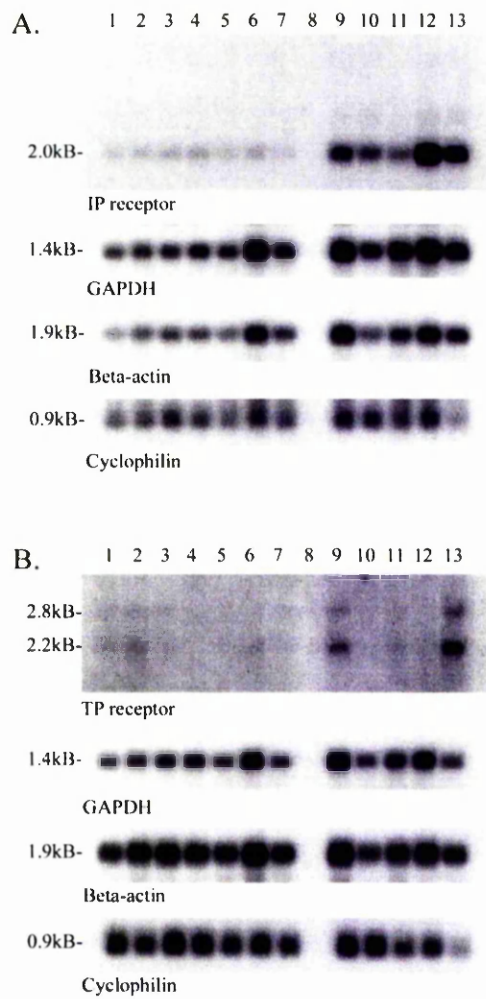


Figure 6.4. Northern blot of non-EP prostanoid receptor mRNA in chorion. Either 2 or 4 μ g each lane, same quantity loaded in each lane in given blot, probed for A. IP receptor gene, and B. TP receptor gene. All membranes consecutively stripped and re-probed for three housekeeping genes. Estimate size of transcripts estimated in kilobases (kB).

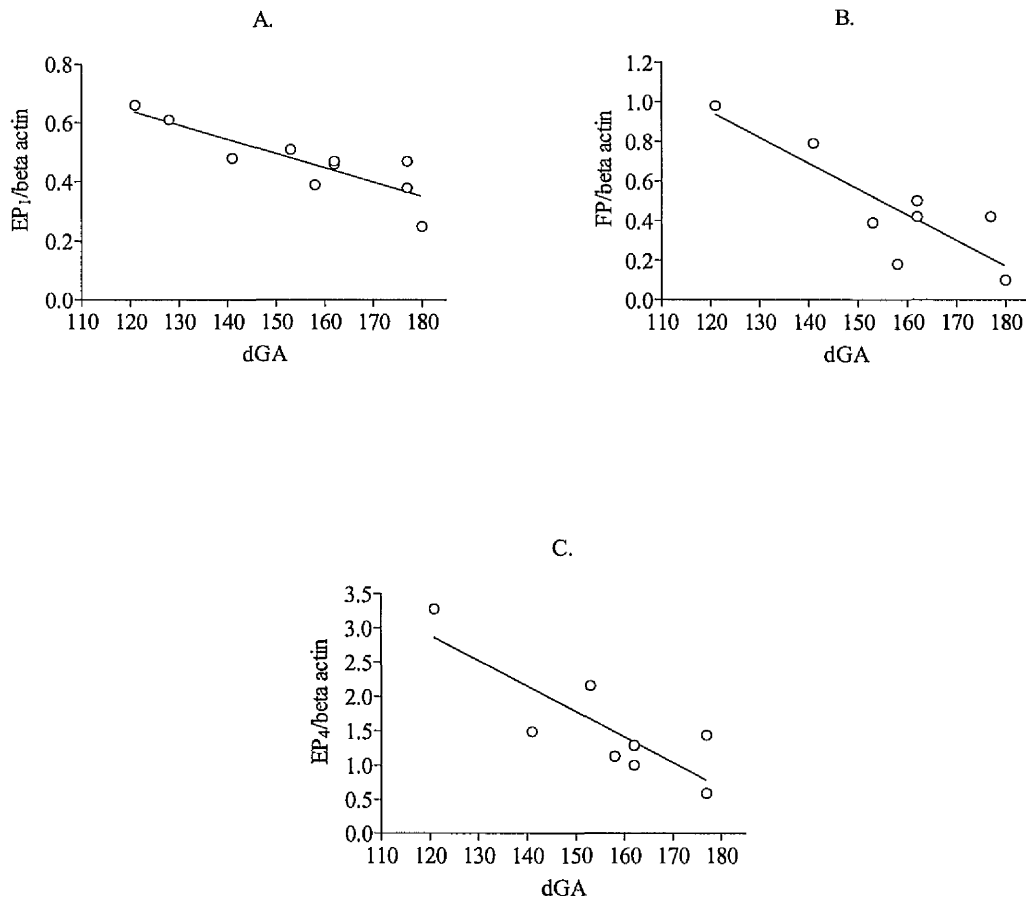


Figure 6.5. Prostanoid receptor gene expression related to gestational age among animals not in labour in decidua and chorion. A. EP₁ receptor gene in decidua ($r^2 = 0.7$, $P=0.002$), B. FP receptor gene in decidua ($r^2 = 0.7$, $P=0.008$), and C. EP₄ receptor gene in chorion ($r^2 = 0.7$, $P=0.01$). Signals quantified using densitometry and expressed as a ratio (arbitrary units) to signal for beta-actin.

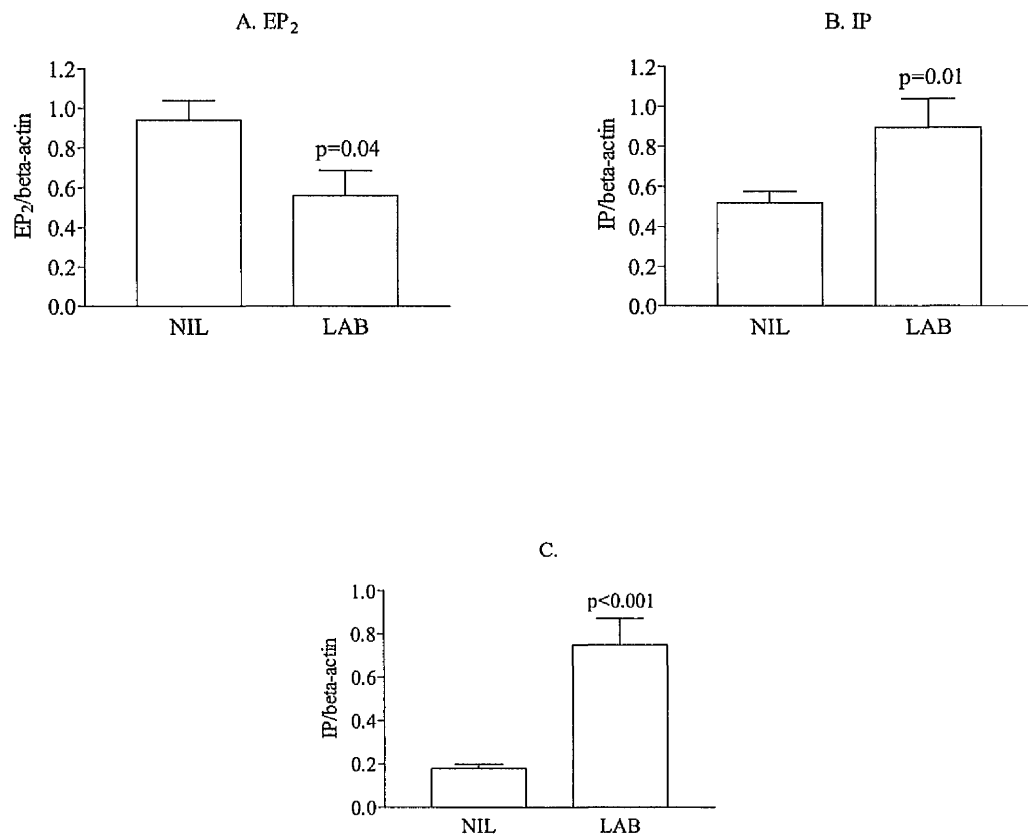


Figure 6.6. Variation in expression of prostanoid receptor genes (expressed as ratio to beta-actin) in relation to labour. A. EP₂ receptor gene in decidua, B. IP receptor gene in decidua, C. IP receptor gene in chorion. NIL = not in labour (n=8-10), LAB = labour (n=5). Statistical comparison by Student's t test. Columns are means, bars are SEM.

Table 6.1. Variation in expression of prostanoid receptor genes (relative to beta-actin) related to advancing gestational age and labour.

Gene	Decidua					Chorion				
	GA	NIL	LAB	NIL vs. LAB	GA	NIL	LAB	NIL vs. LAB	P=	P=
	P (r ²)¶	mean (SEM)	mean (SEM)	P=	P (r ²)¶	mean (SEM)	mean (SEM)	P=		
EP ₁	0.002 (0.73)	0.47 (0.04)	0.37 (0.06)	0.17	0.05 (0.51)	0.85 (0.08)	0.89 (0.15)	0.79		
EP ₂	0.17	0.94 (0.10)	0.56 (0.13)	0.04	0.32	0.42 (0.09)	0.43 (0.08)	0.91		
EP ₃	0.99	1.25 (0.09)	1.36 (0.07)	0.42	0.08	2.21 (0.28)	1.45 (0.13)	0.06		
EP ₄	0.11	0.98 (0.09)	1.18 (0.23)	0.35	0.01 (0.68)	1.55 (0.29)	0.95 (0.14)	0.16		
IP	0.94	0.52 (0.06)	0.89 (0.14)	0.01	0.08	0.18 (0.02)	0.75 (0.12)	0.0003		
FP	0.008 (0.72)	0.47 (0.10)	0.26 (0.09)	0.19	—	—	—	—		

Where GA denotes gestational age, NIL denotes not in labour, LAB denotes labour, and SEM denotes standard error of the mean.

¶Correlation co-efficient: gestational age independent variable, prostanoid receptor gene/beta-actin dependent variable, all animals not in labour at time of hysterectomy.

6.3 Discussion.

Studies on the control of parturition have naturally tended to focus on the myometrium, since uterine contraction is clearly central to the process of expulsion of the fetus. However, 30-40% of preterm births follow PPROM¹⁴⁵. The relative ease of study of the effect of drugs and hormones on myometrium has probably increased the focus on this tissue. However, given the direct contribution of membrane integrity to the process of preterm delivery, and the secondary paracrine effects of choriodecidual activation on myometrial contractility⁹⁷, it is possible that drugs that directly target the membranes and decidua may be effective in the treatment or prophylaxis of preterm labour. Indeed, it is likely, that the apparent efficacy of indomethacin in delaying delivery may be due in a major part to its effects on PG synthesis in the fetal membranes and decidua.

Consequently, it is important to understand the regulation of function of the fetal membranes and decidua. In Chapter 3, I demonstrated that there is a complex pattern of expression of prostanoid receptor genes in baboon chorion and decidua. The genes expressed in chorion and decidua were also expressed in myometrium and cervix. Although receptors are often broadly described as inhibitory or stimulatory, these generalizations usually refer to the effects of receptor activation on smooth muscle. However, it does not follow that a receptor which usually mediates an inhibitory response on smooth muscle will always tend to inhibit non-smooth muscle tissues. A good example of this of the effect of beta-2 adrenoceptor agonists on different excitable tissues: these drugs typically inhibit myometrial and vascular smooth muscle contractility²²⁰, whereas they increase myocardial contractility²²¹. The factors determining the effect of an agonist on a tissue include the cell types in the tissue, the receptors expressed on the different cell types, the second messenger to which the receptor is coupled, the nature of the coupling between the receptor and second messenger, the effect of activation (or

inhibition) of the second messenger on the activity of the cell type, and the role of the given cell type in the regulation of the tissue. Clearly, it does not follow that an agonist which inhibits myometrial contractility will also inhibit the labour-promoting activities of all other intra-uterine tissues where the given signal transduction system is active. A good recent example of this are nitric oxide donors which inhibit myometrial contractility¹⁷¹ but actually promote cervical dilatation¹⁷⁴.

The complex pattern of expression of prostanoid receptors in myometrium (Chapter 4) and the widespread clinical use of prostanoids and their synthesis inhibitors in the therapeutic control of myometrial contractility suggest possible roles for drugs with selective activity at prostanoid receptors in the clinical manipulation of the uterus in term and preterm labour. The demonstration that the genes encoding these receptors are expressed in the chorion and decidua suggested that these receptors might have a role in the control of these key tissues in labour. As stated above, it cannot be safely assumed that receptors which typically inhibit myometrium would also exert inhibitory effects on the chorion and decidua. In order to understand the effects of drugs with selective activity on prostanoid receptors in the control of labour, their effects on non-myometrial tissues also need to be resolved. One way of drawing inferences about the effect of a given system in the control of parturition is by studying the expression of its component parts in pregnancy and during labour. This approach has demonstrated (for example) increased expression in labour of genes encoding gap junctions in myometrium^{179;214}, COX-2 in amnion²⁵ and nitric oxide synthase in cervix²²². There is evidence in each of these cases that the increased expression of the given gene is involved in the control of labour. Furthermore, the realization of the profound increase in COX-2 gene expression has led to the successful exploration of selective inhibitors of this enzyme in the management of preterm labour²²³ and the recognition of the possible role of nitric oxide in the control of

the cervix led to successful trials of NO donors to prime the cervix clinically ^{174;224}. Therefore, as an initial investigation of the possible role of prostanoid receptors in the control of the primate chorion and decidua, I sought to determine whether expression of these genes varied in these tissues with advancing gestational age or in association with spontaneous term labour.

Four patterns of change in expression of prostanoid receptor genes in the decidua and chorion were observed. The first pattern observed was a decrease in expression of genes encoding receptors which are positively coupled to the phosphatidyl inositol cascade (EP₁ and FP) in decidua with advancing gestational age (Figure 6.5A and B). The parallel decrease in these receptors with advancing gestational age suggests that a prostanoid mediated response acting through the phosphatidyl inositol cascade might inhibit changes in the decidua which promote labour. However, activation of these receptors increases myometrial contractility ⁸⁶ and, in the cervix, EP₁ receptor gene expression increases with advancing gestational age (Chapter 5). Therefore, these data suggest that EP₁ and FP receptor-mediated effects may promote parturition in some tissues and inhibit the process in others.

The second pattern that emerged was a decrease in the expression of a gene which encodes a receptor (EP₄) which is positively coupled to adenylate cyclase in chorion with advancing gestational age (Figure 6.5C). This suggests that an adenylate cyclase mediated effect may inhibit changes in chorion which promote parturition. This parallels myometrium, where adenylate cyclase coupled receptors inhibit myometrial contractility ^{86;220}.

The third pattern was a lower level of expression of the gene encoding an adenylate cyclase coupled receptor (EP₂) in decidua obtained from animals in spontaneous labour (Figure 6.6A). This suggests that an adenylate cyclase-mediated effect may inhibit

parturition in decidua in cells expressing the EP₂ receptor. This parallels decreased EP₂ receptor expression in myometrium and cervix in labour (Chapters 4 and 5). I hypothesize that reduced expression of this inhibitory receptor may have a key role in promoting parturition in the primate. The absence of any change in expression of the EP₂ receptor in chorion suggests differential transcriptional control in maternal and fetal intra-uterine tissues.

The fourth pattern, observed in both the decidua and chorion, was an increase in the expression of a gene encoding an adenylate cyclase coupled receptor (IP) and this was particularly marked in the chorion (Figure 6.6B and C). This suggests that adenylate cyclase activation might promote parturition in cells expressing the IP receptor in these tissues. It was interesting to observe two adenylate cyclase coupled prostanoid receptors (EP₂ and IP) vary in opposite directions in decidua in association with labour. This underlines the potential for the same second messenger system to mediate different effects in different cells types and this presumably has its basis in differential effects of protein kinase A in different cell types. Activation of the IP receptor in myometrium inhibits myometrial contractility and this observation, like decreased EP₁ and FP expression in decidua with advancing gestational age, suggests dissociation of roles of receptors in choriodecidua compared with other key tissues. The present findings suggest that extreme caution should be applied in any studies seeking to exploit the IP receptor pathway as a means of uterine tocolysis, since any inhibitory, therapeutic effect on myometrium³⁶ may be undermined by effects on the choriodecidua.

Cellular localization of expression of these receptors may allow some inferences to be drawn regarding their physiological roles. I attempted in situ hybridisation for the EP₂, EP₃, EP₄ and IP receptor genes. However, reliable signals were not obtained due to low levels of expression and high levels of non-specific binding. Other techniques to localize

signal have certain limitations. Receptor autoradiography is limited by the extensive cross reactivity of available ligands ⁴⁷, and immunocytochemistry is limited by the lack of well characterized and specific antibodies.

There are relatively few data on prostanoid receptors in the fetal membranes and decidua. A previous study failed to demonstrate variation in either EP₂ or EP₄ receptor gene expression in labour in ovine endometrium in association with labour ²¹⁵. However, species differences between the sheep and baboon have previously been demonstrated for EP₂ receptor mediated effects on myometrium ^{86;89;90}. Furthermore, myometrial EP₂ receptor gene expression decreases in both the baboon and human ^{91;225} in labour or at term, whereas it is unchanged in sheep ²¹⁵. These findings, taken together, suggest significant species difference between primates and sheep in terms of uterine prostanoid receptors and underline the importance of studies in non-human primates.

A number of studies have postulated a role for prostanoid receptors in the endometrium in association with implantation ²²⁶. An important role for PGs in this context is supported by the failure of implantation and decidualisation in mice which are null mutant for the COX-2 gene ²²⁷, and this may be due to the absence of PGI₂ ⁴⁵. Other studies have demonstrated expression of the EP₁ receptor on amnion WISH cells. Expression of the EP₁ receptor in these tissues is stimulated by CRH ²²⁸, interleukin-1beta ²²⁹, interleukin-4 ²³⁰ and tumour necrosis factor-alpha ²³¹. Interestingly, the effect of CRH appeared to be mediated by PGE₂ ²²⁸, suggesting the possibility of a feedback loop. However, the data presented in this chapter are the first, to my knowledge, to support the concept that a changing pattern of prostanoid receptor expression may have a role in choriodecidual activation in labour.

Chapter 7. The effect of corticosteroids, gestational age and birth on expression of EP receptor genes in ductus arteriosus

The ductus arteriosus is a shunt blood vessel of fetal life. It diverts deoxygenated blood from the main pulmonary artery to the descending aorta. Following birth, the direction of flow across the ductus reverses and the vessel ultimately closes in the first 1-2 days of neonatal life.

Patency of the ductus in fetal life is an active state and is maintained by the tonic dilator effect of high circulating concentrations of PGE₂, which relaxes ductal smooth muscle through the adenylate cyclase coupled EP₄ receptor³⁹. Cyclic adenosine monophosphate (cAMP) inhibits the intrinsically high sensitivity of ductal smooth muscle to the [Ca²⁺]_i¹⁰¹. Following birth, circulating concentrations of PGE₂ fall and loss of PGE₂-mediated activation of adenylate cyclase uncovers the vessel's intrinsically high sensitivity to [Ca²⁺]_i⁹⁹. Furthermore, the physiological increase in arterial oxygen tension which follows birth depolarises ductal smooth muscle by closing oxygen sensitive delayed rectifier potassium channels resulting in intracellular calcium influx in ductal smooth muscle cells^{232;233}. The central role of the prostanoid EP₄ receptor in the control of ductal patency has been underlined by the observed failure of normal closure of the vessel following birth in EP₄ receptor gene null mutant mice¹⁰³. However, the paradox that the absence of a gene encoding an inhibitory receptor results in failure of ductal contraction following birth remains to be satisfactorily explained.

The sensitivity of the ductus arteriosus to the dilator effect of PGE₂ is decreased by antenatal exposure to corticosteroids²³⁴, by advancing gestational age²³⁵ and following birth²³⁶. However, these factors also affect the sensitivity of the vessel to a range of other vaso-active agents [see⁹⁹ for review], and it is not clear whether the altered response to PGE₂ represents a specific change in ductal PG signal transduction. Furthermore,

functional studies also suggest that the ductus expresses a contractile EP receptor, probably EP₃¹⁰². It is not clear whether an alteration in the balance of contractile and dilator EP receptors has a physiological role in the ontogenic and perinatal control of the vessel.

The ductus arteriosus, like myometrium, is profoundly under the control of PGs and most current models of ductal control – like models of parturition - focus on the potential role of changes in PG concentration. In the preceding chapters, I demonstrated that advancing gestational age and labour are associated with changes in the relative expression of prostanoid receptor genes in key intra-uterine tissues. The aim of this part of my study was to determine whether this was also true of the fetal preparation for birth. Therefore, I sought to test the hypothesis that loss of EP₄ receptor gene expression might have a role in mediating ductal closure following birth. I also predicted that the ratio of EP₄ to EP₃ gene expression might change following birth, due to decreased EP₄ expression, increased EP₃ expression or both. I also sought to determine the effect of antenatal corticosteroids and advancing gestational age on the expression of these genes and to determine whether EP₄ receptor mRNA expression could be demonstrated in the non-human primate.

7.1 Methods.

The general methods described in Chapter 2 were employed. In these experiments, chronically instrumented fetal lamb preparations were used.

7.1.1 Obtaining fetal and neonatal lamb ductus arteriosus

Pregnant Rambouillet-Colombia ewes and baboons of known gestational age were anaesthetised (ketamine induction, then halothane), the fetus was delivered by Caesarean section, and exsanguinated. Neonatal Rambouillet-Colombia lambs were anaesthetized with halothane and exsanguinated.

7.1.2 Instrumented fetal lamb preparations

Rambouillet-Colombia ewes bred on a single occasion and of known gestational age were acclimated to the animal facilities for at least 5 days before surgery and kept in rooms with controlled light/dark cycles. Alfalfa cubes and water were provided ad libitum. Surgery was performed at 104 ± 1 (mean \pm SEM) days gestational age (dGA). Anaesthesia was induced with ketamine and maintained with halothane. Catheters were inserted into the ewe's carotid artery and jugular vein, into the fetal carotid and axillary arteries and jugular and pedal veins, and into the amniotic cavity. Electrodes were implanted bilaterally on the fetal parietal bone and above the orbit and on each side on the surface of the maternal uterine body and pregnant horn.

After surgery the ewe was returned to a metabolism cage and provided free access to food and water. Animals were allowed at least five days of post-operative recovery and all ewes received a daily antibiotics (ampicillin) and twice daily analgesics (phenylbutazone, until day three post-op).

Animals were randomly assigned to receive dexamethasone (n=4) or vehicle (n=4) at 110 dGA. Infusion of either vehicle or dexamethasone (2.2 µg/kg/min at 1 ml/h) was maintained for forty two hours. Immediately preceding and following the infusion, the fetuses were exposed to a continuous series of 2 minute infusions of phenylephrine (2.5, 5, 10, 20, and 40 µg/min) and, after a 60 minute recovery period and a set of fetal and maternal blood samples, a continuous series of 2 minute infusions of nitroprusside (1, 2, 4, 8, and 16 µg/min). Infusion of either phenylephrine or nitroprusside was stopped if the change in blood pressure exceeded 30% of baseline or if arrhythmia was noted on the pressure tracing. Ewes were euthanised by intravenous injection of pentobarbital sodium solution (Fatal-Plus, Vortech Pharmaceuticals, Dearborn, MI).

7.1.3 Instrumented fetal baboons

Five pregnant baboons (*Papio* sp.) of known gestational age were employed as described in Chapter 2. At 118-134 days of gestational age (dGA, term = 185dGA) animals were instrumented under halothane (1-2% in 2 l/min O₂) general anaesthesia. Maternal femoral artery and vein and fetal carotid artery catheters were placed and a 22G catheter was used for fetal carotid cannulation. Five electrodes (New England Wire Corp., Lisbon, New Hampshire) were implanted on the uterine body for electromyographic recording of uterine activity.

On the morning of the fifth day after surgery, animals were given an intra-muscular injection of 87.5µg/kg betamethasone (Celestone, betamethasone sodium phosphate injection, USP, Schering Corp., Kenilworth, NJ;) or vehicle. A total of 4 doses at 12 hour intervals were given and the fetus was delivered by Caesarean section 12 hours after the last dose. In addition, further baboon fetuses were delivered by planned Caesarean section as described in Chapter 2.

7.2 Results.

Northern analysis demonstrated clear signals for the EP₄ and EP₃ receptor genes from RNA extracted from fetal and neonatal lambs (Figure 7.1). The size of transcripts was similar to other reports of sheep prostanoid receptors in non-vascular tissue²¹⁵. The blots were also probed with riboprobes for the EP₁, EP₂, FP and IP receptors, but none of these probes resulted in a signal that was clearly greater than background.

The level of expression of the EP₄ receptor gene in lambs at approximately 110dGA was not significantly different comparing un-instrumented lambs and instrumented lambs receiving vehicle or dexamethasone infusion (Table 7.1). However, EP₄ receptor gene expression was significantly lower in late term fetuses (145dGA) and lower still in neonates (Table 7.1, Figures 1 and 2). The pattern of EP₄ expression and statistical significance was similar when expression was normalized to either beta-actin or glyceraldehyde-3-phosphate dehydrogenase (GAPDH) (Table 7.1 and Figure 7.2). The level of expression of the 4.2 kB EP₃ receptor gene transcript was similar in all the groups studied (Figure 7.3A and Table 7.1). The ratio of the size of the EP₄ signal to the 4.2kB EP₃ signal was lower in late gestation and lower still in neonates (Table 7.1 and Figure 7.3B). The results were virtually identical for the 7.5 and 3.5 kB EP₃ transcripts (data not shown).

Samples were available from a number of fetal baboons, both controls and four steroid treated animals (Figure 7.4). In these animals there was no reduction in the level of EP₄ mRNA expression following treatment with betamethasone (Figure 7.5).

In situ hybridization of the baboon ductus confirmed that the gene was expressed in the smooth muscle cells of the vessel and demonstrated the specificity of the EP₄ signal for the ductus over the aorta (Figure 7.6A and C). The hybridization was specific as confirmed by the use of a sense probe (Figure 7.6B).

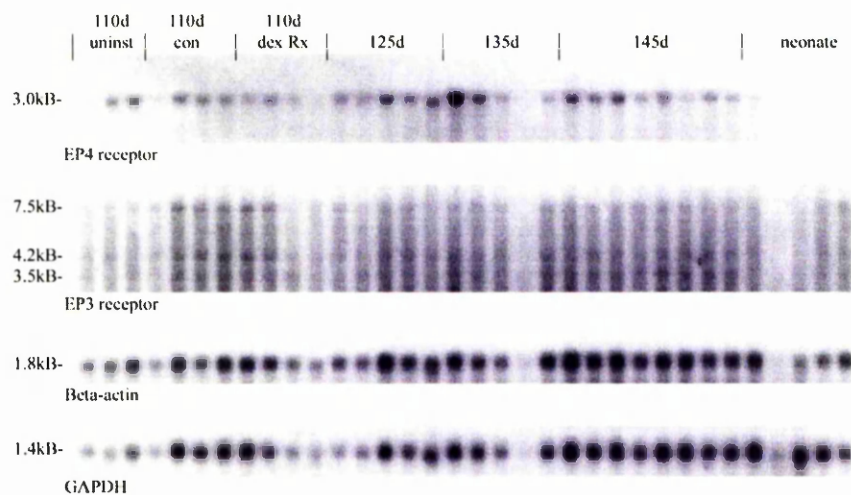


Figure 7.1. Northern blot of fetal and neonatal lamb mRNA, probed for EP₄, EP₃, beta-actin and GAPDH genes. One microgram of polyadenylated RNA was loaded in each lane. The estimated size of transcripts is expressed in kilobases (kB). Lanes 20 and 31 were not used in subsequent analysis due to the absence of a reliable signal to either GAPDH or beta-actin. Numbers equal gestational age at time of delivery, e.g. 145 = animals delivered at 145 days gestational age. “110d uninst” = un-instrumented animals at 110 days. “con” = instrumented animals at 110 days acting as controls for dexamethasone (received vehicle). “dex Rx” = instrumented animals treated with dexamethasone. All other animals un-instrumented.

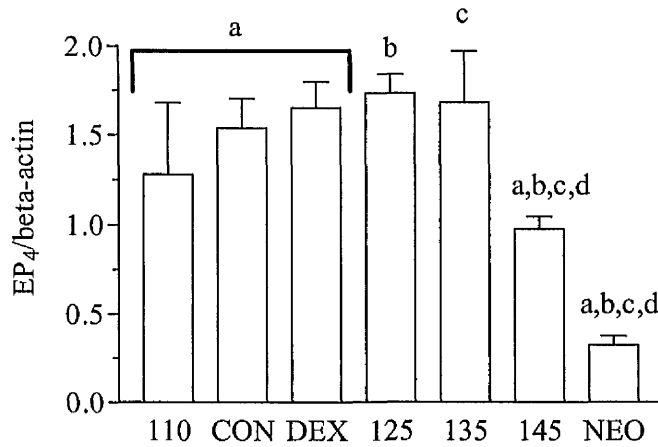


Figure 7.2. Densitometric quantitation of EP₄ signal in each of the seven groups relative to beta-actin. Key to X axis: un-instrumented at 110 days ('UN', n=3), instrumented and treated with dexamethasone vehicle at 110 days ('CON', n=4), instrumented and treated with intravenous dexamethasone at 110 days ('DEX', n=4), un-instrumented at 125 days ('125', n=5), un-instrumented at 135 days ('135', n=4), un-instrumented at 145 days ('145', n=8), un-instrumented neonates ('NEO', n=4). Groups with the same letter were statistically significantly different during post-hoc multiple comparisons (Tukey's method).

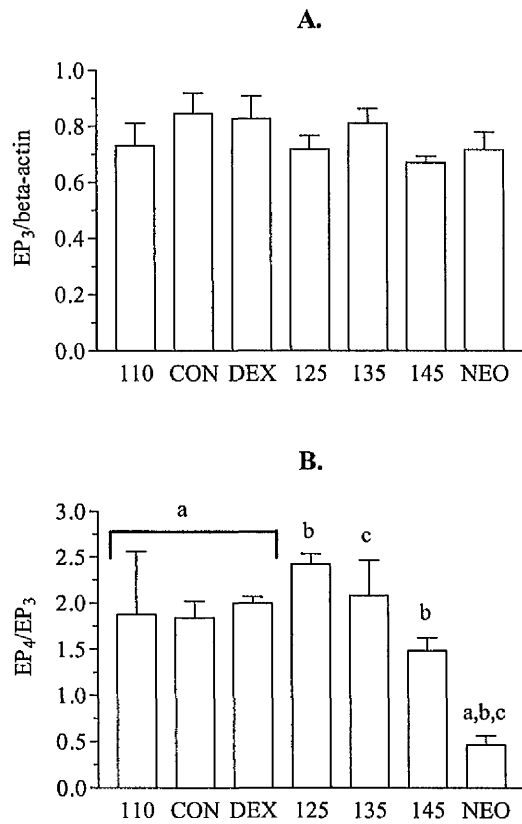


Figure 7.3. Densitometric quantitation of 4.2kB EP₃ transcript to A. ratio of EP₃ to beta-actin B. ratio of EP₄ to EP₃ in each of the seven groups. Key to X axis: un-instrumented at 110 days ('UN', n=3), instrumented and treated with dexamethasone vehicle at 110 days ('CON', n=4), instrumented and treated with intravenous dexamethasone at 110 days ('DEX', n=4), un-instrumented at 125 days ('125', n=5), un-instrumented at 135 days ('135', n=4), un-instrumented at 145 days ('145', n=8), un-instrumented neonates ('NEO', n=4). Groups with the same letter were statistically significantly different during post-hoc multiple comparisons (Tukey's method).

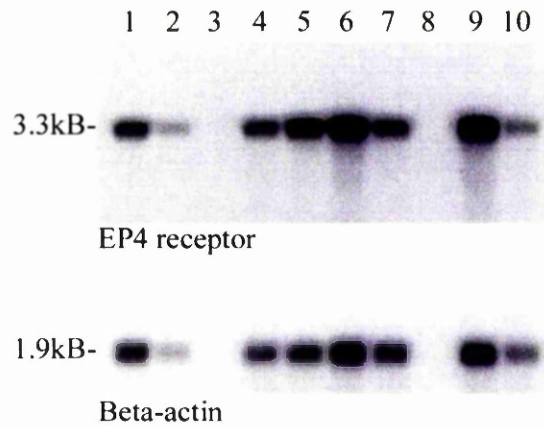


Figure 7.4. Northern blot of fetal baboon ductus arteriosus probed for EP₄ gene and beta-actin. 0.5 μ g of polyadenylated RNA was loaded in each lane, except lane two, where 0.35 μ g was loaded. The estimated size of transcripts is expressed in kilobases (kB). Lane 1=128dGA, un-instrumented; Lane 2=133dGA, instrumented, vehicle control for betamethasone; Lane 3=blank; Lanes 4-7, all instrumented and exposed to maternally administered betamethasone (dGA in order: 124, 127, 129, and 137); Lane 8=blank; Lane 9=un-instrumented 153dGA, Lane 10 = un-instrumented 158dGA.

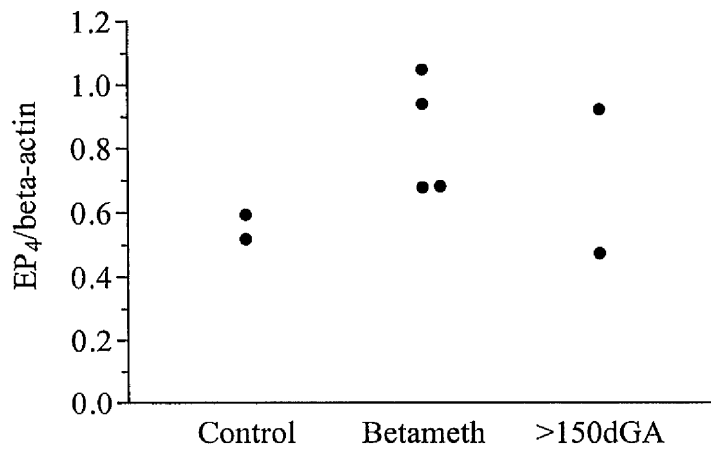


Figure 7.5. Densitometric quantitation of ratio of EP₄ signal to beta-actin in baboon ductus arteriosus. “Control” = 128 dGA, un-instrumented and 133 dGA, instrumented, vehicle control for betamethasone. “Betameth” = all instrumented and exposed to maternally administered betamethasone (dGA in order: 124, 127, 129, and 137). “>150dGA” = un-instrumented 153dGA and un-instrumented 158dGA. One way ANOVA of the three groups: P=0.35. Unpaired t-test of specific hypothesis that EP₄ expression was lower in the betamethasone treated animals compared with “Control”: p=0.97.

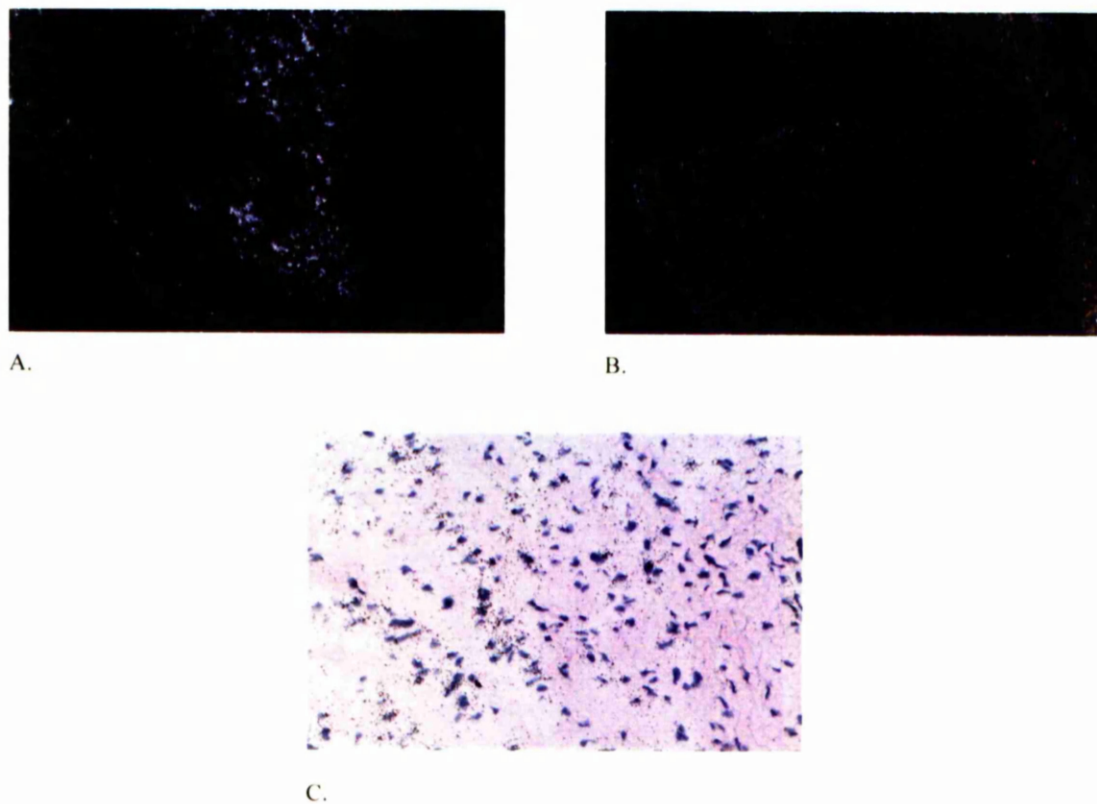


Figure 7.6. In situ hybridisation of EP₄ receptor gene expression in baboon ductus arteriosus and aorta. Sections from un-instrumented animal delivered by Caesarean section at 172 days gestational age, having received no antenatal drug treatment. A. Dark field, anti-sense probe, B. Dark field, sense probe, C. Haematoxylin and eosin section, anti-sense probe, EP₄ signal visualized as silver grains. A and B, low power: ductal tissue in middle of field. C, high power: ductal tissue on upper left of field and aorta on lower right.

Table 7.1. Three way analysis of variance: effect of fetal surgery, dexamethasone and gestational age and birth on expression of prostanoid receptor genes.

	Fetal surgery	Dexamethasone	Gestational age & birth
	P=	P=	P=
EP ₄ /beta-actin	0.35	0.65	<0.001
EP ₄ /GAPDH	0.36	0.31	<0.001
EP ₃ /beta-actin	0.22	0.60	0.36
(4.2kB transcript)			
EP ₄ / EP ₃	0.28	0.79	<0.001
(4.2kB EP ₃ transcript)			

GAPDH denotes glyceraldehydes-3-phosphate dehydrogenase, kB denotes kilobases.

7.3 Discussion

The main finding reported in this chapter is that expression of the gene encoding the prostanoid EP₄ receptor in the ductus arteriosus was lower in term fetal lambs compared with preterm lambs, and was lower still in neonatal lambs. However, there was no variation in expression of the gene encoding the contractile EP₃ receptor in the same animals. The main role of the EP₄ receptor in the control of the ductus had been thought to be to modulate variation in the local and circulating concentrations of PGE₂⁹⁹. The present data suggests that variation in the level of the EP₄ receptor gene may itself have a role in the control of the vessel. It seems likely that an altered balance of EP₄ and EP₃ mediated effects on the ductus, in the face of a given concentration of PGE₂, has a role in promoting contraction of the vessel in neonatal life.

The EP₄ receptor appears to be important in the control of the ductus with conservation in a number of species. It was first demonstrated in rabbits³⁹ and subsequently in mice¹⁰³. In the present study I have now demonstrated expression of the gene encoding the receptor in the lamb and baboon ductus arteriosus (Figures 7.1 and 7.4). Given this high degree of species conservation, it seems likely that the EP₄ receptor will have a similar key role in the control of the human ductus. This is supported by the high sensitivity of the human ductus to PGE₂. In human infants with ductus dependent heart defects the vessel is dilated by exogenous synthetic PGE₂ in the sub-nanomolar range of circulating concentrations²³⁷. This is consistent with the known EC₅₀ for the EP₄ receptor, which is more sensitive to PGE₂ than the other inhibitory EP receptor, the EP₂ receptor^{13;32;198;238}.

The current findings help explain the apparently paradoxical observation that the ductus is patent in utero in EP₄ null mutant mice, but fails to close following birth¹⁰³. Taken at face value, this finding might be thought to suggest that the EP₄ receptor somehow mediates contraction of the vessel in neonatal life. This is implausible both from the

pharmacological perspective, as the receptor mediates relaxation of ductal smooth muscle³⁹, and on the basis of the current findings, since birth was associated with a decrease in expression of the EP₄ receptor gene (Figures 7.1 and 7.2). A major factor in the control of the ductus is the vessel's intrinsically very high level of endogenous contractile tone: acute withdrawal of PGE₂-mediated inhibition using a PG synthesis inhibitor (indomethacin) results in ductal closure and fetal heart failure¹⁰⁰. The ductus of the EP₄ null mouse does not contract in response to indomethacin¹⁰³. I hypothesize that the high level of EP₄ receptor-mediated inhibition of ductal tone is essential for the normal development of the vessel's intrinsic tone in utero. Since contraction of the vessel following birth is dependent on the vessel's intrinsic contractile tone, this model would explain the apparent paradoxical finding of patent ductus arteriosus in EP₄ null mutant mice¹⁰³. This model would also explain the observation that human fetuses exposed to indomethacin in utero are more likely to have a patent ductus arteriosus in neonatal life than infants born at a similar gestational age that were not exposed to indomethacin²³⁹. The lack of effect of corticosteroids on the expression of the EP₄ and EP₃ receptor genes was of interest. It had previously been shown that treatment with corticosteroids decreased the sensitivity of the preterm lamb ductus to the effects of exogenous PGE₂²³⁴. On the basis of the current data, it seems that this is unlikely to be due to variation in the relative balance of EP₄ and EP₃ gene expression. However, I cannot rule out altered translation of EP₄ and EP₃ mRNA following steroid treatment or an effect of corticosteroid on receptor mRNA transcription in late gestation and following birth. Furthermore, steroids may also affect the balance of EP₄ and EP₃ mediated effects by altering transcription and/or translation of other genes which might affect the relative balance of activation of the two receptors, such as genes encoding G-proteins and second messengers. However, it is of interest that there was a decrease in expression of the EP₄

receptor gene with advancing gestational age, but no apparent change following exposure of the preterm fetal lamb to corticosteroids (Figure 7.2). Many of the effects of advancing gestational age can be induced by antenatal exposure of the preterm fetus to corticosteroids and the use of corticosteroids is widespread in the management of threatened preterm delivery, as a means of advancing pulmonary maturity ²⁴⁰. The present study demonstrates an effect of gestational age which was not induced by exogenous corticosteroids, despite the fact that corticosteroids have been shown to affect other aspects of ductal function in the preterm lamb ²⁴¹. This underlines the potential importance of endocrine systems other than the adrenocortical system in ontogenic development of the fetus. Thyroid hormones also have maturational effects on the preterm fetus ²⁴⁰ and been shown to alter contractility of the ductus ²⁴².

It is possible, however, that the apparent lack of effect of corticosteroids is artefactual. It may represent a type II error due to lack of statistical power. However, using the standard deviations obtained from densitometry, I estimate that with an n of 4 in each group, the analysis had 71% power to detect the same magnitude of difference with steroid treatment as was observed comparing 110dGA and 145dGA fetuses. The apparent lack of effect of steroids may be methodological, e.g. the dose of corticosteroid was insufficiently high to induce altered gene expression. However, as in previous studies, the fetus was noted to be hypertensive during steroid treatment, both the lambs (unpublished data, Koenen, Nijland and Nathanielsz, 1999) and baboons ²⁴³ which suggests that the dose was not sub-therapeutic. Finally, the instrumented lambs were also exposed to both phenylephrine and sodium nitroprusside as part of another experiment. Administration of these drugs may have altered prostanoid receptor gene expression and masked an effect of dexamethasone. This seems unlikely, however, as there was no difference in EP₄ gene expression among 110dGA fetuses, comparing un-instrumented animals and

instrumented animals which were controls for dexamethasone and received phenylephrine and sodium nitroprusside (Table 7.1).

A study of the ductus arteriosus in piglets demonstrated several important species differences with respect to prostanoid receptors in the vessel ²⁴⁴. These authors demonstrated the presence of EP₂, EP₃ and EP₄ receptors in the fetal piglet ductus, but that only EP₂ receptors were present in the neonatal piglet ductus. This is in contrast to the fetal rabbit ductus which does not express functional EP₂ receptors, as evidenced by the lack of response to the selective EP₂ agonist, AH13205 ³⁹. It also suggests species differences in terms of the EP₄ receptor, which continued to be expressed at the level of mRNA in the neonatal ductus in the mouse ¹⁰³ and in the present study (albeit at lower levels) in the lamb, and in terms of the EP₃ receptor since, in the present study, expression of EP₃ mRNA was observed in the ductus from neonatal lambs. Furthermore, since the EP₃ receptor mediates contraction of the ductus ¹⁰², it remains to be seen how loss of this receptor might be involved in ductal closure in the neonatal piglet.

Another aspect of prostanoid receptor pharmacology which appears to differ in the fetal piglet when compared with other species is the affinity of EP receptor sub-types. The neonatal piglet ductus expressed only EP₂ receptors, whereas the fetal ductus expressed approximately equivalent levels of EP₂, EP₃ and EP₄ receptors ²⁴⁴. The EP₂ receptor has a five to tenfold lower affinity for PGE₂ than either the EP₃ or EP₄ receptor, which has been observed in functional studies in a number of species ³⁷ and confirmed in recombinant mouse prostanoid receptors ⁴⁷. However, PGE₂ was bound with greater affinity in neonatal ductus than fetal ductus (K_d , 6.1nM versus 10.8nM, respectively), despite the apparent loss of receptors which bind PGE₂ with five to tenfold greater affinity than the EP₂ receptor in other species. Taken together, these observations suggest that the piglet ductus may be somewhat atypical in terms of prostanoid pharmacology.

Since the expression of EP receptor sub-types in the neonatal ductus is highly relevant for the development of drugs for the clinical manipulation of the human ductus, the expression of EP receptors in primate neonates should be an area of further study.

Chapter 8. The effect of labour and gestational age on expression of lipoyxygenase genes in intra-uterine tissues

A large number of studies have underlined the key role of PGs in the control of parturition in a number of species ⁵. PGs are formed from AA by one of two isoforms of the enzyme COX. Expression of the inducible form of this enzyme (COX-2) has been shown to increase both with advancing gestational age and labour ^{25;80;245} and these studies led directly to clinical studies of selective COX-2 inhibitors in the management of preterm labour ²²³.

The PG pathway is only one of several pathways of AA metabolism which results in biologically active products ¹⁰. Others include the epoxide pathway (cytochrome P450) ¹⁷, the isoprostane pathway (non-enzymatic generation of isoprostanes such as 8-epi-PGF_{2α}) ¹⁸ and the LOX pathway ¹⁹. The last of these generates HPETE compounds which are metabolised to a range of eicosanoids, including the leukotrienes (LTs) and some of these compounds have been shown to have effects on myometrium ¹⁵⁹.

Whereas the expression of COX genes has been systematically studied in a number of species, there are few studies systematically addressing variation in the expression of LOX genes with advancing gestation and in relation to labour. In this chapter I report expression of the three main recognized human LOX genes in myometrium (both fundus and lower segment), cervix, decidua and chorion.

8.1 Methods

The same animals described in Chapter 4 were employed and the general methods were as outlined in Chapter 2. All LOX probes were cDNA.

8.2 Results.

Clear signals of similar size to previous reports of Northern analysis of human RNA were detected using the 5-LOX and platelet-12-LOX probes in myometrium (Figure 8.1), cervix (Figure 8.2), decidua (Figure 8.3) and chorion (Figure 8.4). In the case of 5-LOX there was a single band of approximately 2.4kB. In the case of platelet-12-LOX there was a single band of 2.5kB in decidua, but in myometrium, cervix and chorion, there was also an additional band at 2.2kB. A clear 6.3kB band was observed using the human 15-LOX cDNA probe in myometrium and cervix (Figure 8.5). However, this differs markedly in size from previous reports of the 15-LOX gene in Northern analysis of human tissues²⁴⁶ and it remains to be determined whether this signal truly represents the baboon 15-LOX gene. No signal in any tissues was observed using the murine leukocyte 12-LOX cDNA probe.

Expression of the 5-LOX gene (relative to beta-actin) decreased in the cervix with advancing gestational age, but did not vary significantly with gestational age in any of the other tissues (Table 8.1). Expression of 5-LOX was greater in decidua obtained from animals in labour than animals not in labour, whereas 5-LOX expression in chorion was lower in tissues obtained from animals in labour (Figure 8.6). There was no significant difference in 5-LOX expression comparing labour and not in labour in either cervix or myometrium, and myometrial expression was not significantly different comparing samples obtained from the fundus and lower segment (Table 8.1).

Expression of the 2.5 kB transcript of the platelet 12-LOX gene (relative to beta actin) decreased with advancing gestational age in chorion, but did not vary significantly with advancing gestational age in any of the other tissues (Table 8.1). Expression of the 2.5 kB transcript of the platelet 12-LOX gene in cervix was lower in tissues obtained from animals in labour (Figure 8.7A). There was no significant difference in the 2.5 kB

transcript of the platelet 12-LOX gene comparing labour and not in labour in myometrium, decidua or chorion and myometrial expression was not significantly different comparing the fundus and lower segment (Table 8.1).

Although the 2.2kB platelet 12-LOX transcript was also observed in myometrium and chorion, the signals were too weak for meaningful quantitative analysis. When related to beta-actin, there was no significant change in the level of expression of the 2.2kB transcript in association with either advancing gestational age or labour in the cervix (Table 8.1). The ratio of expression of the 2.5kB platelet 12-LOX gene transcript to the 2.2kB transcript was lower in cervical tissue obtained from animals in labour (Figure 8.7B) but there was no change in the ratio of the two transcripts in association with advancing gestational age in the cervix (Table 8.1).

The apparent 15-LOX signal showed no variation with either gestational age or labour in myometrium and cervix (Figure 8.5).

All cases where significant variation was observed when signals were related to beta-actin were also significant when related to at least one of the other two housekeeping genes (data not shown). Where signals were observed to change with gestational age prior to labour (namely, 5-LOX in the cervix and platelet 12-LOX in the chorion) adjusting the comparison between animals in labour and not in labour for the effect of gestational age (using analysis of covariance) did not alter the statistical significance.

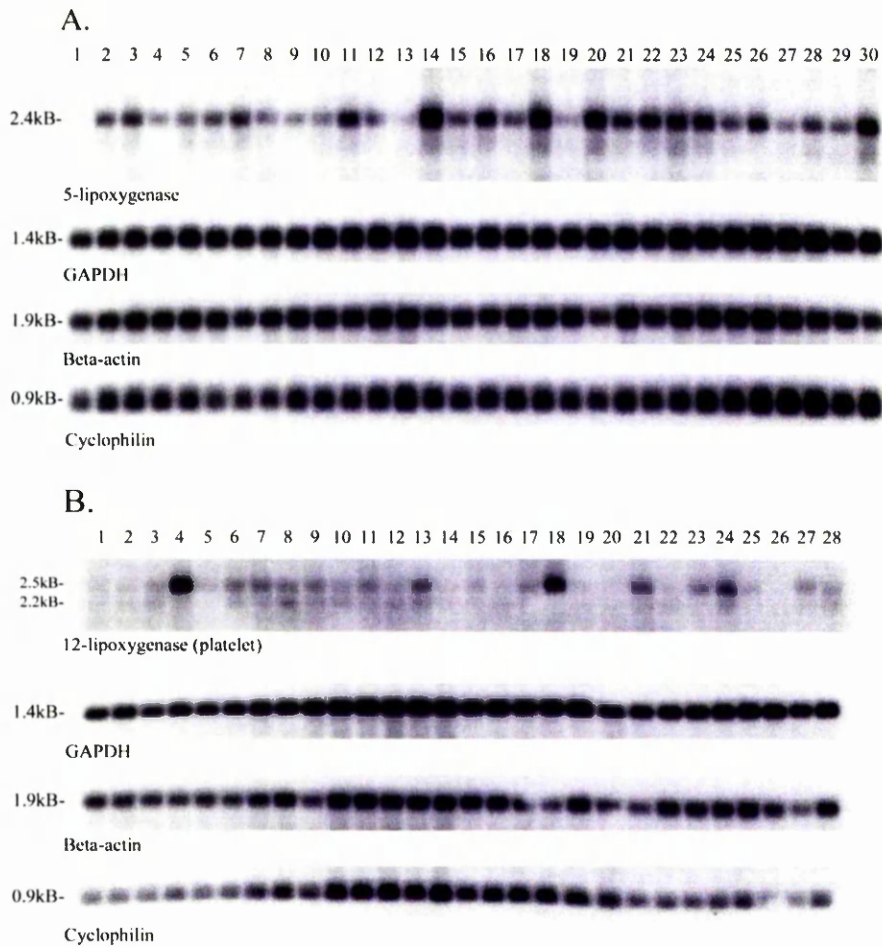


Figure 8.1. Northern blot of LOX expression in myometrium from lower uterine segment (LUS) and fundus (FUN), from animals not in labour (NIL) and in labour (LAB). A. 5-LOX, B. platelet 12-LOX. The estimated size of transcripts is given in kilobases (kB). 2 μ g of RNA was loaded in each lane. Lanes: A. 1-10 LUS, NIL; 11-15 LUS, LAB; 16-25 FUN, NIL; and 26-30 FUN, LAB. B. 1-9 LUS, NIL; 10-14 LUS, LAB; 15-23 FUN, NIL; 24-28 FUN, LAB.

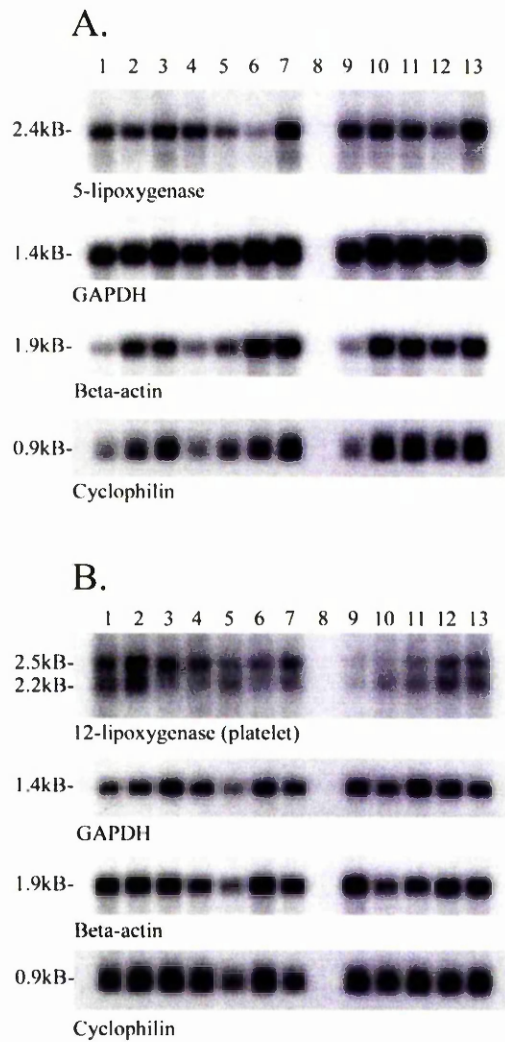


Figure 8.2. Northern blot of LOX expression in cervix from animals not in labour (NIL) and in labour (LAB). A. 5-LOX, B. platelet 12-LOX. The estimated size of transcripts is given in kilobases (kB). 4 μ g of RNA loaded in each lane in A and 2 μ g loaded in each lane in B. Both blots, lanes 1-7 NIL and lanes 9-13 LAB.

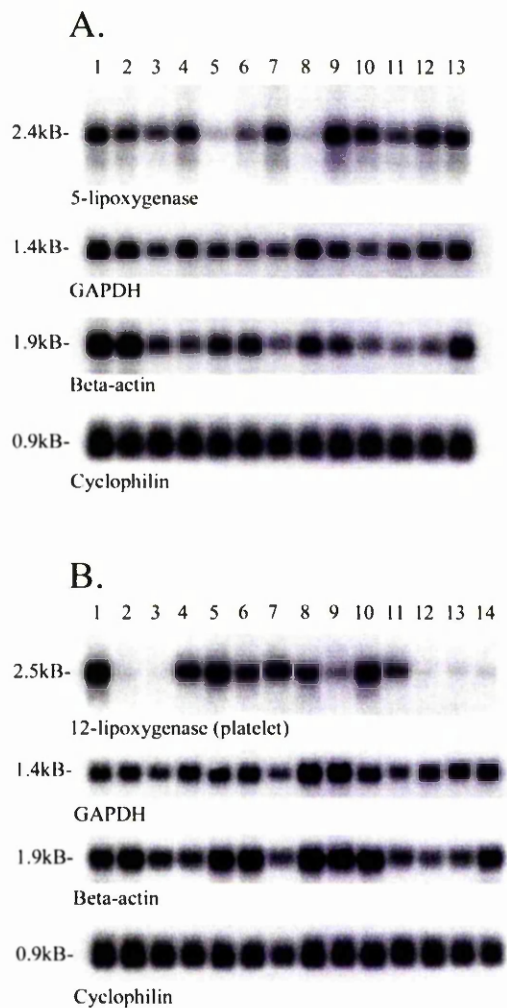


Figure 8.3. Northern blot of LOX expression in decidua from animals not in labour (NIL) and in labour (LAB). A. 5-LOX, B. platelet 12-LOX. The estimated size of transcripts is given in kilobases (kB). 5 μ g of RNA loaded in each lane. Lanes: A. 1-8 NIL, 9-13 LAB; B. 1-9 NIL, 10-14 LAB.

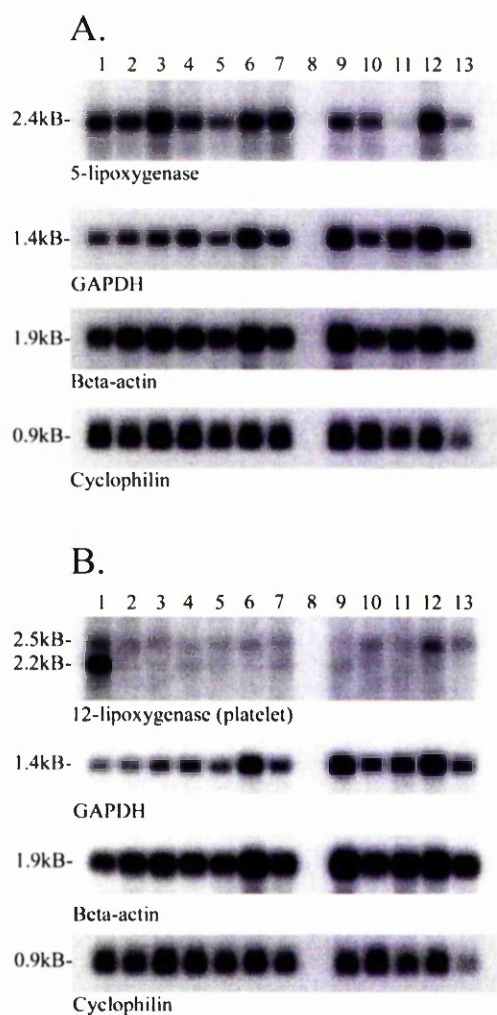


Figure 8.4. Northern blot of LOX expression in chorion from animals not in labour (NIL) and in labour (LAB). A. 5-LOX, B. platelet 12-LOX. The estimated size of transcripts is given in kilobases (kB). 5 μ g of RNA loaded in each lane. Both blots, lanes 1-7 NIL and lanes 9-13 LAB.

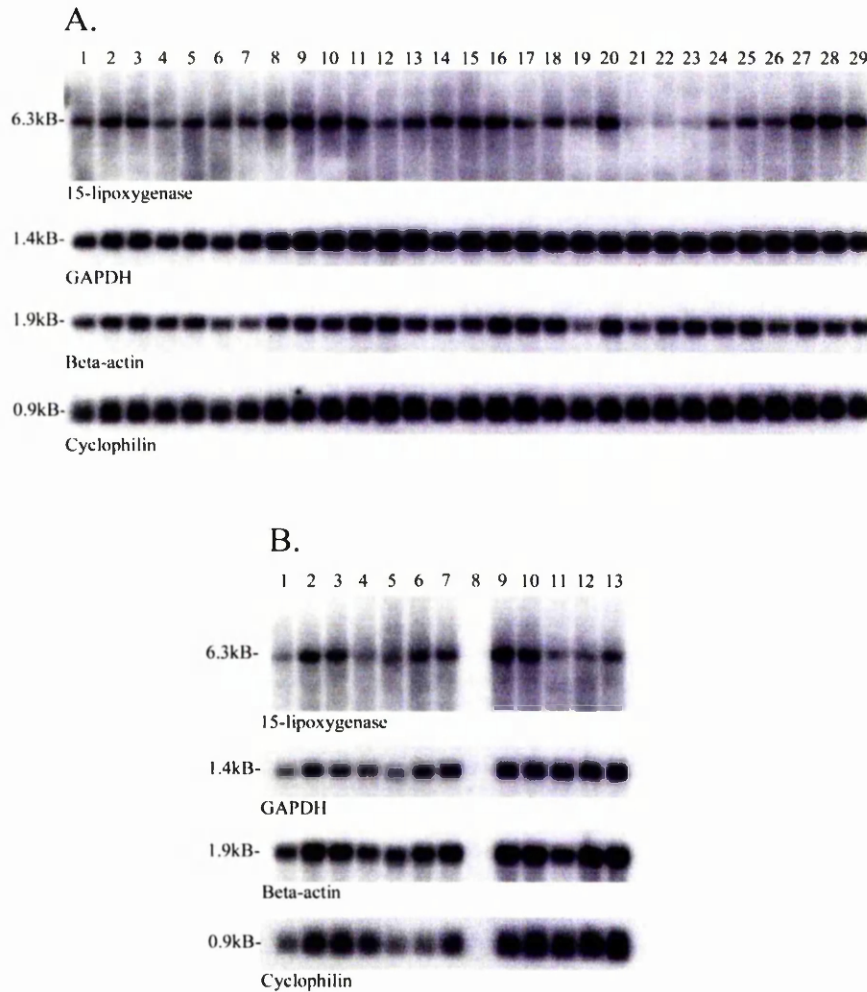


Figure 8.5. Northern blot of 15-LOX expression in A. Myometrium, B. Cervix. Samples of myometrium from both lower uterine segment (LUS) and fundus (FUN), and samples from both tissues obtained from animals not in labour (NIL) and in labour (LAB). The estimated size of transcripts is given in kilobases (kB). Lanes: A. 1-9 LUS, NIL; 10-14 LUS, LAB; 15-24 FUN, LAB; and 25-29 FUN, LAB. B. 1-7 NIL and 9-13 LAB. Statistical comparison (densitometry relative to beta-actin): A. FUN vs. LUS, $p=0.19$ (ANOVA), NIL vs. LAB, $p=0.18$ (ANOVA); B. NIL vs. LAB, $p=0.38$ (Student's t-test). Similar results obtained when signal quantified relative to GAPDH or cyclophilin.

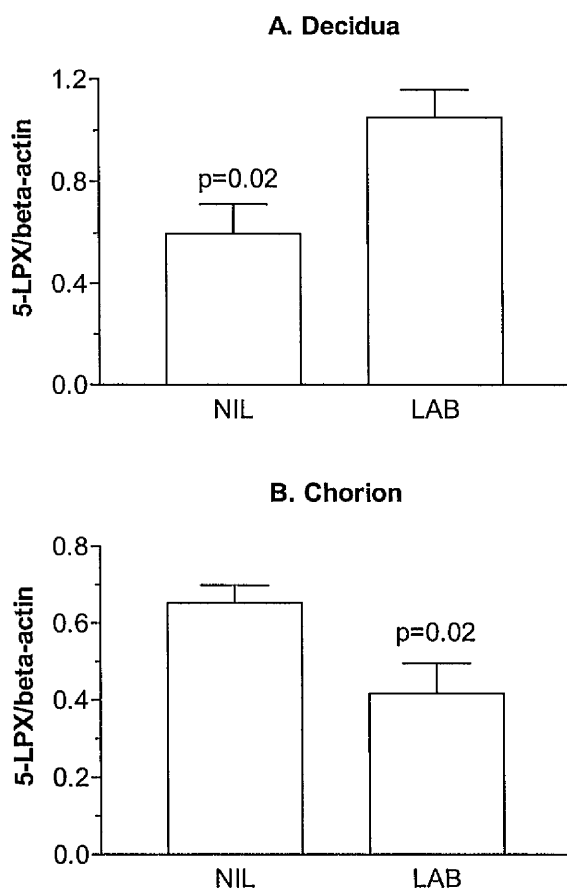


Figure 8.6. Densitometric analysis of 5-LOX expression (relative to beta-actin) decidua and chorion. A. Decidua and B. Chorion, comparing animals in labour (LAB, n=5) with animals not in labour (NIL, n=7-8). Columns are means, bars are SEM. Statistical comparison by Student's t-test.

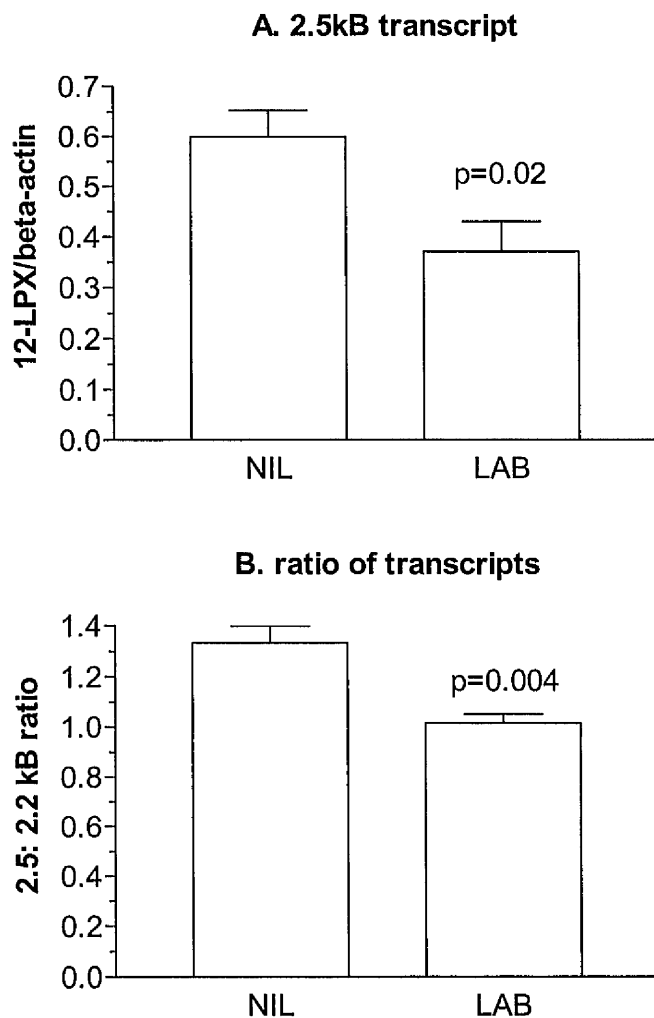


Figure 8.7. Densitometric analysis of 12-LOX expression in the cervix. A. 2.5kb transcript relative to beta-actin and B. 2.5kb transcript relative to 2.2kb transcript, comparing animals in labour (LAB, n=5) with animals not in labour (NIL, n=7). Columns are means, bars are SEM. Statistical comparison by Student's t-test.

Table 8.1. Expression of 5-LOX and platelet 12-LOX in relation to gestational age and labour in baboon intra-uterine tissues.

	5-lipoxygenase				Platelet 12-lipoxygenase¶			
	Means (SEM)		P =		Means (SEM)		P =	
	NIL	LAB	GA		NIL	LAB	GA	LAB
	(n=7-10)	(n=5)	(NIL)	LAB	(n=7-10)	(n=5)	(NIL)	
LUS	0.25 (0.03)	0.35 (0.10)	0.96		0.49 (0.14)	0.24 (0.04)	0.57	
Fundus	0.39 (0.05)	0.34 (0.09)	0.77	0.69	0.38 (0.13)	0.26 (0.07)	0.69	0.33
Fundus* labour				0.20				0.17
Cervix	1.23 (0.28)	1.11 (0.22)	0.02	0.75	2.5kB: 0.60 (0.05)	0.37 (0.05)	0.79	0.02
					2.2kB: 0.46 (0.05)	0.37 (0.06)	0.68	0.27
					ratio: 1.33 (0.07)	1.02 (0.03)	0.61	0.004
Decidua	0.59 (0.12)	1.05 (0.11)	0.72	0.02	0.90 (0.17)	0.62 (0.20)	0.95	0.32
Chorion	0.65 (0.05)	0.42 (0.08)	0.56	0.02	0.23 (0.04)	0.20 (0.02)	0.02	0.51

¶Unless stated otherwise, expression of platelet 12-LOX refers to the 2.5kB transcript.

GA denotes gestational age, NIL denotes not in labour, LAB denotes in labour, kB denotes kilobases, SEM denotes standard error of the mean.

8.3 Discussion.

A key role has been established for PGs in the control of labour in a number of species ⁵. The elucidation of the physiological role of PGs in labour led directly to the clinical use of PG synthesis inhibitors in preterm labour, and these drugs are currently widely clinically employed ²¹⁰. The PG pathway is, however, just one of several pathways through which AA can be metabolised. Much less is understood about the role of these other pathways in the control of parturition. A wide range of drugs have been developed which both inhibit LOX enzymes and block receptors through which LOX products act ^{247;248} If LOX products have a role in parturition, there is the potential for these drugs to be employed in the clinical management of preterm labour. A key line of evidence in understanding the role and site of PG synthesis in the control of parturition was elucidation of quantitative changes in COX gene expression in association with labour ^{25;245} The aim of the present study was, therefore, the systematic study of LOX gene expression in key intra-uterine tissues with advancing gestational age and labour.

The three main LOX genes identified in the human are 5-LOX, platelet 12-LOX and 15-LOX ¹⁹. In the mouse there is another isoform, namely, leukocyte 12-LOX. However, this enzyme also has 15-LOX activity and shares 65% amino acid homology with human 15-LOX ¹⁹. It is thought that murine leukocyte 12-LOX is the homologue of 15-LOX in the human and, consistent with this hypothesis, I could not detect leukocyte 12-LOX expression in any of the baboon tissues. The signals observed in the present study using the 5-LOX and platelet 12-LOX probes were similar to previous reports of Northern analysis in human tissues ^{249;250} However, the transcript observed using the 15-LOX probe was approximately twice as large as previous descriptions of Northern blot of human tissue ²⁴⁶. I have reported these findings with this probe for completeness (Figure

8.5), but suggest that the results be interpreted with some caution until more is known about the baboon 15-LOX gene.

The changes observed in expression of 5-LOX were tissue specific. Expression of the gene decreased with advancing gestational age in cervix and was lower in animals in labour in chorion. However, 5-LOX expression was greater in decidua obtained from animals in labour. Increased 5-LOX expression suggests that 5-HPETE or its metabolites might promote parturition. This could either be due to autocrine effects on the decidua or paracrine effects on myometrium. Consistent with this, 5-HETE has been shown to stimulate contractility in myometrial strips obtained from the LUS of pregnant women¹⁵⁹. The decrease in 5-LOX expression in cervix and chorion may indicate that in these tissues, 5-HPETE or its metabolites may act to inhibit the process of parturition. Consistent with this hypothesis, LTB₄ has been shown to exert inhibitory effects on cervical tissue obtained from pregnant women at term²⁵¹. An alternative possibility is that 5-HPETE and its metabolites are without significant effects in these tissues and that the main biological consequence of lower levels of 5-LOX expression may be to divert free AA to the PG pathway. These mechanisms may also act additively, since exogenous LTB₄ has been shown to inhibit PGE₂ release in human fetal membranes²⁵². Therefore, decreased 5-LOX may stimulate PGE₂ release due to both an increase in the free AA pool and direct effects of LTs on PG biosynthesis.

The size of the platelet 12-LOX transcript observed in the baboon (2.5kB) was very similar in size to the human transcript (2.4kB) which supports the interpretation that this signal is likely to represent the platelet 12-LOX gene in the baboon²⁵⁰. However, in myometrium, cervix and chorion a second transcript of 2.2kB was also observed. The level of expression of the smaller transcript was lower than the 2.5kB transcript and quantitative analysis of the level of expression was only possible in cervix. Interestingly,

labour appeared to affect differentially expression of the two transcripts in the cervix (Figure 8.2, Table 8.1). The practical significance of these observed changes in relative expression is as yet unclear although isoforms of 12-LOX have previously been described¹⁹. However, the fact that changes were observed is consistent with a role for platelet 12-LOX, in some form, in the control of parturition in these tissues. As in the case of 5-LOX this could be by effects on PG synthesis or by effects of 12-HPETE (or its metabolites) on key intra-uterine tissues. It has been shown that 12-HETE had no effect on the contractility of isolated strips of human myometrium¹⁵⁹. As in the case of 5-LOX, it may be that reduced platelet 12-LOX activity might be a factor in determining increased PG levels in labour, amplifying the effect of other known factors which increase PG synthesis in labour such as increased expression of PLA₂ and COX-2⁸⁰.

Comparing the findings of this study with other studies, it has also been shown in the human that mRNA encoding the 5-LOX gene increases in the choriodecidua in association with labour and this was paralleled by increased 5-LOX protein¹⁵⁸. This is consistent with the observation that amniotic fluid concentrations of both 5-HETE and LTC₄ are higher in women in samples obtained from women in labour compared with women not in labour^{253;254}. The finding that 5-LOX and 12-LOX were the main LOX enzymes in intra-uterine tissues is consistent with biochemical studies of human intra-uterine tissues that have demonstrated that 5-HETE and 12-HETE are the main LOX products of AA in human intra-uterine tissues^{255;256}. My hypothesis that decreased 12-HETE formation in labour may play a role in increasing free AA and thus promoting formation of PGs is supported by the observation that 12-HETE is quantitatively the major AA metabolite formed in key intra-uterine tissues²⁵⁶ and that human labour is associated with an increased ratio of COX: LOX metabolites of AA²⁵⁷.

There were no apparent differences in myometrial expression of LOX genes comparing the upper and lower segment of the uterus. This is in contrast to COX genes which have been shown to demonstrate differential expression according to uterine site in the baboon⁸¹. These observations also tend to support an interpretation of different physiological roles for COX and LOX products in the control of parturition.

These data do not strongly suggest, at present, a potential role for LOX inhibitors in the treatment of preterm labour. Indeed, the observations that both advancing gestational age and labour are associated with decreases in the expression of some LOX genes in key tissues suggests that pharmacological inhibition of LOX genes may actually promote parturition by diverting free AA to the PG pathway. However, increased expression of 5-LOX in decidua is consistent with a possible role for 5-LOX products (such as leukotrienes) in parturition. Multiple receptors exist for these compounds and drugs with selective antagonist activity have been described²⁴⁷. Unlike enzyme inhibitors, receptor antagonists would not be expected to increase the pool of free AA and the effect of available drugs on parturition in animals warrants study.

Chapter 9. Overall conclusions.

The specific aims outlined in Chapter 1 were largely achieved. The immediate significance of observations to a given area of study were discussed in each chapter. The purpose of this chapter is, briefly, to draw together these observations into such general conclusions as can be drawn from this work.

9.1 Variation in prostanoid receptor expression with advancing gestational age and labour

As outlined in Chapter 1, there has been a great deal of study of the regulation of the genes which control PG concentrations in key intra-uterine tissues during parturition. One of the key aims of this thesis was to determine whether variation in the level of expression of the receptors may also have a role in the control of primate parturition. This aim was achieved and I demonstrated variation in the level of expression of genes comparing animals in labour and animals not in labour in myometrium, cervix, decidua and chorion. Of particular significance was the observation that expression of the prostanoid EP₂ receptor gene was lower in labour in myometrium (both lower segment and fundus [Chapter 4]), cervix (Chapter 5) and decidua (Chapter 6). This gene encodes an adenylate cyclase coupled receptor which typically inhibits myometrial contractility. I speculate that this gene may also exert inhibitory effects in the cervix and decidua and that loss of EP₂ receptor mediated inhibition in these tissues may be a key molecular event in primate parturition. I also demonstrated that in a key, PG mediated fetal adaptive response at birth, namely, contraction of the ductus arteriosus, one could demonstrate altered expression of stimulatory and inhibitory prostanoid receptor genes. These data suggest that variation in the expression of prostanoid receptor genes may be a key feature of dynamic physiological control of smooth muscle by PGs.

9.2 Regional variation in myometrial function

Many studies in pregnant woman have examined the function of myometrium obtained at the time of lower uterine segment Caesarean section ⁸⁶. The assumption (often unstated) of these studies was that the function of LUS myometrium is the same as myometrium from the corpus and fundus. In this thesis I demonstrated that this assumption is unsound. Variation exists in the expression of prostanoid receptor genes in tissue obtained prior to the onset of labour (Chapter 3) and the pattern of variation is consistent with functional variation in the response to native PGs ¹⁹⁵. Furthermore, the nature of the differences between the LUS and fundus is affected by labour in terms of inhibitory but not excitatory receptor genes (Chapter 4). These observations underline the potential pitfalls of studying LUS samples in isolation from the rest of the uterus. The studies also demonstrated that there was no regional variation in expression of LOX genes (Chapter 8) in contrast to studies of COX2 expression in these baboon tissues ⁸¹.

9.3 Tissue specific expression of prostanoid receptors

In the initial screen of expression of receptor genes, it was apparent that there was variation in the level of expression of different prostanoid receptor types and sub-types comparing different intra-uterine tissues (Chapter 3). When levels of expression were then compared in samples obtained at the time of labour and from animals prior to the onset of labour, it was observed that certain genes changed in some tissues but not others. For instance, there was a lower level of expression of the prostanoid EP₂ receptor gene in maternal intra-uterine tissues (Chapters 4-6) but not chorion (Chapter 6). In contrast, IP receptor gene expression was increased in both decidua and chorion (Chapter 6), but did not significantly differ in relation to labour in myometrium (Chapter 4) or cervix

(Chapter 5). These observations suggest that different PGs may have differential effects on different intra-uterine tissues and, consequently, that drugs with selective activity at prostanoid receptor types and sub-types may allow more specific manipulation of intra-uterine tissues.

9.4 Cell specific expression of prostanoid receptors

A number of lines of study in this thesis demonstrated that even in a given tissue a given second messenger system could not be characterized as either inhibitory or stimulatory. This possibility was underlined by findings in the decidua. Labour was associated with lower levels of expression of the EP₂ receptor gene but higher levels of expression of the IP receptor gene (Chapter 6). Both receptors are coupled to adenylate cyclase¹². This suggests that in cells which express the EP₂ receptor, cAMP dependent protein kinases have effects which inhibit the process of labour, whereas in cells expressing the IP receptor the same enzymes may have effects which promote parturition. The potential for these observations to have rational physiological explanations was apparent using *in situ* hybridization in myometrium which demonstrated differential expression of the prostanoid EP₃ receptor in myometrial and vascular smooth muscle (Chapter 4). This is likely to be due to the fact that uterine resistance vessels should be most widely dilated when uterine smooth muscle is most intensely activated.

9.5 The concept of stimulatory and inhibitory systems

The factors regulating myometrial contractility have been the natural focus of studies of the control of parturition. This reflects a key role for uterine contraction in the process of labour, but also probably also reflects the relative ease of study of myometrial function. Techniques have been available for more than 50 years that allow the study of the effects

of drugs on myometrial contractility. However, as discussed elsewhere in this thesis, other intra-uterine tissues have key roles in the process of parturition, such as the fetal membranes and cervix. However, it is much less straightforward to characterize the effect of a drug on the process of cervical priming or the integrity of the fetal membranes than uterine contraction. This has led both to the natural focus on myometrium and a tendency to characterize a given system as either promoting or inhibiting the process of parturition on the basis of its effect on myometrial contractility. It is clear, however, that the simple fact of a drug having an inhibitory effect on myometrial contractility does not ensure that the same drug will inhibit the processes leading to expulsion of the fetus in the other key intra-uterine tissues. A recent, important clinical example of this is nitric oxide, which inhibits myometrial contractility¹⁷¹, but which promotes the process of cervical ripening^{174;222}.

The current studies also demonstrated that receptors which would be anticipated to stimulate myometrial contractility were seen to have lower levels of mRNA expression in other tissues in association with labour. Both TP and FP receptor activation stimulate myometrial contractility⁸⁶, but expression of the genes encoding both receptors was lower in the cervix in association with labour (Chapter 5). Conversely, IP receptor activation inhibits myometrial contractility⁸⁶, but expression of the IP receptor gene was much higher in the chorion (and to a lesser extent in the decidua) obtained from animals in labour (Chapter 6). These observations suggest that key prostanoid receptor types might be stimulatory to the process of labour in some tissues but inhibit the process in others. These findings underline the necessity for caution when investigating a drug in the inhibition of labour on the basis of a negative effect on myometrial contractility.

9.6 Regulation of prostanoid receptor gene transcription

It is apparent from the studies reported in this thesis that there is variation in the expression of prostanoid receptor genes in association with advancing gestational age and labour which is manifested by altered levels of expression of the same receptor gene in some tissues but not others, or in a given tissue of some genes but not others. These findings suggest that factors modulate the control of expression of these genes in labour within key intra-uterine cells in a tissue specific manner. The global reduction in EP₂ receptor gene expression was taken to suggest a key role for this gene in the control of primate parturition. If true, the factors which control the onset of parturition may possibly be inferred by understanding the transcriptional regulation of this gene. There are very few data on the factors which control expression of any of the prostanoid receptors. For instance, one of the basic techniques in the study of factors which control gene transcription is DNA foot printing. A Medline search (September 2000) failed to yield any citations which used the MESH terms “DNA foot printing” and “receptors, prostaglandin”. This area, particularly as it relates to parturition, would appear to be ripe for future research.

9.7 Technical considerations relating to prostanoid gene expression

Over the course of the three year project, I clearly modified the technical approach to the study of prostanoid receptor genes. The major change was the use of riboprobes instead of cDNA probes for Northern blot analysis. In my initial studies I could not detect EP₁, IP or TP receptor signals using cDNA probes (Chapter 3). I then found that using riboprobes, appropriate sized transcripts could be detected for these genes (Chapter 4). There are a number of advantages to the use of riboprobes over cDNA probes. First, cDNA probes are double stranded whereas riboprobes are single stranded. The two

complementary DNA molecules will tend to bind each other and therefore not be available to bind the target. In the case of riboprobes, this is not an issue. Secondly, the higher stability of RNA: RNA duplexes compared with DNA:RNA duplexes ²⁵⁸ means that higher temperatures can be used for both hybridisation and washing which results in a higher signal to noise ratio. Thirdly, the unlabelled template DNA can be removed from the riboprobe using RNase free DNase I. This can clearly not be done using a cDNA probe. Finally, as part of the random priming process, one employs random decamers to create stretches of double stranded DNA to allow the Klenow DNA polymerase to synthesize DNA. Consequently, only a proportion of the given dNTP (for instance dCTP) is radioactively labelled since dCTP will also be included in the random hexamers, which are unlabelled. In contrast every NTP (UTP in the present study) in a riboprobe is radioactively labelled unless 'cold' UTP is added with the intention of making the probe less active.

Another issue is that the present study focused entirely on the expression of genes. I did not use other techniques to confirm that the RNA was translated into protein and that the protein was formed into functional receptors. The close parallel between gene expression and functional responses when comparing uterine regions ¹⁹⁵ (Chapter 3) supports my assumption that genes were translated. Ideally, one would employ protein analysis (such as immunocytochemistry or Western blot analysis) or receptor binding studies to confirm these assumptions. Unfortunately, antibodies are not widely available for the prostanoid receptor genes. Furthermore, such ligands as are available are not specific for a given prostanoid receptor type or sub-type ⁴⁷.

9.8 Summary

Many genes encoding prostanoid receptor types and sub-types are expressed in key intra-uterine tissues and the level of expression varies systematically within the gravid baboon uterus in relation to anatomical site, gestational age and labour. Comparable changes were demonstrated in the fetal ductus arteriosus. The nature of this variation suggests that altered expression of prostanoid receptor genes may have an important role in the control of parturition and in the preparation of the fetus for birth. Drugs with selective activity at prostanoid receptor types and sub-types are likely to allow safer and more effective control of the human uterus in the clinical management of term and pre-term labour.

.

References

1. Nathanielsz PW. Comparative studies on the initiation of labor.
Eur.J.Obstet.Gynecol.Reprod.Biol. 1998;**78**:127-32.
2. Sugimoto Y, Yamasaki A, Segi E, Tsuboi K, Aze Y, Nishimura T *et al.* Failure of parturition in mice lacking the prostaglandin F receptor. *Science* 1997;**277**:681-3.
3. Gross GA, Imamura T, Luedke C, Vogt SK, Olson LM, Nelson DM *et al.* Opposing actions of prostaglandins and oxytocin determine the onset of murine labor. *Proc Natl Acad Sci U.S.A* 1998;**95**:11875-9.
4. Mahendroo MS, Porter A, Russell DW, Word RA. The parturition defect in steroid 5alpha-reductase type 1 knockout mice is due to impaired cervical ripening. *Mol Endocrinol* 1999;**13**:981-92.
5. Challis JRG, Lye SJ, Gibb W. Prostaglandins and parturition. In Bulletti. C, ed. *Annals of the New York Academy of Sciences. The uterus: Endometrium and myometrium.*, pp 254-67. New York, New York, USA: New York Academy of Sciences, 1997.
6. Morgan MA, Silavin SL, Wentworth RA, Figueroa JP, Honnebier BO, Fishburne JIJ *et al.* Different patterns of myometrial activity and 24-h rhythms in myometrial contractility in the gravid baboon during the second half of pregnancy. *Biol Reprod* 1992;**46**:1158-64.
7. Shinozuka N, Yen A, Nathanielsz PW. Increased myometrial contracture frequency at 96-140 days accelerates fetal cardiovascular maturation. *Am.J Physiol Heart Circ.Physiol* 2000;**278**:H41-H49.

8. Caldeyro-Barcia R, Poseiro JJ. Physiology of the uterine contraction. *Clin Obstet Gynecol* 1960;**3**:386-408.
9. Drife J. Management of primary postpartum haemorrhage. *Br J Obstet Gynaecol* 1997;**104**:275-7.
10. Campbell WB, Halushka PV. Lipid derived autacoids: eicosanoids and platelet-activating factor. In Hardman JG, Limbird LE, Molinoff PB, Ruddon RW, Gilman AGG, eds. *Goodman & Gilman's The Pharmacological Basis of Therapeutics*, pp 601-16. New York: McGraw-Hill, 1996.
11. Coleman RA, Smith WL, Narumiya S. VIII. International Union of Pharmacology classification of prostanoid receptors: Properties, distribution, and structure of the receptors and their subtypes. *Pharmacol Rev* 1994;**46**:205-29.
12. Narumiya S, Sugimoto Y, Ushikubi F. Prostanoid receptors: structures, properties and functions. *Physiol Rev* 1999;**79**:1193-226.
13. Coleman RA, Kennedy I, Humphrey PPA, Bunce K, Lumley P. Prostanoids and their receptors. In Emmett. JC, ed. *Comprehensive Medicinal Chemistry: The Rational Design, Mechanistic Study and Therapeutic Application Of Chemical Compounds, Vol. 3. Membranes and Receptors*, pp 643-714. Oxford, England: Pergamon Press, 1990.
14. Smith WL, Marnett LJ, Dewitt DL. Prostaglandin and thromboxane biosynthesis. *Pharmacol Ther.* 1991;**49**:153-80.

15. Clyman RI, Mauray F, Roman C, Rudolph AM, Heymann MA. Circulating prostaglandin E2 concentrations and patent ductus arteriosus in fetal and neonatal lambs. *J Pediatr*. 1980;**97**:455-61.
16. Pierce KL, Regan JW. Prostanoid receptor heterogeneity through alternative mRNA splicing. *Life Sci* 1998;**62**:1479-83.
17. Harder DR, Campbell WB, Roman RJ. Role of cytochrome P-450 enzymes and metabolites of arachidonic acid in the control of vascular tone. *J Vasc Res* 1995;**32**:79-92.
18. Roberts LJ, Morrow JD. The generation and actions of isoprostanes. *Biochim Biophys Acta* 1997;**1345**:121-35.
19. Funk CD. Lipoxygenases of mice and men. In Folco GC, Samuelsson B, Macclouf J, Velo GP, eds. *Eicosanoids: from biotechnology to therapeutic applications*, pp 13-20. New York: Plenum Press, 1996.
20. Somlyo AP, Somlyo AV. Signal transduction and regulation in smooth muscle [published erratum appears in *Nature* 1994 Dec 22-29;372(6508):812]. *Nature* 1994;**372**:231-6.
21. Hartshorne DJ, Ito M, Erdodi F. Myosin light chain phosphatase: subunit composition, interactions and regulation. *J Muscle Res Cell Motil*. 1998;**19**:325-41.
22. Murakami M, Nakatani Y, Atsumi G, Inoue K, Kudo I. Regulatory functions of phospholipase A2. *Crit Rev Immunol*. 1997;**17**:225-83.

23. Vane JR, Bakhle YS, Botting RM. Cyclooxygenases 1 and 2. *Annu Rev Pharmacol Toxicol* 1998;**38**:97-120.
24. Habermehl DA, Janowiak MA, Vagnoni KE, Bird IM, Magness RR. Endothelial vasodilator production by uterine and systemic arteries. IV. Cyclooxygenase isoform expression during the ovarian cycle and pregnancy in sheep. *Biol Reprod* 2000;**62**:781-8.
25. Slater D, Dennes W, Sawdy R, Allport V, Bennett P. Expression of cyclooxygenase types-1 and -2 in human fetal membranes throughout pregnancy. *J Mol Endocrinol* 1999;**22**:125-30.
26. Lipsky PE, Brooks P, Crofford LJ, DuBois R, Graham D, Simon LS *et al.* Unresolved issues in the role of cyclooxygenase-2 in normal physiologic processes and disease. *Arch.Intern.Med* 2000;**160**:913-20.
27. Mitchell JA, Warner TD. Cyclo-oxygenase-2: pharmacology, physiology, biochemistry and relevance to NSAID therapy. *Br J Pharmacol* 1999;**128**:1121-32.
28. Langenbach R, Morham SG, Tiano HF, Loftin CD, Ghanayem BI, Chulada PC *et al.* Prostaglandin synthase 1 gene disruption in mice reduces arachidonic acid-induced inflammation and indomethacin-induced gastric ulceration. *Cell* 1995;**83**:483-92.
29. Morham SG, Langenbach R, Loftin CD, Tiano HF, Vouloumanos N, Jennette JC *et al.* Prostaglandin synthase 2 gene disruption causes severe renal pathology in the mouse. *Cell* 1995;**83**:473-82.

30. Davis BJ, Lennard DE, Lee CA, Tiano HF, Morham SG, Wetsel WC *et al.*
Anovulation in cyclooxygenase-2-deficient mice is restored by prostaglandin E2
and interleukin-1beta. *Endocrinology* 1999;**140**:2685-95.
31. Gardiner PJ. Characterization of prostanoid relaxant/inhibitory receptors (psi)
using a highly selective agonist, TR4979. *Br J Pharmacol* 1986;**87**:45-56.
32. Coleman RA, Grix SP, Head SA, Louttit JB, Mallett A, Sheldrick. A novel
inhibitory prostanoid receptor in piglet saphenous vein. *Prostaglandins*
1994;**47**:151-68.
33. Sturzebecher S, Hildebrand M, Schobel C, Seifert W, Staks T. Platelet inhibitory
and haemodynamic effects of a new stable PGI2 analogue, cicaprost (ZK
96480), in different animal species and in man. *Biomed.Biochim Acta*
1988;**47**:S45-S47.
34. Whittle BJ, Moncada S, Mullane K, Vane JR. Platelet and cardiovascular activity
of the hydantoin BW245C, a potent prostaglandin analogue. *Prostaglandins*
1983;**25**:205-23.
35. Wu KK, Le Breton GC, Tai HH, Chen YC. Abnormal platelet response to
thromboxane A2. *J Clin Invest* 1981;**67**:1801-4.
36. Rossdale PD, Pashen RL, Jeffcott LB. The use of synthetic prostaglandin
analogue (fluprostenol) to induce foaling. *J Reprod Fertil.Suppl* 1979;521-9.
37. Nials AT, Vardey CJ, Denyer LH, Thomas M, Sparrow SJ, Shepherd *et al.*
AH13205, a selective prostanoid EP-2-receptor agonist. *Cardiovasc Drug Rev*
1993; **11**:165-79.

38. Bunce KT, Clayton NM, Coleman RA, Collington EW, Finch H, Humphray JM *et al.* GR63799X--a novel prostanoid with selectivity for EP3 receptors. *Adv Prostaglandin Thromboxane Leukot Res* 1991;**21A**:379-82.
39. Smith GCS, Coleman RA, McGrath JC. Characterization of dilator prostanoid receptors in the fetal rabbit ductus arteriosus. *J Pharmacol Exp Ther* 1994;**271**:390-6.
40. Ritter JM, Benjamin N, Doktor HS, Barrow SE, Mant TG, Schey S *et al.* Effects of a selective thromboxane receptor antagonist (GR32191B) and of glyceryl trinitrate on bleeding time in man. *Br J Clin Pharmacol* 1990;**29**:431-6.
41. Giles H, Leff P, Bolofo ML, Kelly MG, Robertson AD. The classification of prostaglandin DP-receptors in platelets and vasculature using BW A868C, a novel, selective and potent competitive antagonist. *Br J Pharmacol* 1989;**96**:291-300.
42. Coleman RA, Kennedy I, Sheldrick RL. New evidence with selective agonists and antagonists for the subclassification of PGE2-sensitive (EP) receptors. *Adv Prostaglandin Thromboxane Leukot Res* 1987;**17A**:467-70.
43. Ito S, Okuda E, Sugama K, Negishi M, Hayaishi O. Evaluation of ZK110841 and AH6809, an agonist and an antagonist of prostaglandin DP-receptors on human platelets, with a PGD2-responsive cell line from bovine embryonic trachea. *Br J Pharmacol* 1990;**99**:13-4.
44. Bhattacharya M, Peri KG, Almazan G, Ribeiro-da-Silva A, Shichi H, Durocher Y *et al.* Nuclear localization of prostaglandin E2 receptors. *Proc Natl Acad Sci U.S.A* 1998;**95**:15792-7.

45. Lim H, Gupta RA, Ma WG, Paria BC, Moller DE, Morrow JD *et al.* Cyclo-oxygenase-2-derived prostacyclin mediates embryo implantation in the mouse via PPARdelta. *Genes Dev.* 1999;**13**:1561-74.
46. Ushikubi F, Hirata M, Narumiya S. Molecular biology of prostanoid receptors; an overview. *J Lipid Mediat.Cell Signal.* 1995;**12**:343-59.
47. Kiriyaama M, Ushikubi F, Kobayashi T, Hirata M, Sugimoto Y, Narumiya. Ligand binding specificities of the eight types and subtypes of the mouse prostanoid receptors expressed in Chinese hamster ovary cells. *Br J Pharmacol* 1997;**122**:217-24.
48. Matsuoka T, Hirata M, Tanaka H, Takahashi Y, Murata T, Kabashima K *et al.* Prostaglandin D2 as a mediator of allergic asthma. *Science* 2000;**287**:2013-7.
49. Hirata M, Kakizuka A, Aizawa M, Ushikubi F, Narumiya S. Molecular characterization of a mouse prostaglandin D receptor and functional expression of the cloned gene. *Proc Natl Acad Sci U.S.A* 1994;**91**:11192-6.
50. Watabe A, Sugimoto Y, Honda A, Irie A, Namba T, Negishi M *et al.* Cloning and expression of cDNA for a mouse EP1 subtype of prostaglandin E receptor. *J Biol Chem* 1993;**268**:20175-8.
51. Hizaki H, Segi E, Sugimoto Y, Hirose M, Saji T, Ushikubi F *et al.* Abortive expansion of the cumulus and impaired fertility in mice lacking the prostaglandin E receptor subtype EP(2). *Proc Natl Acad Sci U.S.A* 1999;**96**:10501-6.

52. Tilley SL, Audoly LP, Hicks EH, Kim HS, Flannery PJ, Coffman TM *et al.*
Reproductive failure and reduced blood pressure in mice lacking the EP2
prostaglandin E2 receptor [see comments]. *J Clin Invest* 1999;**103**:1539-45.
53. Katsuyama M, Nishigaki N, Sugimoto Y, Morimoto K, Negishi M, Narumiya S *et al.*
The mouse prostaglandin E receptor EP2 subtype: cloning, expression, and
northern blot analysis. *FEBS Lett.* 1995;**372**:151-6.
54. Takeuchi K, Ukawa H, Kato S, Furukawa O, Araki H, Sugimoto Y *et al.* Impaired
duodenal bicarbonate secretion and mucosal integrity in mice lacking
prostaglandin E-receptor subtype EP(3). *Gastroenterology* 1999;**117**:1128-35.
55. Sugimoto Y, Namba T, Honda A, Hayashi Y, Negishi M, Ichikawa A *et al.*
Cloning and expression of a cDNA for mouse prostaglandin E receptor EP3
subtype. *J Biol Chem* 1992;**267**:6463-6.
56. Nguyen M, Camenisch T, Snouwaert JN, Hicks E, Coffman TM, Anderson PA *et al.*
The prostaglandin receptor EP4 triggers remodelling of the cardiovascular
system at birth. *Nature* 1997;**390**:78-81.
57. Segi E, Sugimoto Y, Yamasaki A, Aze Y, Oida H, Nishimura T *et al.* Patent
ductus arteriosus and neonatal death in prostaglandin receptor EP4-deficient
mice. *Biochem.Biophys.Res.Commun.* 1998;**246**:7-12.
58. Nishigaki N, Negishi M, Honda A, Sugimoto Y, Namba T, Narumiya S *et al.*
Identification of prostaglandin E receptor 'EP2' cloned from mastocytoma cells
EP4 subtype. *FEBS Lett.* 1995;**364**:339-41.

59. Sugimoto Y, Yamasaki A, Segi E, Tsuboi K, Aze Y, Nishimura T *et al.* Failure of parturition in mice lacking the prostaglandin F receptor. *Science* 1997; **277**:681-3.
60. Sugimoto Y, Hasumoto K, Namba T, Irie A, Katsuyama M, Negishi M *et al.* Cloning and expression of a cDNA for mouse prostaglandin F receptor. *J Biol Chem* 1994; **269**:1356-60.
61. Murata T, Ushikubi F, Matsuoka T, Hirata M, Yamasaki A, Sugimoto Y *et al.* Altered pain perception and inflammatory response in mice lacking prostacyclin receptor. *Nature* 1997; **388**:678-82.
62. Namba T, Oida H, Sugimoto Y, Kakizuka A, Negishi M, Ichikawa A *et al.* cDNA cloning of a mouse prostacyclin receptor. Multiple signaling pathways and expression in thymic medulla. *J Biol Chem* 1994; **269**:9986-92.
63. Thomas DW, Mannon RB, Mannon PJ, Latour A, Oliver JA, Hoffman M *et al.* Coagulation defects and altered hemodynamic responses in mice lacking receptors for thromboxane A₂. *J Clin Invest* 1998; **102**:1994-2001.
64. Namba T, Sugimoto Y, Hirata M, Hayashi Y, Honda A, Watabe A *et al.* Mouse thromboxane A₂ receptor: cDNA cloning, expression and northern blot analysis. *Biochem Biophys Res Commun.* 1992; **184**:1197-203.
65. Namba T, Sugimoto Y, Negishi M, Irie A, Ushikubi F, Kakizuka A *et al.* Alternative splicing of C-terminal tail of prostaglandin E receptor subtype EP₃ determines G-protein specificity [see comments]. *Nature* 1993; **365**:166-70.

66. Negishi M, Sugimoto Y, Irie A, Narumiya S, Ichikawa A. Two isoforms of prostaglandin E receptor EP3 subtype. Different COOH-terminal domains determine sensitivity to agonist-induced desensitization. *J Biol Chem* 1993;**268**:9517-21.
67. Hirata M, Hayashi Y, Ushikubi F, Yokota Y, Kageyama R, Nakanishi S *et al.* Cloning and expression of complementary DNA for a human thromboxane A2 receptor. *Nature* 1991;**349**:617-20.
68. Boie Y, Rushmore TH, Darmon GA, Grygorczyk R, Slipetz DM, Metters KM *et al.* Cloning and expression of a cDNA for the human prostanoid IP receptor. *J Biol Chem* 1994;**269**:12173-8.
69. Armstrong RA, Lawrence RA, Jones RL, Wilson NH, Collier A. Functional and ligand binding studies suggest heterogeneity of platelet prostacyclin receptors. *Br.J Pharmacol.* 1989; **97**:657-68.
70. Armstrong RA, Humphrey PP, Lumley P. Characteristics of the binding of [3H]-GR32191 to the thromboxane (TP-) receptor of human platelets. *Br.J Pharmacol.* 1993;**110**:539-47.
71. Challis JR, Patel FA, Pomini F. Prostaglandin dehydrogenase and the initiation of labor. *J Perinat.Med* 1999;**27**:26-34.
72. Bygdeman M, Van Look PF. Anti-progesterones for the interruption of pregnancy. *Baillieres Clin Obstet Gynaecol* 1988;**2**:617-29.
73. Aiken JW. Aspirin and indomethacin prolong parturition in rats: evidence that prostaglandins contribute to expulsion of fetus. *Nature* 1972;**240**:21-5.

74. Lewis RB, Schulman JD. Influence of acetylsalicylic acid, an inhibitor of prostaglandin synthesis, on the duration of human gestation and labour. *Lancet* 1973;**2**:1159-61.
75. Wu WX, Unno N, Ma XH, Nathanielsz PW. Inhibition of prostaglandin production by nimesulide is accompanied by changes in expression of the cassette of uterine labor-related genes in pregnant sheep. *Endocrinol* 1998;**139**:3096-4003.
76. Unno N, Wu WX, Wong CH, Bennett PR, Shinozuka N, Nathanielsz PW. Prostaglandin regulation of fetal plasma adrenocorticotropin and cortisol concentrations in late-gestation sheep. *Biol Reprod* 1998;**58**:514-9.
77. Noort WA, van Bulck B, Vereecken A, de Zwart FA, Keirse MJ. Changes in plasma levels of PGF2 alpha and PGI2 metabolites at and after delivery at term. *Prostaglandins* 1989;**37**:3-12.
78. Keirse MJ, Mitchell MD, Turnbull AC. Changes in prostaglandin F and 13,14-dihydro-15-keto-prostaglandin F concentrations in amniotic fluid at the onset of and during labour. *Br J Obstet Gynaecol* 1977;**84**:743-6.
79. Romero R, Baumann P, Gonzalez R, Gomez R, Rittenhouse L, Behnke E *et al.* Amniotic fluid prostanoid concentrations increase early during the course of spontaneous labor at term. *Am.J Obstet Gynecol* 1994;**171**:1613-20.
80. Zhang Q, Wu WX, Brenna JT, Nathanielsz PW. The expression of cytosolic phospholipase A2 and prostaglandin endoperoxide synthase in ovine maternal uterine and fetal tissues during late gestation and labor. *Endocrinology* 1996;**137**:4010-7.

81. Wu WX, Ma XH, Smith GCS, Koenen SV, Nathanielsz PW. An integrated analysis of prostaglandin H synthase (PGHS) 2 mRNA in intrauterine tissues in late gestation and spontaneous labor in the pregnant baboon (abstract). *J Soc Gynecol Invest* 2000;**7 (supplement)**:52A-3A.
82. Romero R, Gonzalez R, Baumann P, Behnke E, Rittenhouse L, Barberio D *et al.* Topographic differences in amniotic fluid concentrations of prostanoids in women in spontaneous labor at term. *Prostaglandins Leukot Essent Fatty Acids* 1994;**50**:97-104.
83. Bocking AD. Preterm labour: recent advances in understanding of pathophysiology, diagnosis and management. *Curr. Opin. Obstet Gynecol* 1998;**10**:151-6.
84. Kniss DA, Jams JD. Regulation of parturition update. Endocrine and paracrine effectors of term and preterm labor. *Clin Perinatol.* 1998;**25**:819-36, v.
85. Thomson AJ, Telfer JF, Young A, Campbell S, Stewart CJ, Cameron IT *et al.* Leukocytes infiltrate the myometrium during human parturition: further evidence that labour is an inflammatory process. *Hum Reprod* 1999;**14**:229-36.
86. Senior J, Marshall K, Sangha R, Clayton JK. In-vitro characterization of prostanoid receptors on human myometrium at term pregnancy. *Br J Pharmacol* 1993;**108**:501-6.
87. Asboth G, Phaneuf S, Europe-Finner GN, Toth M, Lopez BA. Prostaglandin E-2 activates phospholipase C and elevates intracellular calcium in cultured myometrial cells: Involvement of EP1 and EP3 receptor subtypes. *Endocrinology.* 1996;**137**:2572-9.

88. Carrasco MP, Phaneuf S, Asboth G, Bernal AL. Fluprostenol activates phospholipase C and Ca-2+ mobilization in human myometrial cells. *Journal.of.Clinical.Endocrinology & Metabolism*. 1996;**81**:2104-10.
89. Garcia-Villar R, Green LR, Jenkins SL, Wentworth RA, Coleman RA, Nathanielsz PW. Evidence for the presence of AH13205-sensitive EP2 prostanoid receptors in the pregnant baboon but not in the pregnant sheep myometrium near term. *J Soc Gynecol Invest* 1995;**2**:6-12.
90. Crankshaw DJ, Gaspar V. Pharmacological characterization in vitro of prostanoid receptors in the myometrium of nonpregnant ewes. *J Reprod Fertil* 1995;**103**:55-61.
91. Brodt-Eppley J, Myatt L. Prostaglandin receptors in lower segment myometrium during gestation and labor. *Obstet Gynecol* 1999;**93**:89-93.
92. Brodt-Eppley J, Myatt L. Changes in expression of contractile FP and relaxatory EP2 receptors in pregnant rat myometrium during late gestation, at labor, and postpartum. *Biol Reprod* 1998;**59**:878-83.
93. Dong YL, Yallampalli C. Pregnancy and exogenous steroid treatments modulate the expression of relaxant EP(2) and contractile FP receptors in the rat uterus. *Biol Reprod* 2000;**62**:533-9.
94. Ou CW, Chen ZQ, Qi S, Lye SJ. Expression and regulation of the messenger ribonucleic acid encoding the prostaglandin F(2alpha) receptor in the rat myometrium during pregnancy and labor. *Am.J Obstet Gynecol* 2000;**182**:919-25.

95. Greer IA. Cervical ripening. In Drife JO, Calder AA, eds. *Prostaglandins and the uterus*, pp 191-209. London: Springer-Verlag, 1992.
96. Uldbjerg N, Ekman G, Malmstrom A, Sporrang B, Ulmsten U, Wingerup L. Biochemical and morphological changes of human cervix after local application of prostaglandin E2 in pregnancy [letter]. *Lancet* 1981;1:267-8.
97. French JI, McGregor JA. The pathobiology of premature rupture of membranes. *Semin Perinatol* 1996;**20**:344-68.
98. Heymann MA, Rudolph AM. Control of the ductus arteriosus. *Physiol Rev*. 1975;**55**:62-78.
99. Smith GCS. The pharmacology of the ductus arteriosus. *Pharmacol Rev* 1998;**50**:35-58.
100. Momma K, Takao A. Right ventricular concentric hypertrophy and left ventricular dilatation by ductal constriction in fetal rats. *Circ Res* 1989;**64**:1137-46.
101. Crichton CA, Smith GCS, Smith GL. alpha-Toxin-permeabilised rabbit fetal ductus arteriosus is more sensitive to Ca²⁺ than aorta or main pulmonary artery. *Cardiovasc Res* 1997;**33**:223-9.
102. Smith GCS, McGrath JC. Contractile effects of prostanoids on fetal rabbit ductus arteriosus. *J Cardiovasc Pharmacol* 1995;**25**:113-8.
103. Nguyen M, Camenisch T, Snouwaert JN, Hicks E, Coffman TM, Anderson PA *et al*. The prostaglandin receptor EP4 triggers remodelling of the cardiovascular system at birth. *Nature* 1997;**390**:78-81.

104. ZhuGe R, Li S, Chen TH, Hsu WH. Oxytocin induced a biphasic increase in the intracellular Ca^{2+} concentration of porcine myometrial cells: participation of a pertussis toxin-insensitive G-protein, inositol 1,4,5-trisphosphate-sensitive Ca^{2+} pool, and Ca^{2+} channels. *Mol Reprod Dev.* 1995;**41**:20-8.
105. Molnar M, Rigo J, Jr., Romero R, Hertelendy F. Oxytocin activates mitogen-activated protein kinase and up-regulates cyclooxygenase-2 and prostaglandin production in human myometrial cells. *Am.J Obstet Gynecol* 1999;**181**:42-9.
106. Chibbar R, Miller FD, Mitchell BF. Synthesis of oxytocin in amnion, chorion, and decidua may influence the timing of human parturition. *J Clin Invest* 1993;**91**:185-92.
107. Thornton S, Davison JM, Baylis PH. Plasma oxytocin during the first and second stages of spontaneous human labour. *Acta Endocrinol (Copenh.)* 1992;**126**:425-9.
108. Nishimori K, Young LJ, Guo Q, Wang Z, Insel TR, Matzuk MM. Oxytocin is required for nursing but is not essential for parturition or reproductive behavior. *Proc Natl Acad Sci U.S.A* 1996;**93**:11699-704.
109. Shubert PJ. Atosiban. *Clin Obstet Gynecol* 1995;**38**:722-4.
110. Goodwin TM, Valenzuela GJ, Silver H, Creasy G. Dose ranging study of the oxytocin antagonist atosiban in the treatment of preterm labor. Atosiban Study Group. *Obstet Gynecol* 1996;**88**:331-6.

111. Takemura M, Kimura T, Nomura S, Makino Y, Inoue T, Kikuchi T *et al.*
Expression and localization of human oxytocin receptor mRNA and its protein
in chorion and decidua during parturition. *J Clin Invest* 1994;**93**:2319-23.
112. Chibbar R, Wong S, Miller FD, Mitchell BF. Estrogen stimulates oxytocin gene
expression in human chorio-decidua. *J Clin Endocrinol Metab.* 1995;**80**:567-72.
113. Wu WX, Ma XH, Yoshizato T, Shinozuka N, Nathanielsz PW. Differential
expression of myometrial oxytocin receptor and prostaglandin H synthase 2, but
not estrogen receptor alpha and heat shock protein 90 messenger ribonucleic
acid in the gravid horn and nongravid horn in sheep during betamethasone-
induced labor. *Endocrinology* 1999;**140**:5712-8.
114. Fang X, Wong S, Mitchell BF. Effects of RU486 on estrogen, progesterone,
oxytocin, and their receptors in the rat uterus during late gestation.
Endocrinology. 1997;**138**:2763-8.
115. Ou CW, Chen ZQ, Qi S, Lye SJ. Increased expression of the rat myometrial
oxytocin receptor messenger ribonucleic acid during labor requires both
mechanical and hormonal signals. *Biol.Reprod.* 1998;**59**:1055-61.
116. Johnson CT. Induction of parturition in cattle. *Acta Vet.Scand.Suppl* 1981;**77**:311-
9.
117. Liggins GC, Fairclough RJ, Grieves SA, Forster CS, Knox BS. Parturition in the
sheep. *Ciba Found.Symp.* 1977;5-30.
118. McDonald TJ, Nathanielsz PW. Bilateral destruction of the fetal paraventricular
nuclei prolongs gestation in sheep. *Am.J Obstet Gynecol* 1991;**165**:764-70.

119. Anderson AB, Flint AP, Turnbull AC. Mechanism of action of glucocorticoids in induction of ovine parturition: effect on placental steroid metabolism. *J Endocrinol* 1975;**66**:61-70.
120. Challis JR. Sharp increase in free circulating oestrogens immediately before parturition in sheep. *Nature* 1971;**229**:208.
121. Rawlings NC, Howell WE. The use of estradiol benzoate to manage lambing period in ewes bred at synchronized estrus. *J Anim Sci* 1988;**66**:851-4.
122. Tomasi A, Tseng HY, Scommegna A, Burd L. The effect of medroxyprogesterone acetate on premature labor in the sheep. *Am.J Obstet Gynecol* 1985;**151**:694-700.
123. Wu WX, Ma XH, Zhang Q, Buchwalder L, Nathanielsz PW. Regulation of prostaglandin endoperoxide H synthase 1 and 2 by estradiol and progesterone in nonpregnant ovine myometrium and endometrium in vivo. *Endocrinology* 1997;**138**:4005-12.
124. Pepe GJ, Albrecht ED. Actions of placental and fetal adrenal steroid hormones in primate pregnancy. *Endocr.Rev.* 1995;**16**:608-48.
125. Walsh SW, Stanczyk FZ, Novy MJ. Daily hormonal changes in the maternal, fetal, and amniotic fluid compartments before parturition in a primate species. *J Clin Endocrinol Metab* 1984;**58**:629-39.
126. Walsh SW. Regulation of progesterone and estrogen production during rhesus monkey pregnancy. In Albrecht E, Pepe G, eds. *Perinatal Endocrinology*, pp 219-25. Ithaca, NY: Perinatology Press, 1985.

127. Haluska GJ, Cook MJ, Novy MJ. Inhibition and augmentation of progesterone production during pregnancy: effects on parturition in rhesus monkeys. *Am.J Obstet Gynecol* 1997;**176**:682-91.
128. Haluska GJ, Novy MJ. Hormonal modulation of uterine activity during primate parturition. *Semin Reprod Endocrinol* 1993;**11**:261-71.
129. Weiss G, Butler WR, Hotchkiss J, Dierschke DJ, Knobil E. Periparturitional serum concentrations of prolactin, the gonadotropins, and the gonadal hormones in the rhesus monkey. *Proc Soc Exp Biol Med* 1976;**151**:113-6.
130. Mesiano S, Jaffe RB. Developmental and functional biology of the primate fetal adrenal cortex. *Endocr.Rev.* 1997;**18**:378-403.
131. Mecnas CA, Giussani DA, Owiny JR, Jenkins SL, Wu WX, Honnebier BO *et al.* Production of premature delivery in pregnant rhesus monkeys by androstenedione infusion. *Nat Med* 1996;**2**:443-8.
132. Nathanielsz PW, Jenkins SL, Tame JD, Winter JA, Guller S, Giussani DA. Local paracrine effects of estradiol are central to parturition in the rhesus monkey. *Nat Med* 1998;**4**:456-9.
133. Horie K, Takakura K, Imai K, Liao S, Mori T. Immunohistochemical localization of androgen receptor in the human endometrium, decidua, placenta and pathological conditions of the endometrium. *Hum Reprod* 1992;**7**:1461-6.
134. Neulen J, Breckwoldt M. Placental progesterone, prostaglandins and mechanisms leading to initiation of parturition in the human. *Exp Clin Endocrinol* 1994;**102**:195-202.

135. Tulchinsky D, Hobel CJ, Yeager E, Marshall JR. Plasma estrone, estradiol, estriol, progesterone, and 17-hydroxyprogesterone in human pregnancy. I. Normal pregnancy. *Am.J Obstet Gynecol* 1972;**112**:1095-100.
136. Caritis SN, Darby MJ, Chan L. Pharmacologic treatment of preterm labor. *Clin Obstet Gynecol* 1988;**31**:635-51.
137. Mitchell BF, Rogers K, Wong S. The dynamics of prostaglandin metabolism in human fetal membranes and decidua around the time of parturition. *J Clin.Endocrinol.Metab* 1993;**77**:759-64.
138. Casey ML, MacDonald PC. Human parturition: distinction between the initiation of parturition and the onset of labor. *Semin Reprod Endocrinol* 1993;**11**:272-84.
139. Wu WX, Derks JB, Zhang Q, Nathanielsz PW. Changes in heat shock protein-90 and -70 messenger ribonucleic acid in uterine tissues of the ewe in relation to parturition and regulation by estradiol and progesterone. *Endocrinology* 1996;**137**:5685-93.
140. Peiber D, Allport VC, Slater DM, Bennett PR. Repression of the activity of the progesterone receptor B form by the A form may cause the functional progesterone withdrawal associated with human labor. *J Soc Gynecol Invest* 2000;**7** (supplement 1):51A.
141. Goodwin TM. A role for estriol in human labor, term and preterm. *Am.J Obstet Gynecol* 1999;**180**:S208-S213.
142. Jones CT, Boddy K, Robinson JS, Ratcliffe JG. Developmental changes in the responses of the adrenal glands of foetal sheep to endogenous

- adrenocorticotrophin, as indicated by hormone responses to hypoxaemia. *J Endocrinol* 1977;**72**:279-92.
143. Albrecht ED, Pepe GJ. Central integrative role of oestrogen in modulating the communication between the placenta and fetus that results in primate fecal-placental development. *Placenta* 1999;**20**:129-39.
144. Majzoub JA, McGregor JA, Lockwood CJ, Smith R, Taggart MS, Schulkin J. A central theory of preterm and term labor: putative role for corticotropin-releasing hormone. *Am.J Obstet Gynecol* 1999;**180**:S232-S241.
145. Creasy RK, Iams JD. Preterm labor and delivery. In Creasy RK, Resnik R, eds. *Maternal-Fetal Medicine*, pp 498-531. Philadelphia: WB Saunders, 1999.
146. Novy MJ, Walsh SW. Dexamethasone and estradiol treatment in pregnant rhesus macaques: effects on gestational length, maternal plasma hormones, and fetal growth. *Am.J Obstet Gynecol* 1983;**145**:920-31.
147. Korebrits C, Yu DH, Ramirez MM, Marinoni E, Bocking AD, Challis JR. Antenatal glucocorticoid administration increases corticotrophin-releasing hormone in maternal plasma. *Br J Obstet Gynaecol* 1998;**105**:556-61.
148. Patel FA, Clifton VL, Chwalisz K, Challis JR. Steroid regulation of prostaglandin dehydrogenase activity and expression in human term placenta and chorion-decidua in relation to labor. *J Clin Endocrinol Metab* 1999;**84**:291-9.
149. Pinto RM, Leon C, Mazzocco N, Scasserra V. Action of estradiol-17-beta at term and at onset of labor. *Am.J Obstet Gynecol* 1967;**98**:540-6.

150. Vessey MP. Epidemiological studies of the effects of diethylstilboestrol. *IARC Sci Publ.* 1989;335-48.
151. Allen J, Uldbjerg N, Petersen LK, Secher NJ. Intracervical 17 beta-oestradiol before induction of second-trimester abortion with a prostaglandin E1 analogue. *Eur.J Obstet Gynecol Reprod Biol* 1989;32:123-7.
152. Magann EF, Perry KGJ, Dockery JRJ, Bass JD, Chauhan SP, Morrison JC. Cervical ripening before medical induction of labor: a comparison of prostaglandin E2, estradiol, and oxytocin. *Am J Obstet Gynecol* 1995;172:1702-6.
153. Riley SC, Challis JR. Corticotrophin-releasing hormone production by the placenta and fetal membranes. *Placenta* 1991;12:105-19.
154. Dibbs KI, Anteby E, Mallon MA, Sadovsky Y, Adler S. Transcriptional regulation of human placental corticotropin-releasing factor by prostaglandins and estradiol. *Biol Reprod* 1997;57:1285-92.
155. Majzoub JA, Karalis KP. Placental corticotropin-releasing hormone: function and regulation. *Am.J Obstet Gynecol* 1999;180:S242-S246.
156. Simpkin JC, Kermani F, Palmer AM, Campa JS, Tribe RM, Linton EA *et al.* Effects of corticotrophin releasing hormone on contractile activity of myometrium from pregnant women. *Br J Obstet Gynaecol* 1999;106:439-45.
157. Stevens MY, Challis JR, Lye SJ. Corticotropin-releasing hormone receptor subtype 1 is significantly up-regulated at the time of labor in the human myometrium. *J.Clin.Endocrinol.Metab* 1998;83:4107-15.

158. Brown NL, Slater DM, Alvi SA, Elder MG, Sullivan MH, Bennett PR.
Expression of 5-lipoxygenase and 5-lipoxygenase-activating protein in human fetal membranes throughout pregnancy and at term. *Mol Hum Reprod* 1999;**5**:668-74.
159. Bennett PR, Elder MG, Myatt L. The effects of lipoxygenase metabolites of arachidonic acid on human myometrial contractility. *Prostaglandins* 1987;**33**:837-44.
160. Morrow JD, Minton TA, Roberts LJ. The F2-isoprostane, 8-epi-prostaglandin F2 alpha, a potent agonist of the vascular thromboxane/endoperoxide receptor, is a platelet thromboxane/endoperoxide receptor antagonist. *Prostaglandins* 1992;**44**:155-63.
161. Kayganich-Harrison KA, Rose DM, Murphy RC, Morrow JD, Roberts LJ.
Collision-induced dissociation of F2-isoprostane-containing phospholipids. *J Lipid Res* 1993;**34**:1229-35.
162. Crankshaw D. Effects of the isoprostane, 8-epi-prostaglandin F-2alpha, on the contractility of the human myometrium in vitro.
European.Journal.of.Pharmacology. 1995;**285**:151-8.
163. Pennefather JN, Paull JD, Story ME, Ziccone SP. Supersensitivity to the stimulant action of noradrenaline on human myometrium near term. *Reprod Fertil Dev.* 1993;**5**:39-48.
164. Martinez-Mir MI, Estan L, Morales-Olivas FJ, Rubio E. Effect of histamine and histamine analogues on human isolated myometrial strips. *Br J Pharmacol* 1992;**107**:528-31.

165. Rudolph MI, Reinicke K, Cruz MA, Gallardo V, Gonzalez C, Bardisa L.
Distribution of mast cells and the effect of their mediators on contractility in human myometrium. *Br.J Obstet.Gynaecol.* 1993;**100**:1125-30.
166. Pitera AE, Smith GC, Wentworth RA, Nathanielsz PW. Parathyroid hormone-related peptide (1 to 34) inhibits in vitro oxytocin-stimulated activity of pregnant baboon myometrium. *Am J Obstet Gynecol* 1998;**179**:492-6.
167. Ferguson JE, Gorman JV, Bruns DE, Weir EC, Burtis WJ, Martin TJ *et al.*
Abundant expression of parathyroid hormone-related protein in human amnion and its association with labor. *Proc Natl Acad Sci U.S.A* 1992;**89**:8384-8.
168. Bani D. Relaxin: a pleiotropic hormone. *Gen.Pharmacol* 1997;**28**:13-22.
169. Brennand JE, Calder AA, Leitch CR, Greer IA, Chou MM, MacKenzie IZ.
Recombinant human relaxin as a cervical ripening agent. *Br J Obstet Gynaecol* 1997;**104**:775-80.
170. Millar LK, Boesche MH, Yamamoto SY, Killeen J, DeBuque L, Chen R *et al.* A relaxin-mediated pathway to preterm premature rupture of the fetal membranes that is independent of infection. *Am.J Obstet Gynecol* 1998;**179**:126-34.
171. Norman JE, Ward LM, Martin W, Cameron AD, McGrath JC, Greer IA *et al.*
Effects of cGMP and the nitric oxide donors glyceryl trinitrate and sodium nitroprusside on contractions in vitro of isolated myometrial tissue from pregnant women. *J Reprod Fertil* 1997;**110**:249-54.

172. Lees C, Campbell S, Jauniaux E, Brown R, Ramsay B, Gibb D *et al.* Arrest of preterm labour and prolongation of gestation with glyceryl trinitrate, a nitric oxide donor. *Lancet* 1994;**343**:1325-6.
173. Ali M, Buhimschi I, Chwalisz K, Garfield RE. Changes in expression of the nitric oxide synthase isoforms in rat uterus and cervix during pregnancy and parturition. *Mol.Hum.Reprod.* 1997;**3**:995-1003.
174. Thomson AJ, Lunan CB, Cameron AD, Cameron IT, Greer IA, Norman JE. Nitric oxide donors induce ripening of the human uterine cervix: a randomised controlled trial. *Br J Obstet Gynaecol* 1997;**104**:1054-7.
175. Wray S. Uterine contraction and physiological mechanisms of modulation. *Am J Physiol.* 1993;**264**:C1-18.
176. Somlyo AP, Wu X, Walker LA, Somlyo AV. Pharmacomechanical coupling: the role of calcium, G-proteins, kinases and phosphatases. *Rev.Physiol Biochem Pharmacol* 1999;**134**:201-34.
177. Luckas MJ, Taggart MJ, Wray S. Intracellular calcium stores and agonist-induced contractions in isolated human myometrium. *Am.J Obstet Gynecol* 1999;**181**:468-76.
178. Karaki H, Ozaki H, Hori M, Mitsui-Saito M, Amano K, Harada K *et al.* Calcium movements, distribution, and functions in smooth muscle. *Pharmacol Rev* 1997;**49**:157-230.
179. Garfield RE, Ali M, Yallampalli C, Izumi H. Role of gap junctions and nitric oxide in control of myometrial contractility. *Semin.Perinatol.* 1995;**19**:41-51.

180. Sanborn BM, Yue C, Wang W, Dodge KL. G protein signalling pathways in myometrium: affecting the balance between contraction and relaxation. *Rev Reprod* 1998;**3**:196-205.
181. Namba T, Sugimoto Y, Negishi M, Irie A, Ushikubi F, Kakizuka A *et al.* Alternative splicing of C-terminal tail of prostaglandin E receptor subtype EP3 determines G-protein specificity [see comments]. *Nature* 1993;**365**:166-70.
182. Europe-Finner GN, Phaneuf S, Tolkovsky AM, Watson SP, Lopez BA. Down-regulation of G alpha s in human myometrium in term and preterm labor: a mechanism for parturition. *J Clin Endocrinol Metab* 1994;**79**:1835-9.
183. McKillen K, Thornton S, Taylor CW. Oxytocin increases the $[Ca^{2+}]_i$ sensitivity of human myometrium during the falling phase of phasic contractions. *Am.J.Physiol.* 1999;**276**:E345-E351.
184. Izumi H, Ichihara J, Uchiumi Y, Shirakawa K. Gestational changes in mechanical properties of skinned muscle tissues of human myometrium. *Am J Obstet Gynecol* 1990;**163**:638-47.
185. Word RA, Stull JT, Casey ML, Kamm KE. Contractile elements and myosin light chain phosphorylation in myometrial tissue from nonpregnant and pregnant women. *J Clin Invest* 1993; **92**:29-37.
186. Gyetvai K, Hannah ME, Hodnett ED, Ohlsson A. Tocolytics for preterm labor: a systematic review. *Obstet Gynecol* 1999;**94**:869-77.
187. Hannah ME, Hannah WJ, Hellmann J, Hewson S, Milner R, Willan A. Induction of labor as compared with serial antenatal monitoring in post-term pregnancy. A

- randomized controlled trial. The Canadian Multicenter Post-term Pregnancy Trial Group. *N Engl J Med* 1992;**326**:1587-92.
188. Grant JM. Induction of labour confers benefits in prolonged pregnancy. *Br J Obstet Gynaecol* 1994;**101**:99-102.
189. Saunders N, Paterson C. Effect of gestational age on obstetric performance: when is "term" over? *Lancet* 1991;**338**:1190-2.
190. Yudkin PL, Wood L, Redman CW. Risk of unexplained stillbirth at different gestational ages. *Lancet* 1987;**1**:1192-4.
191. Feinberg AP, Vogelstein B. A technique for radiolabelling DNA restriction endonuclease fragments to a high specific activity. *Analyt Biochem* 1983;**132**:6-13.
192. Casey ML, MacDonald PC, Mitchell MD, Snyder JM. Maintenance and characterization of human myometrial smooth muscle cells in monolayer culture. *In Vitro* 1984;**20**:396-403.
193. Fuchs AR, Fuchs F, Husslein P, Soloff M. Oxytocin receptors in the human uterus during pregnancy and parturition. *Am J Obstet Gynecol* 1984;**150**:734-41.
194. Wikland M, Lindblom B, Wijkvist N. Myometrial response to prostaglandins during labor. *Gynecol Obstet Invest* 1984;**17**:131-8.
195. Smith GCS, Baguma-Nibasheka M, Wu WX, Nathanielsz PW. Regional variations in contractile responses to prostaglandins and prostanoid receptor messenger ribonucleic acid in pregnant baboon uterus. *Am J Obstet Gynecol* 1998;**179**:1545-52.

196. Gould SF. Anatomy. In Gabbe SG, Niebyl JR, Simpson JL, eds. *Obstetrics: normal and problem pregnancies*, pp 3-38. New York: Churchill Livingstone, 1991.
197. Kotani M, Tanaka I, Ogawa Y, Usui T, Mori K, Ichikawa A *et al*. Molecular cloning and expression of multiple isoforms of human prostaglandin E receptor EP-3 subtype generated by alternative messenger RNA splicing: Multiple second messenger systems and tissue-specific distributions. *Mol Pharmacol* 1995;**48**:869-79.
198. Regan JW, Bailey TJ, Pepperl DJ, Pierce KL, Bogardus AM, Donello JE *et al*. Cloning of a novel human prostaglandin receptor with characteristics of the pharmacologically defined EP-2 subtype. *Mol Pharmacol* 1994;**46**:213-20.
199. An S, Yang J, Xia M, Goetzl EJ. Cloning and expression of the EP2 subtype of human receptors for prostaglandin E2. *Biochem Biophys Res Comm* 1993;**197**:263-70.
200. Lake S, Gullberg H, Wahlqvist J, Sjogren AM, Kinhult A, Lind P *et al*. Cloning of the rat and human prostaglandin F2 alpha receptors and the expression of the rat prostaglandin F2 alpha receptor. *FEBS Lett*. 1994;**355**:317-25.
201. Bastien L, Sawyer N, Grygorczyk R, Metters KM, Adam M. Cloning, functional expression, and characterization of the human prostaglandin E2 receptor EP2 subtype. *J Biol Chem* 1994;**269**:11873-7.
202. Mijovic JE, Zakar T, Nairn TK, Olson DM. Prostaglandin-endoperoxide H synthase-2 expression and activity increases with term labor in human chorion. *Am J Physiol* 1997;**272**:E832-E840.

203. Enkin M, Keirse MJNC, Renfrew M, Neilson JP. A guide to effective care in pregnancy and childbirth. Oxford: Oxford University Press, 1997.
204. Rayburn WF. Clinical experience with a controlled-release prostaglandin E2 intravaginal insert in the USA. *Br J Obstet Gynaecol* 1997;**104**:S8-S12.
205. Carbonne B, Jannet D, Dallot E, Pannier E, Ferre F, Cabrol D. Synthesis of glycosaminoglycans by human cervical fibroblasts in culture: Effects of prostaglandin E-2 and cyclic AMP. *Eur J Obstet Gynecol Reprod Biol* 1996;**70**:101-5.
206. Surbek DV, Boesiger H, Hoesli I, Pavic N, Holzgreve W. A double-blind comparison of the safety and efficacy of intravaginal misoprostol and prostaglandin E2 to induce labor. *Am J Obstet Gynecol* 1997;**177**:1018-23.
207. Gottschall DS, Borgida AF, Mihalek JJ, Sauer F, Rodis JF. A randomized clinical trial comparing misoprostol with prostaglandin E2 gel for preinduction cervical ripening. *Am J Obstet Gynecol* 1997;**177**:1067-70.
208. Fruzzetti F, Melis GB, Strigini F, Vettori C, Ricci C, Fioretti P. Use of sulprostone for induction of preoperative cervical dilation or uterine evacuation a comparison among the effects of different treatment schedules. *Obstet Gynecol* 1988;**72** :704-8.
209. Oleen MA, Mariano JP. Controlling refractory atonic postpartum hemorrhage with hemabate sterile solution. *Am J Obstet Gynecol* 1990;**162**:205-8.
210. Van den Veyver IB, Moise KJ. Prostaglandin synthetase inhibitors in pregnancy. *Obstet Gynecol Surv* 1993;**48**:493-502.

211. Keirse MJNC, van Oppen CC. Preparing the cervix for induction of labour. In Chalmers I, Enkin M, Keirse MJNC, eds. *Effective care in pregnancy and childbirth*, pp 988-1056. Oxford, UK: OUP, 1989.
212. Skinner KA, Challis JR. Changes in the synthesis and metabolism of prostaglandins by human fetal membranes and decidua at labor. *Am J Obstet Gynecol* 1985;**151**:519-23.
213. Phaneuf S, Asboth G, Europe-Finner GN, Watson SP, Lopez BA. Second messenger pathways for oxytocin and prostaglandins in human myometrium. *Biochem Soc Trans* 1995;**23**:21S.
214. Ou CW, Orsino A, Lye SJ. Expression of connexin-43 and connexin-26 in the rat myometrium during pregnancy and labor is differentially regulated by mechanical and hormonal signals. *Endocrinol* 1997;**138**:5398-407.
215. Ma XH, Wu WX, Nathanielsz PW. Differential regulation of prostaglandin EP and FP receptors in the pregnant sheep myometrium and endometrium during spontaneous term labor. *Biol Reprod* 1999;**61**:1281-6.
216. Keirse MJNC, Chalmers I. Methods for inducing labour. In Chalmers I, Enkin M, Keirse MJNC, eds. *Effective care in pregnancy and childbirth*, pp 1057-79. Oxford, UK: OUP, 1989.
217. Abramovitz M, Boie Y, Nguyen T, Rushmore TH, Bayne MA, Metters K *et al*. Cloning and expression of a cDNA for the human prostanoid FP receptor. *J Biol Chem* 1994;**269**:2632-6.

218. Challis JR, Lye SJ, Gibb W. Prostaglandins and parturition. *Ann.N Y.Acad.Sci* 1997;**828**:254-67.
219. Van Meir CA, Ramirez MM, Matthews SG, Calder AA, Keirse MJ, Challis JR. Chorionic prostaglandin catabolism is decreased in the lower uterine segment with term labour. *Placenta* 1997;**18**:109-14.
220. Liu YL, Nwosu UC, Rice PJ. Relaxation of isolated human myometrial muscle by beta2-adrenergic receptors but not beta1-adrenergic receptors. *Am.J.Obstet.Gynecol.* 1998;**179**:895-8.
221. Hoffman BB, Lefkowitz RJ. Adrenergic receptors in the heart. *Annu.Rev.Physiol* 1982;**44**:475-84.
222. Chwalisz K, Garfield RE. Nitric oxide as the final metabolic mediator of cervical ripening. *Hum.Reprod.* 1998;**13**:245-8.
223. Sawdy R, Slater D, Fisk N, Edmonds DK, Bennett P. Use of a cyclo-oxygenase type-2-selective non-steroidal anti-inflammatory agent to prevent preterm delivery [letter]. *Lancet* 1997;**350**:265-6.
224. Thomson AJ, Lunan CB, Ledingham M, Howat RC, Cameron IT, Greer IA *et al.* Randomised trial of nitric oxide donor versus prostaglandin for cervical ripening before first-trimester termination of pregnancy. *Lancet* 1998;**352**:1093-6.
225. Smith GCS, Wu WX, Nathanielsz PW. Baboon labor is associated with decreased uterine prostanoid EP2 receptor expression. *J Soc Gynecol Invest* 1999;**6**:128A-9A.

226. Yang ZM, Das SK, Wang J, Sugimoto Y, Ichikawa A, Dey SK. Potential sites of prostaglandin actions in the periimplantation mouse uterus: Differential expression and regulation of prostaglandin receptor genes. *Biol Reprod* 1997;**56**:368-79.
227. Lim H, Paria BC, Das SK, Dinchuk JE, Langenbach R, Trzaskos JM *et al.* Multiple female reproductive failures in cyclooxygenase 2-deficient mice. *Cell* 1997;**91**:197-208.
228. Spaziani EP, O'Brien WF, Benoit RR, Gould SF. Corticotropin-releasing hormone increases the expression of the prostaglandin E(2) receptor subtype EP1 in amnion WISH cells. *Biol Reprod* 2000;**62**:23-6.
229. Spaziani EP, O'Brien WF, Tsibris JC, Benoit RR, Gould SF. Modulation of the prostaglandin E receptor: a possible mechanism for infection-induced preterm labor. *Obstet Gynecol* 1999;**93**:84-8.
230. Spaziani EP, Tsibris JC, Hunt LT, Benoit RR, O'Brien WF. The effect of interleukin-1 beta and interleukin-4 on the expression of prostaglandin receptors EP1 and EP3 in amnion WISH cells. *Am J Reprod.Immunol.* 1997;**38**:279-85.
231. Spaziani EP, Benoit RR, Tsibris JC, Gould SF, O'Brien WF. Tumor necrosis factor-alpha upregulates the prostaglandin E2 EP1 receptor subtype and the cyclooxygenase-2 isoform in cultured amnion WISH cells. *J Interferon Cytokine Res* 1998;**18**:1039-44.
232. Nakanishi T, Gu H, Hagiwara N, Momma K. Mechanisms of oxygen-induced contraction of ductus arteriosus isolated from the fetal rabbit. *Circ Res* 1993;**72**:1218-28.

233. Tristani-Firouzi M, Reeve HL, Tolarova S, Weir EK, Archer SL. Oxygen-induced constriction of rabbit ductus arteriosus occurs via inhibition of a 4-aminopyridine-, voltage-sensitive potassium channel. *J Clin Invest* 1996;**98**:1959-65.
234. Clyman RI, Mauray F, Roman C, Heymann MA, Ballard PL, Rudolph AM *et al.* Effects of antenatal glucocorticoid administration on ductus arteriosus of preterm lambs. *Am J Physiol* 1981;**241**:H415-H420.
235. Clyman RI, Mauray F, Rudolph AM, Heymann MA. Age-dependent sensitivity of the lamb ductus arteriosus to indomethacin and prostaglandins. *J Pediatr* 1980;**96**:94-8.
236. Clyman RI, Mauray F, Roman C, Heymann MA, Payne B. Factors determining the loss of ductus arteriosus responsiveness to prostaglandin E. *Circulation* 1983;**68**:433-6.
237. Silove ED, Coe JY, Shiu MF, Brunt JD, Page AJ, Singh SP *et al.* Oral prostaglandin E2 in ductus-dependent pulmonary circulation. *Circulation* 1981;**63**:682-8.
238. Marshall FK, Patel K, Lundstrom K, Camacho J, Foord SM, Lee MG. Characterization of 3H-prostaglandin E2 binding to prostaglandin EP4 receptors expressed with Semliki Forest virus. *Br J Pharmacol* 1997;**121**:1673-8.
239. Norton ME, Merrill J, Cooper BAB, Kuller JA, Clyman RI. Neonatal complications after the administration of indomethacin for preterm labor. *N Engl J Med* 1993;**329**:1602-7.

240. Jobe AH. Fetal lung development, tests for maturation, induction of maturation and treatment. In Creasy RK, Resnik R, eds. *Maternal-Fetal Medicine*, pp 404-22. Philadelphia: WB Saunders, 1999.
241. Clyman RI, Mauray F, Roman C, Rudolph AM, Heymann MA. Glucocorticoids alter the sensitivity of the lamb ductus arteriosus to prostaglandin E2. *J Pediatr*. 1981;**98**:126-8.
242. Takizawa T, Arishima K, Yamamoto M, Shirai M, Somiya H, Eguchi Y. Constrictive effect of thyroxine on the ductus arteriosus in fetal rats. *J Vet Med Sci* 1993;**55**:379-81.
243. Mecenas CA, Jenkins SL, Smith GCS, Loris JLN PW. 48 hour maternally administered betamethasone increases maternal and fetal arterial blood pressure and heart rate at 0.7 gestation in the baboon. *J Soc Gynecol Invest* 1999;**6** (Supplement 1):184A (Abstract).
244. Bhattacharya M, Asselin P, Hardy P, Guerguerian AM, Shichi H, Hou X *et al*. Developmental changes in prostaglandin E(2) receptor subtypes in porcine ductus arteriosus. Possible contribution in altered responsiveness to prostaglandin E(2). *Circulation* 1999;**100**:1751-6.
245. Wu WX, Ma XH, Nathanielsz PW. Tissue-specific ontogenic expression of prostaglandin H synthase 2 in the ovine myometrium, endometrium, and placenta during late gestation and at spontaneous term labor. *Am J Obstet Gynecol* 1999;**181**:1512-9.

246. Conrad DJ, Kuhn H, Mulkins M, Highland E, Sigal E. Specific inflammatory cytokines regulate the expression of human monocyte 15-lipoxygenase. *Proc Natl Acad Sci USA* 1992;**89**:217-21.
247. Coleman RA, Eglen RM, Jones RL, Narumiya S, Shimizu T, Smith WL *et al*. Prostanoid and leukotriene receptors: A progress report from the IUPHAR working parties on classification and nomenclature. *Adv Prost Thrombox Leuk Res* 1995;**23**:283-5.
248. Drazen JM, Israel E, O'Byrne PM. Treatment of asthma with drugs modifying the leukotriene pathway. *N Engl J Med* 1999;**340**:197-206.
249. Matsumoto T, Funk CD, Radmark O, Hoog JO, Jornvall H, Samuelsson B. Molecular cloning and amino acid sequence of human 5-lipoxygenase. *Proc Natl Acad Sci USA* 1988;**85**:26-30.
250. Funk CD, Funk LB, Fitzgerald GA, Samuelsson B. Characterization of human 12-lipoxygenase genes. *Proc.Natl.Acad.Sci.U.S.A* 1992;**89**:3962-6.
251. Bryman I, Hammarstrom S, Lindblom B, Norstrom A, Wikland M, Wiqvist N. Leukotrienes and myometrial activity of the term pregnant uterus. *Prostaglandins* 1985;**30**:907-13.
252. Ticconi C, Zicari A, Pontieri G, Salerno A, Mauri A, Piccione E *et al*. Release of arachidonic acid metabolites by human fetal membranes: interrelationship between leukotriene B4 and prostaglandin E2. *Prostaglandins* 1995;**49**:197-204.

253. Romero R, Wu YK, Mazor M, Hobbins JC, Mitchell MD. Increased amniotic fluid leukotriene C4 concentration in term human parturition. *Am J Obstet Gynecol* 1988;**159**:655-7.
254. Romero R, Wu YK, Mazor M, Hobbins JC, Mitchell MD. Amniotic fluid concentration of 5-hydroxyeicosatetraenoic acid is increased in human parturition at term. *Prost Leukot Essent Fatty Acids* 1989;**35**:81-3.
255. Saeed SA, Mitchell MD. Lipoxygenase activity in human uterine and intrauterine tissues: new prospects for control of prostacyclin production in pre-eclampsia. *Clin Exp Hypertens [B]* 1983;**2**:103-11.
256. Schafer WR, Zahradnik HP, Arbogast E, Wetzka B, Werner K, Breckwoldt M. Arachidonate metabolism in human placenta, fetal membranes, decidua and myometrium: lipoxygenase and cytochrome P450 metabolites as main products in HPLC profiles. *Placenta* 1996;**17**:231-8.
257. Bennett PR, Slater D, Sullivan M, Elder MG, Moore GE. Changes in amniotic arachidonic acid metabolism associated with increased cyclo-oxygenase gene expression. *Br J Obstet Gynaecol* 1993;**100**:1037-42.
258. Casey J, Davidson N. Rates of formation and thermal stabilities of RNA:DNA and DNA:DNA duplexes at high concentrations of formamide. *Nucl Acid Res* 1977;**4**:1539-52.